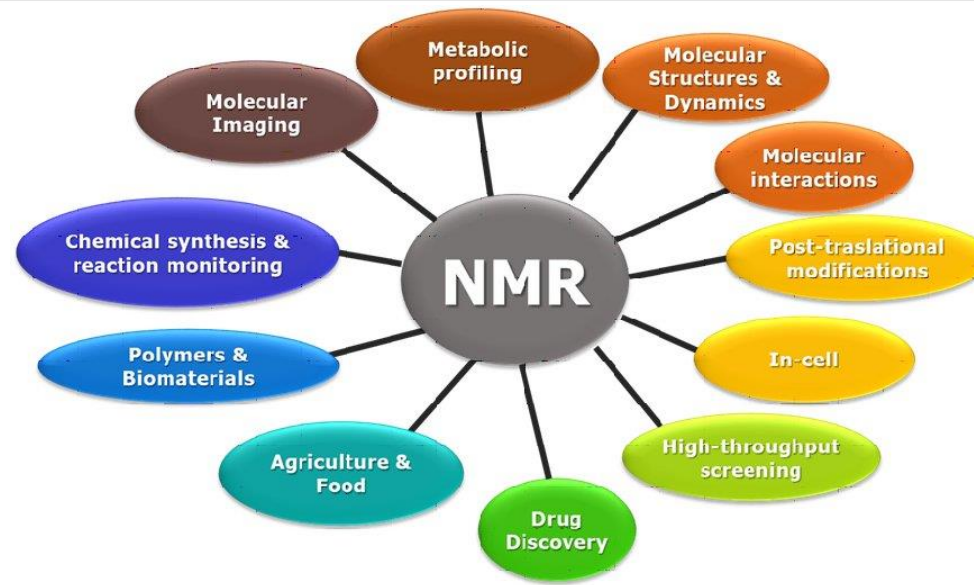


# Applications

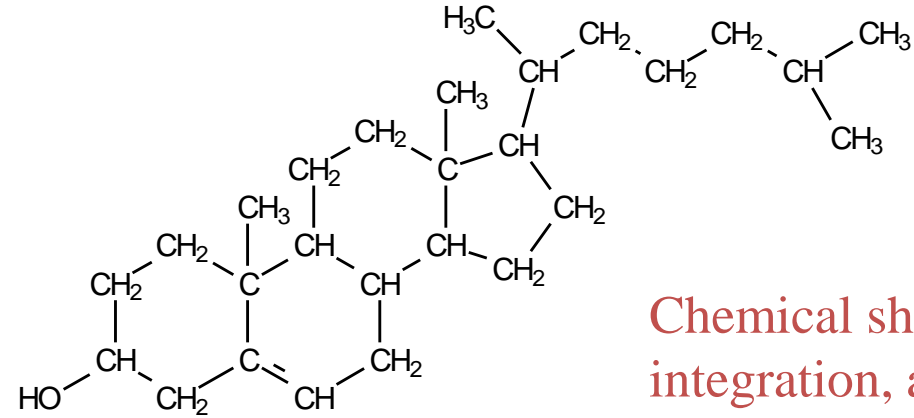
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# Chemistry

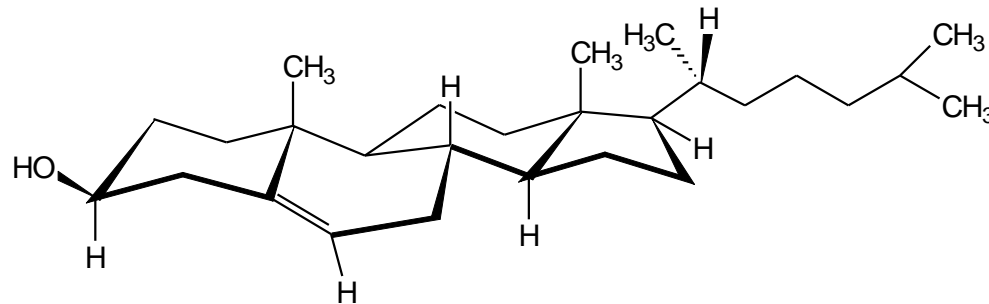
## Chemical structure

$^1\text{H}$ : 46 hydrogen atoms  
 $^{13}\text{C}$ : 27 carbon atoms.



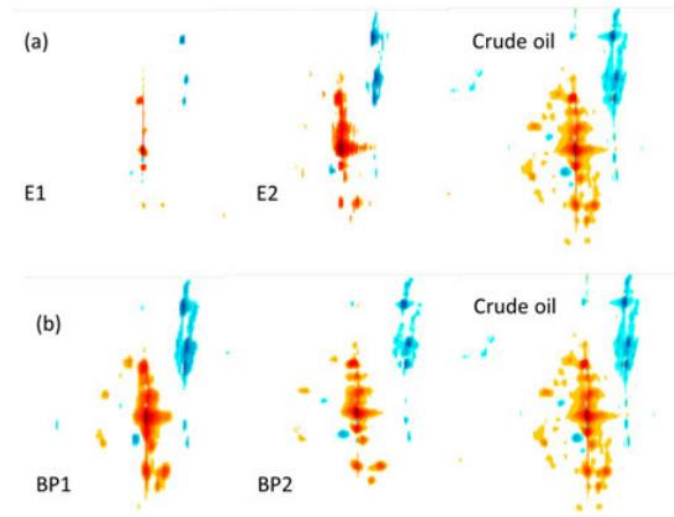
Chemical shift  $\delta$ ,  
integration, and  
J correlations

## Relative configuration



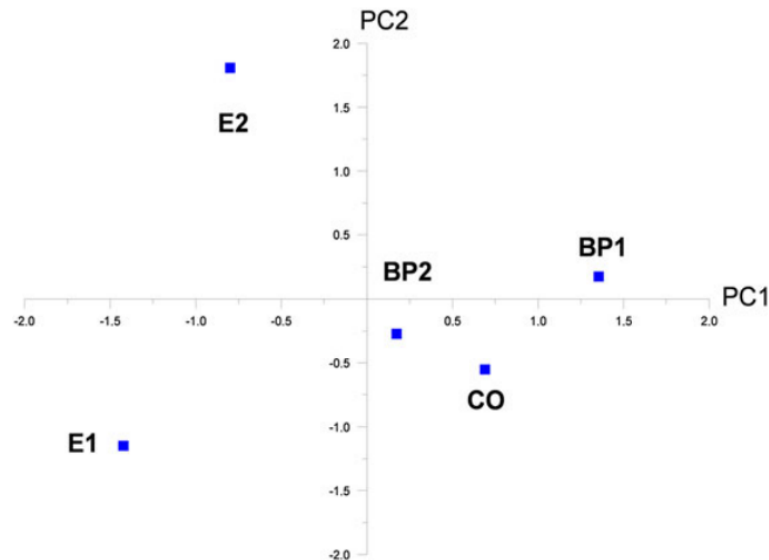
$^1\text{H}$ - $^1\text{H}$  splitting  
and NOE

# Quality control/chemometrics – oil degradation



Characterization of oil complex hydrocarbon mixtures by HSQC-NMR spectroscopy and PCA  
<https://doi.org/10.1002/poc.3233>

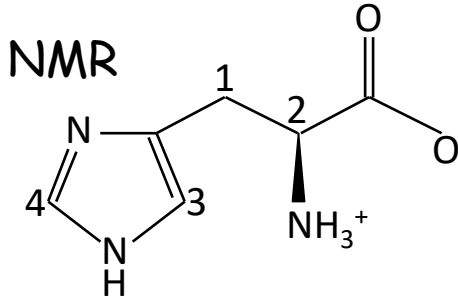
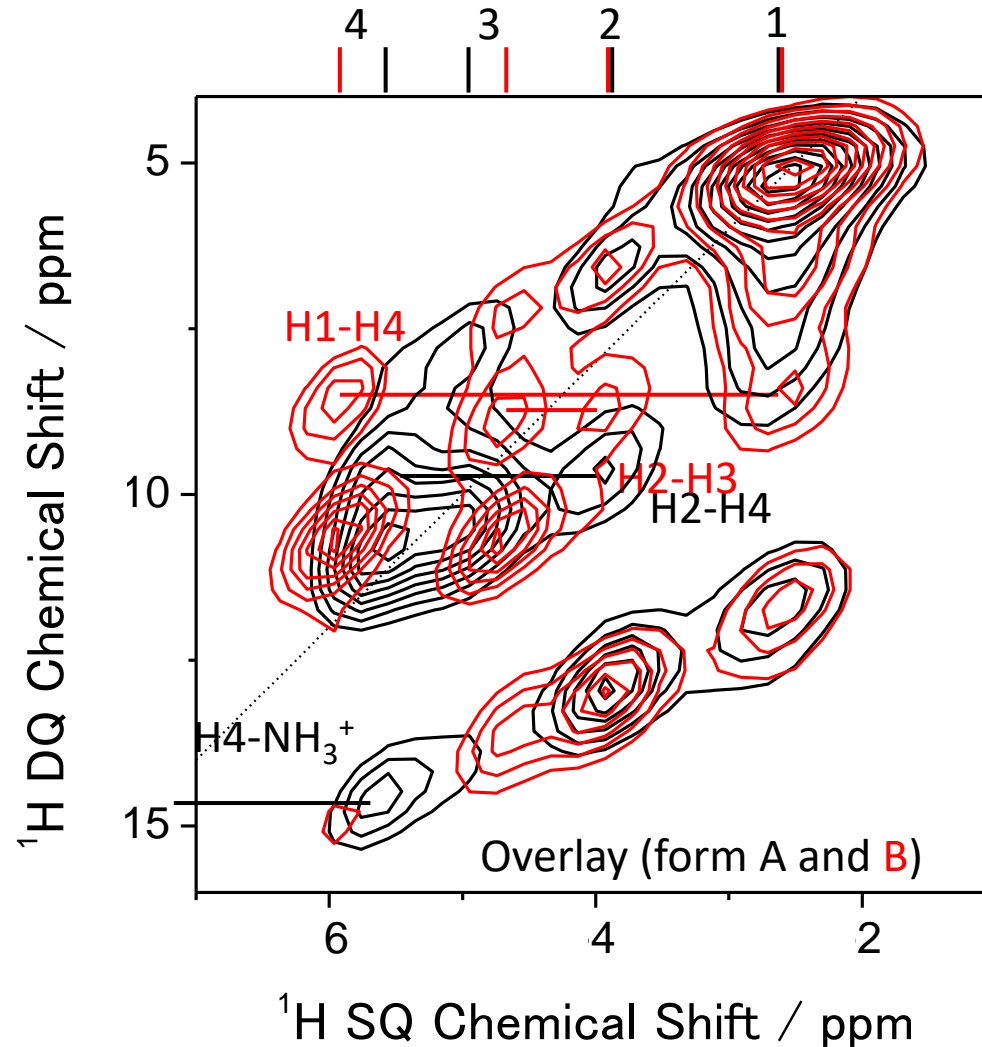
**Figure 1.** Fingerprint of the expanded HSQC-NMR spectra ( $\delta\text{C}/\delta\text{H}$  10–45/0.5–3.0 ppm). (a) Crude oil and samples with natural attenuation (E1 and E2); (b) crude oil and biopiles (BP1 and BP2); f1:  $\delta\text{C}$ ; f2:  $\delta\text{H}$ ;  $-\text{CH}_3$  and  $-\text{CH}$  (blue);  $-\text{CH}_2$  (orange)



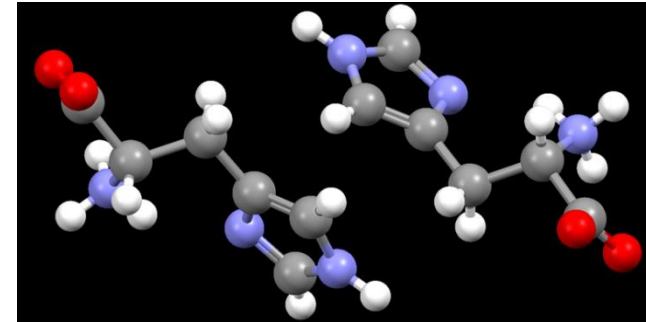
**Figure 3.** Score plot: crude oil (CO), residues with natural attenuation (E1 and E2), and biopiles (BP1 and BP2)

# Intermolecular interactions

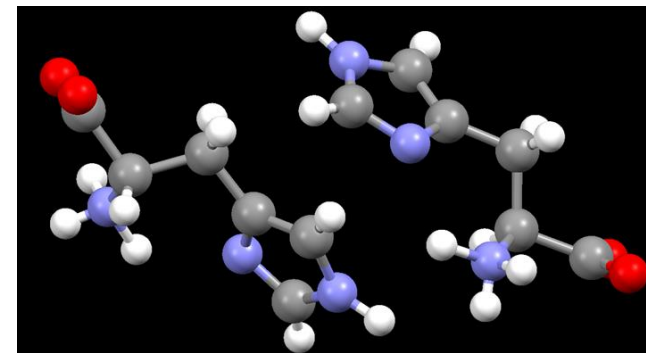
Determining molecular packing of L-histidine by solid-state NMR



Form A

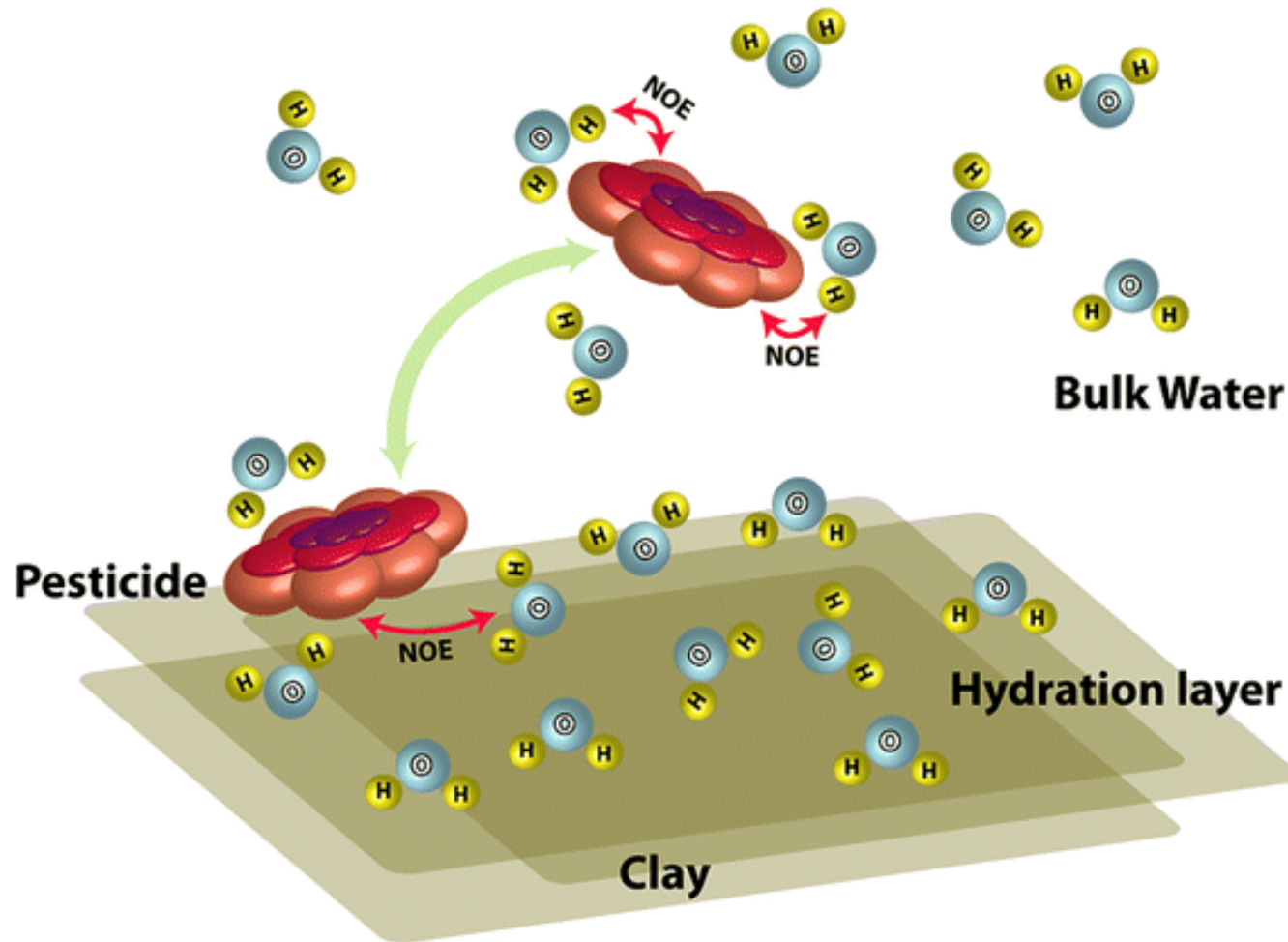


Form B



T. Oikawa, M. Okumura, T. Kimura, Y. Nishiyama, Acta Cryst. C73 (2017) 219–228.

# Intermolecular interactions pesticide – soils, drugs – proteins, etc.



WaterLOGSY  
using liquid-state  
probes

Soong, R.,  
Botana, A. et al.  
Chemical and  
Biological  
Technologies in  
Agriculture  
(2017) 4:3

# Antitrypanosomal 4-phenyl-6-(pyridin-3-yl)pyrimidines interaction with rhodesain

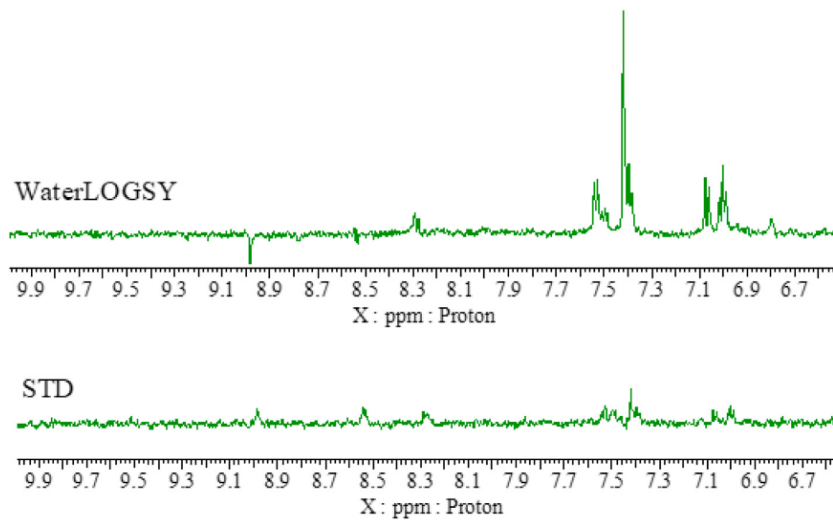
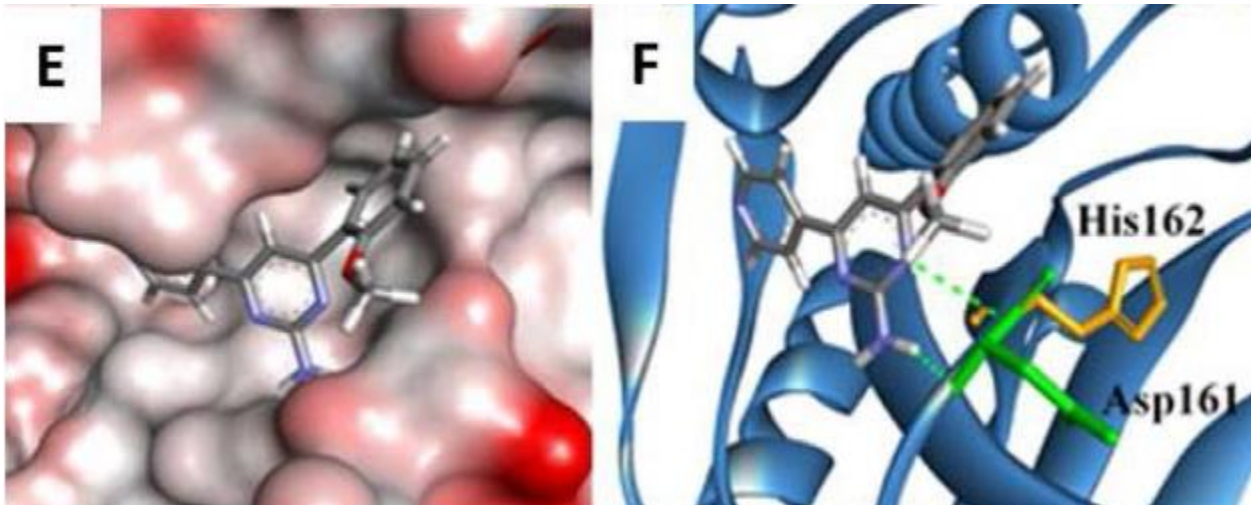
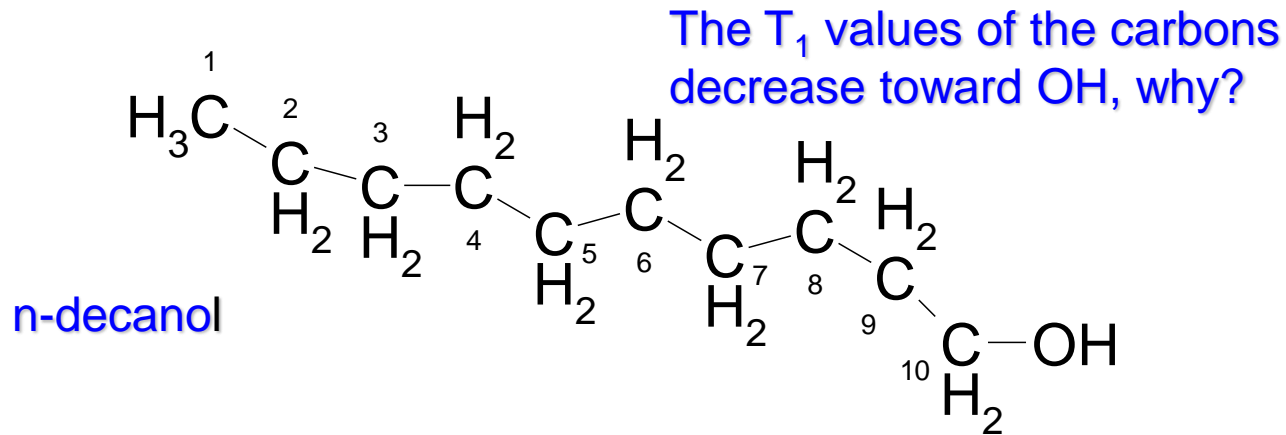


Fig. 6. STD and WaterLOGSY spectra of compound 13 in the presence of rhodesain.

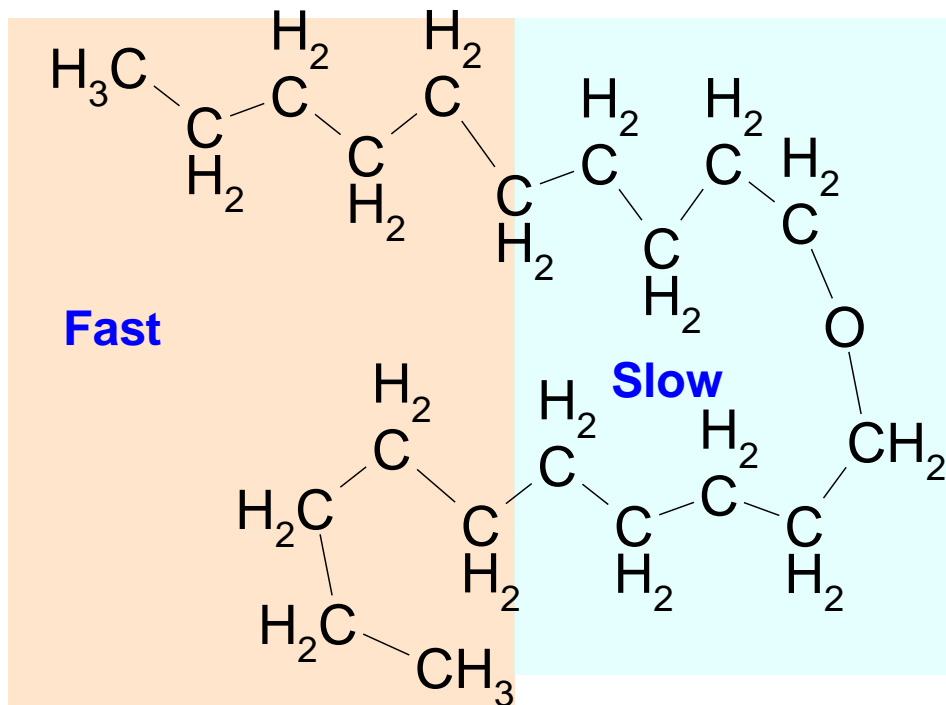
<https://doi.org/10.1016/j.ejmech.2020.112871>



# Information from T<sub>1</sub>



Carbon	T <sub>1</sub> /sec
1	3.1
2	2.2
3	1.6
4	1.1
5	0.84
6	0.84
7	0.84
8	0.77
9	0.77
10	0.65



Those T<sub>1</sub> values prove the two molecular complex by hydrogen bound



# Information from $T_1$ in solids

**Table 2.**  $^1\text{H}$   $T_1$  relaxation times for  $\alpha$ - and  $\gamma$ -CD, PCL-PEO-PCL tri-block copolymer, and inclusion complexes of the CDs and copolymer

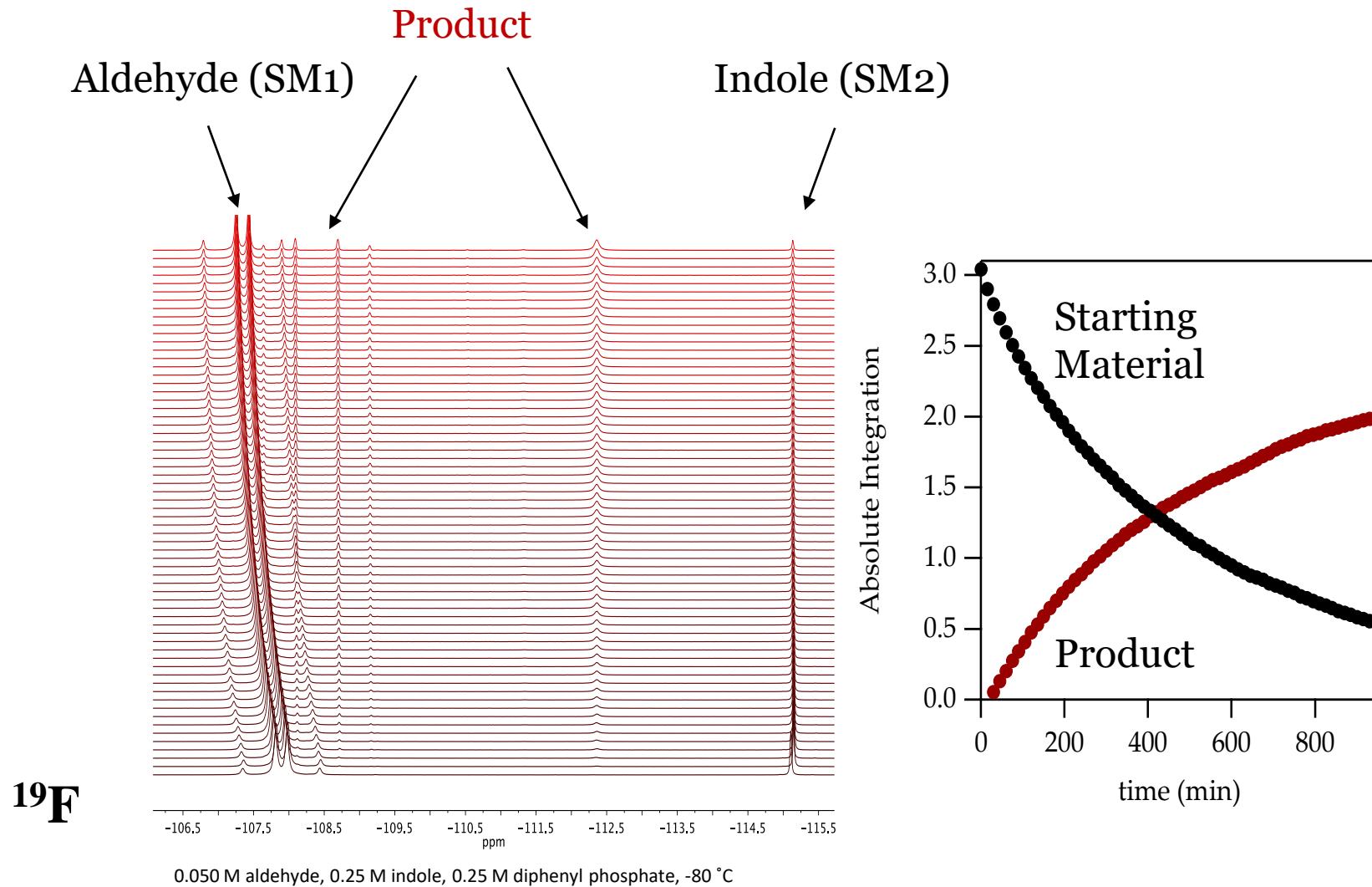
Sample	$T_1$ (s) PCL-PEO-PCL	$T_1$ (s) CD
$\alpha$ -CD	–	3.50
$\gamma$ -CD	–	2.35
PCL-PEO-PCL	1.40	–
PCL-PEO-PCL- $\alpha$ -CD-IC	1.86	1.83
PCL-PEO-PCL- $\gamma$ -CD-IC	1.07	1.02

The similarity of relaxation times for the CDs and the copolymer in the inclusion complexes demonstrates homogeneity of the complex. Adapted from [24].

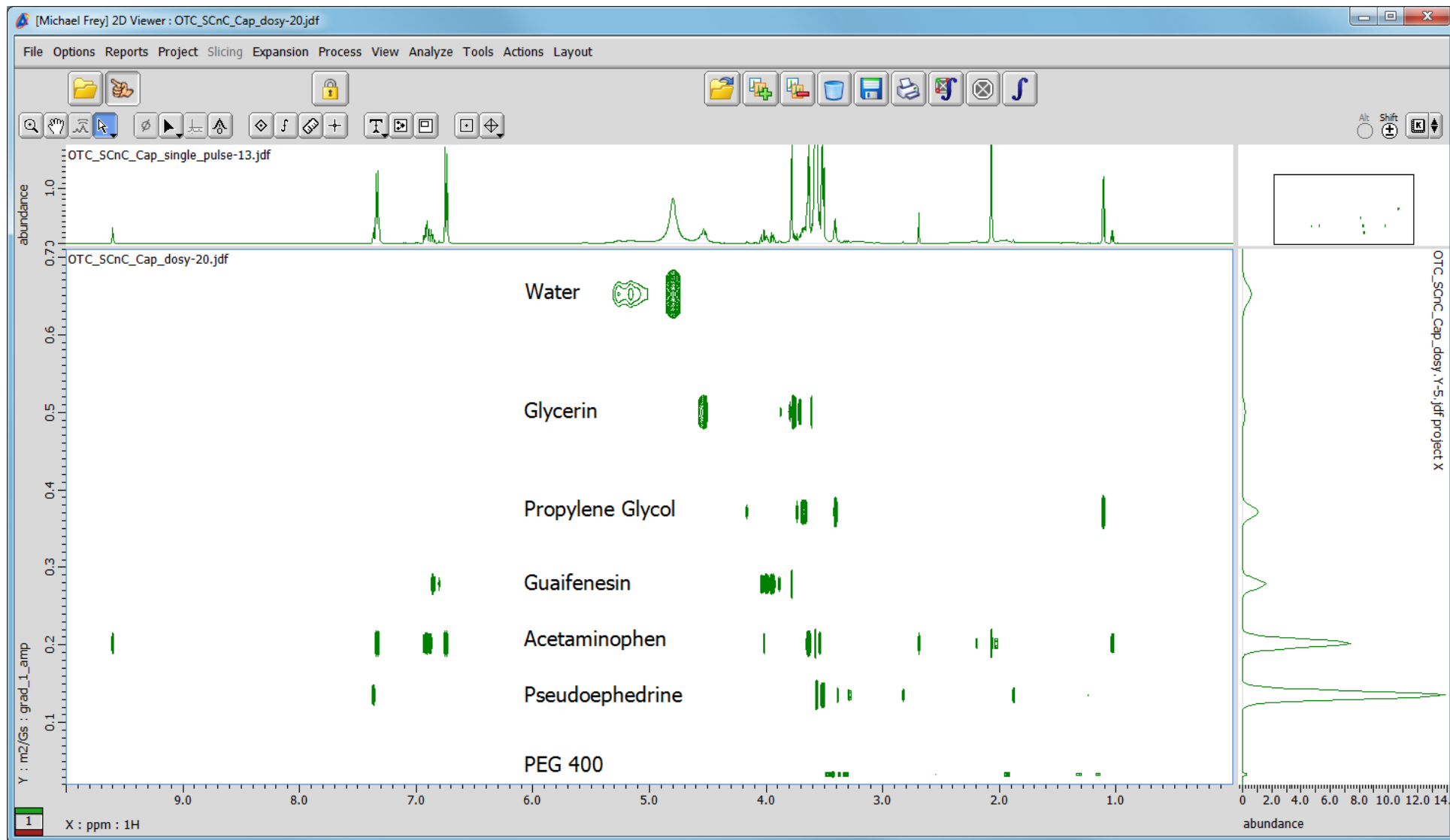
<https://doi.org/10.1016/j.trac.2006.07.006>



# Reaction Monitoring

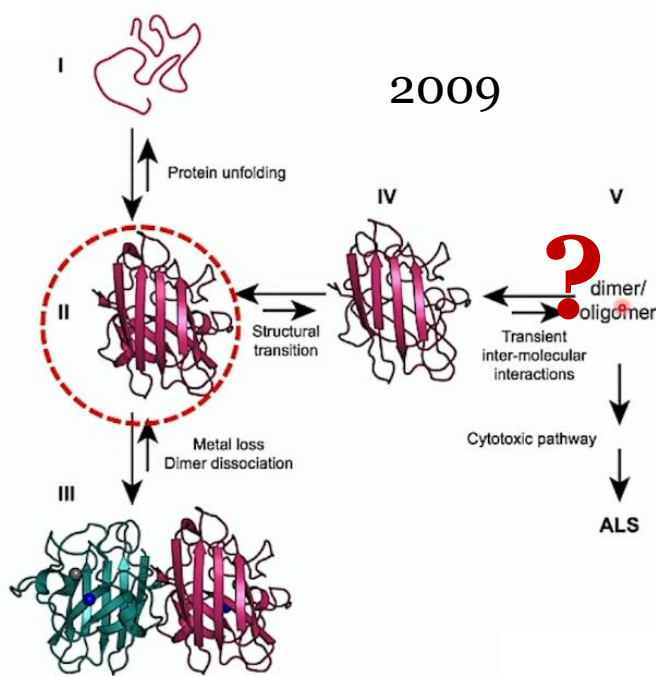


# DOSY NMR of paracetamol tablet

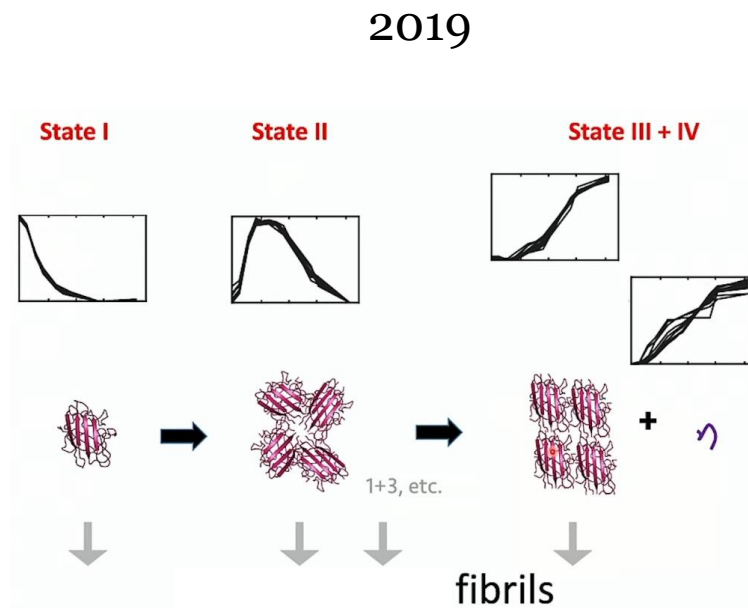


Solvent Methanol- $\text{d}_4$

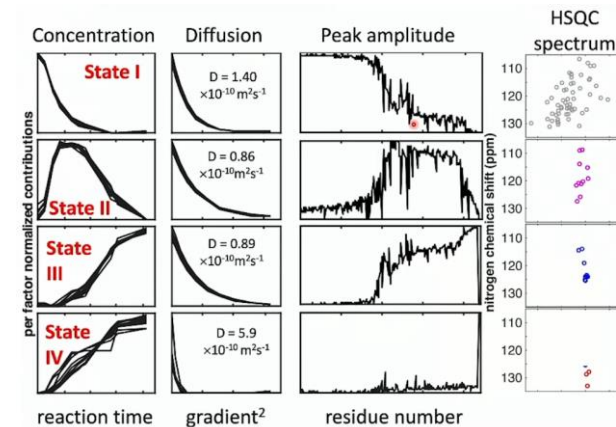
# Insight into Amyotrophic Lateral Sclerosis via DOSY and reaction monitoring



<http://dx.doi.org/10.1073/pnas.0907387106>

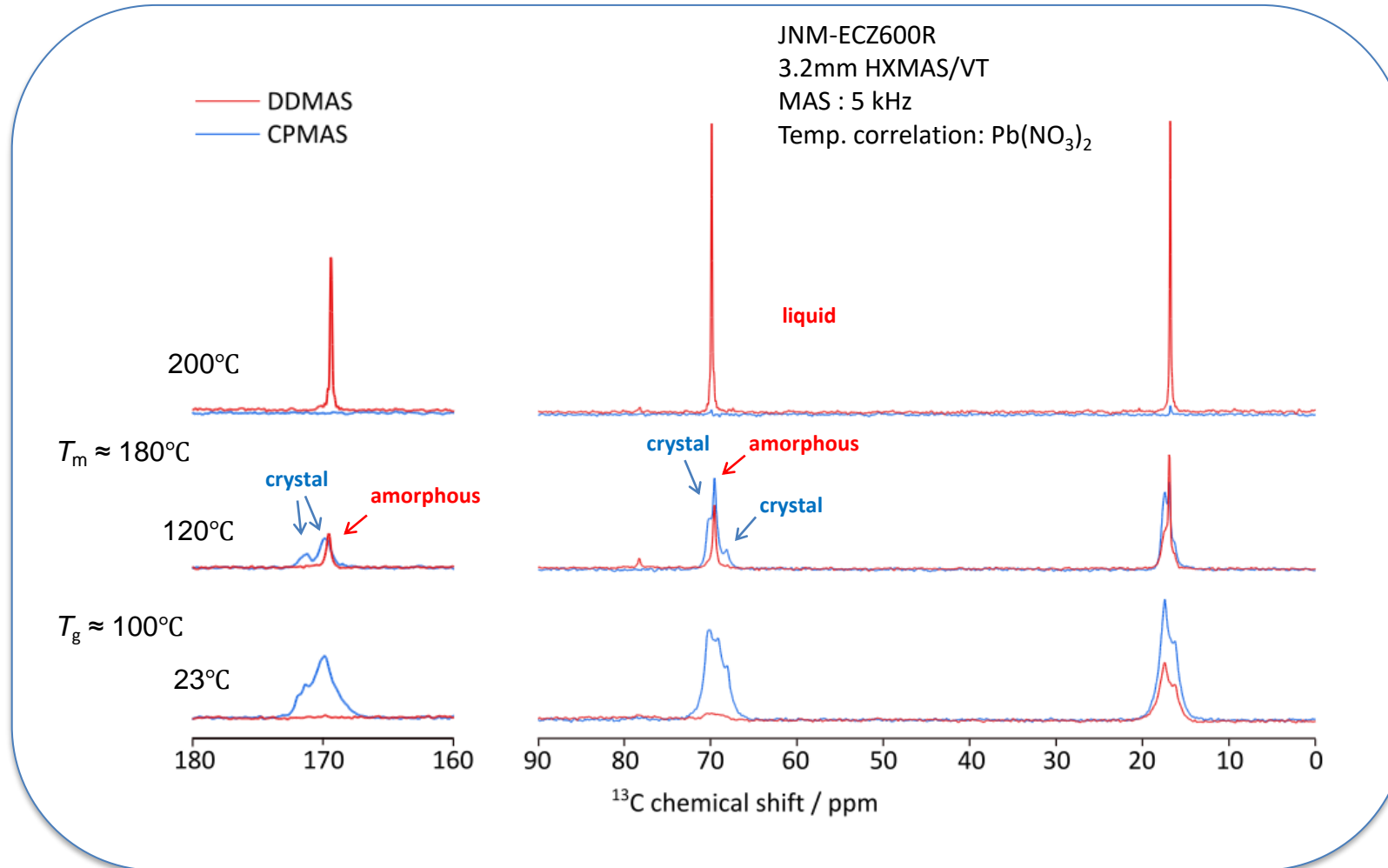


<https://doi.org/10.1021/jacs.9b07952>



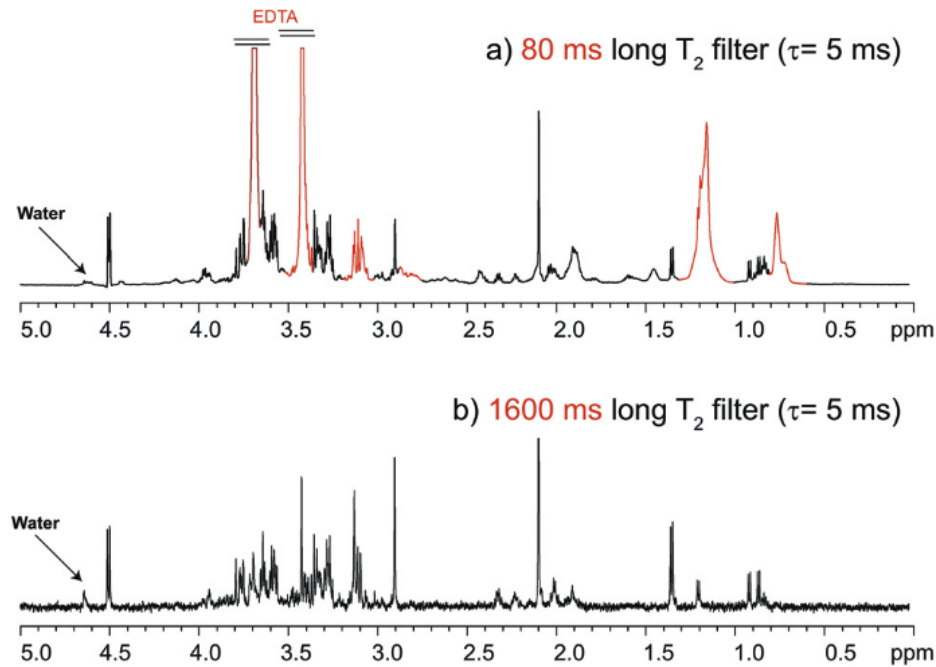
# Polymer analysis

poly(L-lactic acid) using extended VT solid-state probe

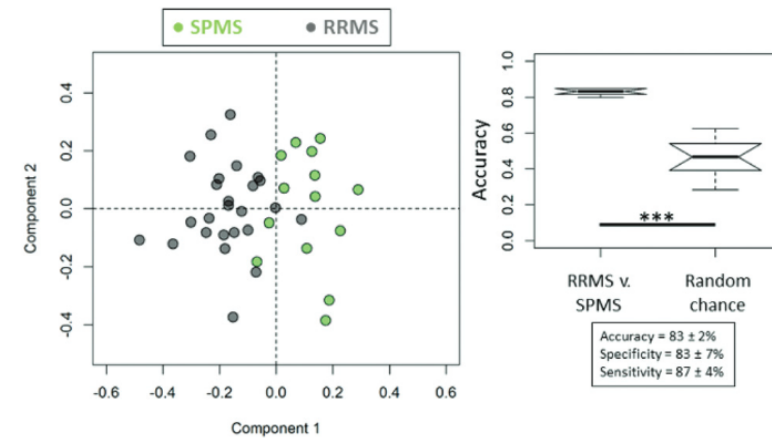


Sample – courtesy of Prof. Naoki Asakawa, Gunma University

# Biological samples analysis



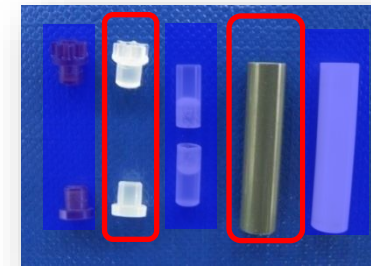
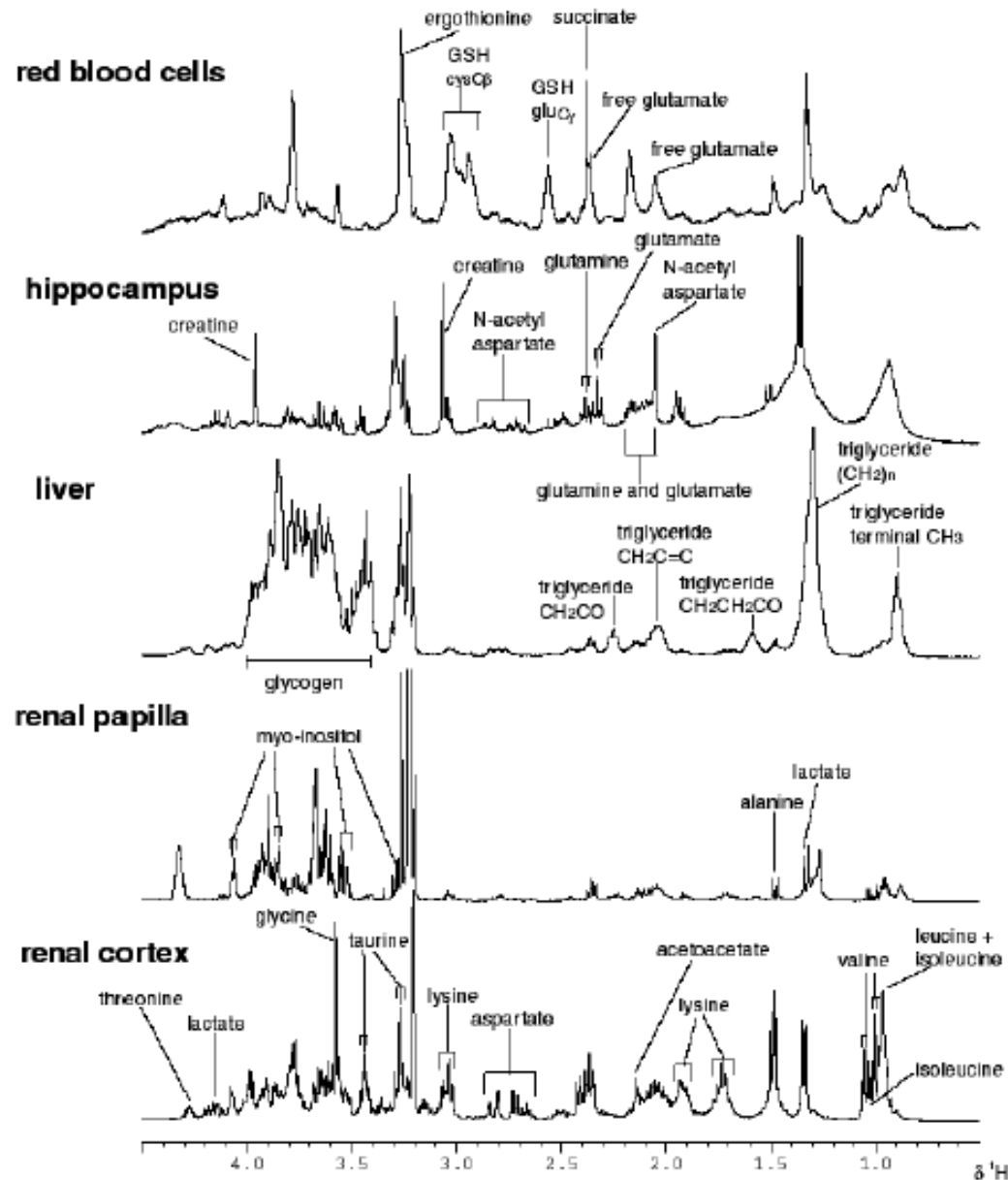
**Fig. 5** Wasted pulse sequences allow the use of  $T_2$  filters of a length impossible to use achieve with presat-CPMG. This allows the introduction of  $T_2$  encoding to distinguish between different metabolite populations. Compare (b), in which a 1600 ms long filter has been made possible by Wasted-II, with (a) in which an 80 ms filter has been used (also using Wasted-II). Note that the suppression of the water signal is excellent even when using these long filters. The sample is human blood with non-deuterated EDTA.



**Fig. 6** OPLS-DA results from blood plasma Wasted-II  $^1\text{H}$  NMR data discriminating SP from RR multiple sclerosis patients. Left, a representative scores plots illustrating separation between SPMS and RRMS plasma spectra in the multivariate models, and right, the accuracy of the cross-validated ensemble of OPLS-DA models is significantly better than random chance. Kolmogorov–Smirnov test  $p$ -values  $<0.001$  are represented by \*\*\*.

<https://doi.org/10.1039/C9AN01005J>

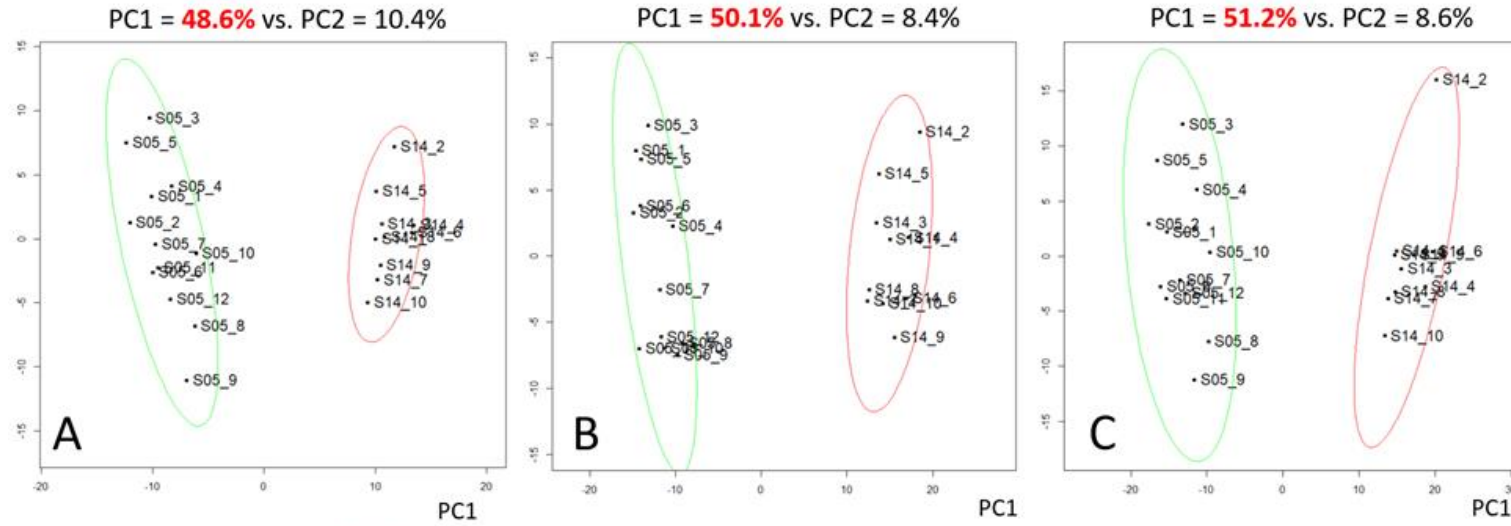
# Semi-solids analysis in biology



Study of tissues using FGMAS probe reveals clear differences between them



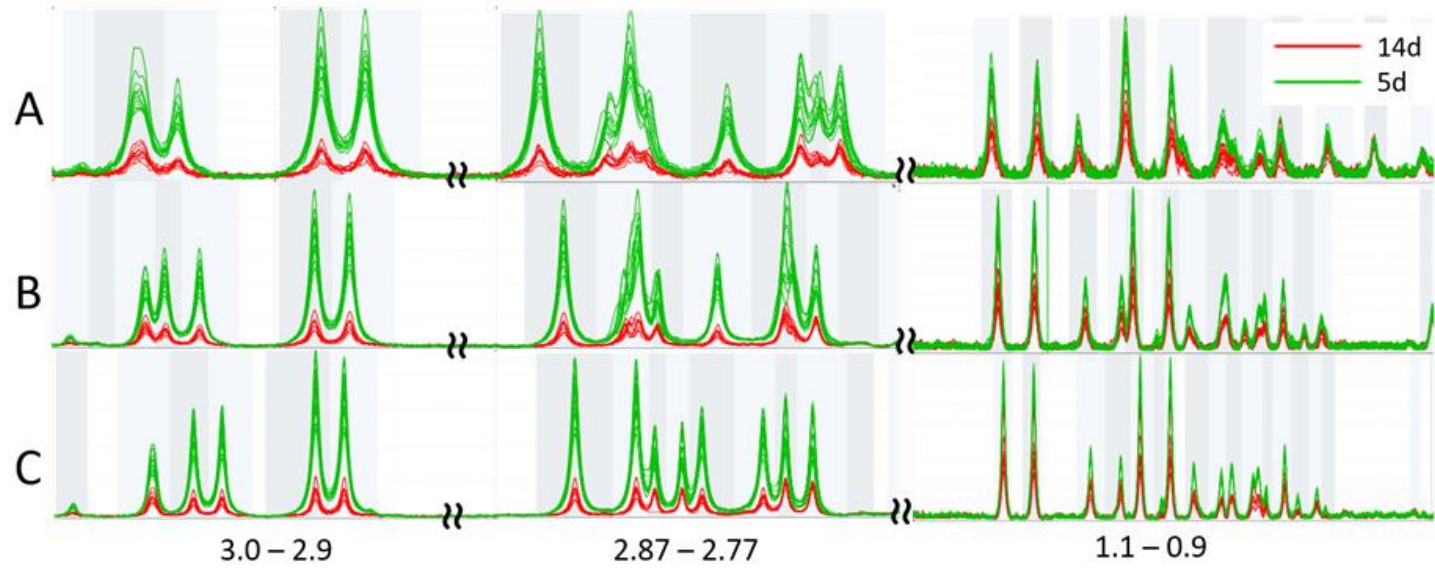
# Metabolomics analysis



**A: JEOL 400**  
**B: Bruker 500**  
**C: Bruker 600**

**Green: 5 days**  
**after flowering**

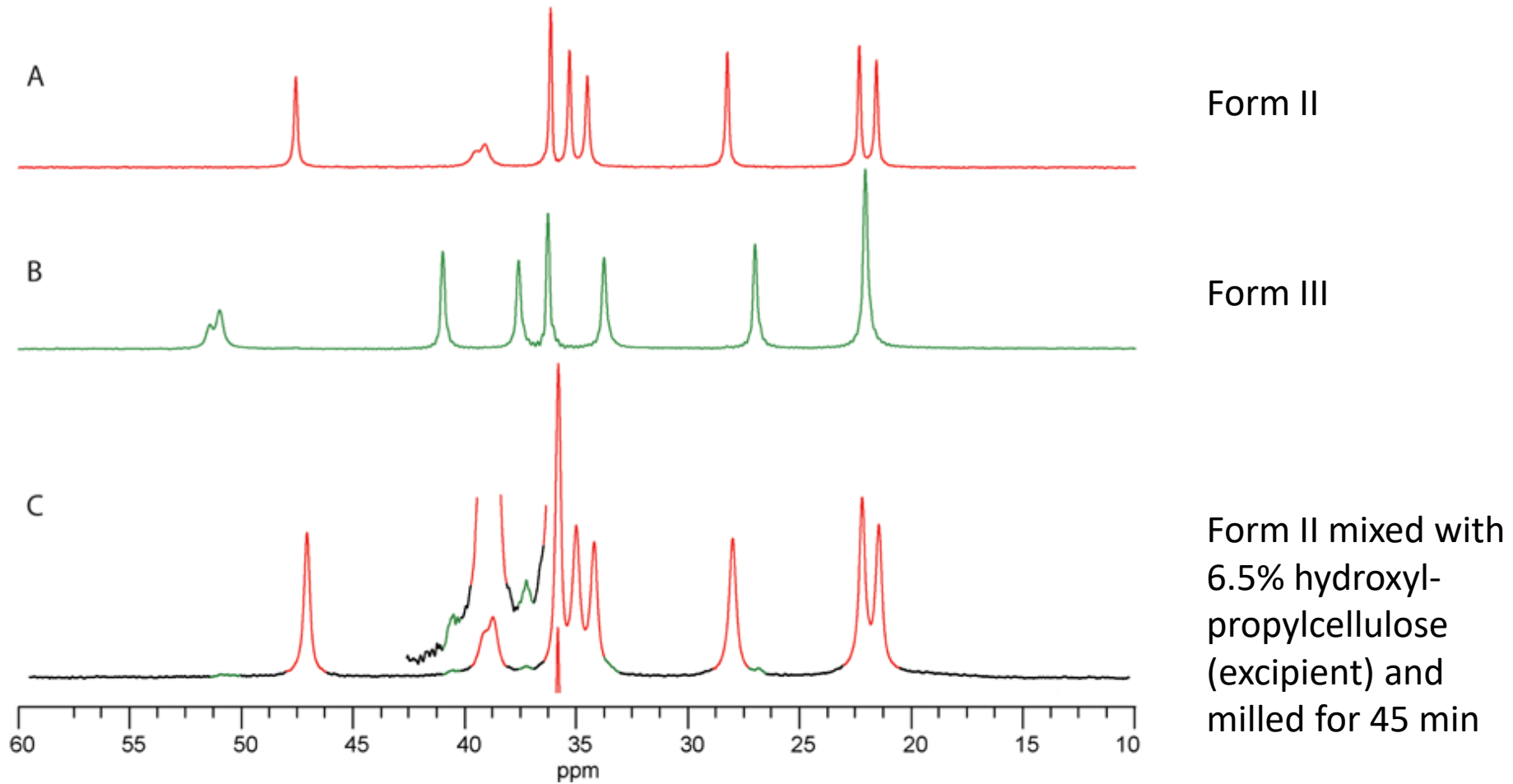
**Red: 14 days**  
**after flowering**



Deborde, C.,  
 Fontaine, JX.,  
 Jacob, D. Botana,  
 A. et al.  
 Metabolomics  
 (2019) 15: 28



# Quantitation of gabapentin polymorphs



<https://www.europeanpharmaceuticalreview.com/article/17182/applications-of-solid-state-nmr-spectroscopy-to-pharmaceuticals/>  
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3581669/>

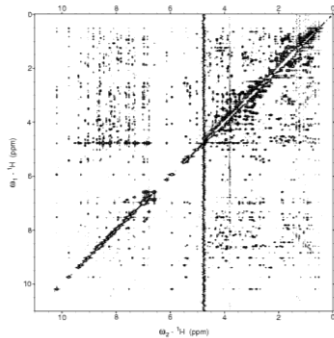
# Protein structure determination

GSDIIDEFGTLDDSATICRVCQKPG  
DLVMCNQCEFCFHLDPALQD  
VPGEEWSCSLCHVLPDLKEEDVDL  
QACKLN

Protein sequence

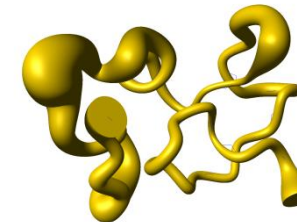
Express and  
purify protein  
(or isolate from  
natural source)

Initial characterisation  
- Identity, composition  
- Concentration  
- Stability (buffers, salt,  
pH, temperature)



Acquire NMR spectra


Evaluation:  
Sequential Assignment  
Extraction of distance restraints  
and other structural data



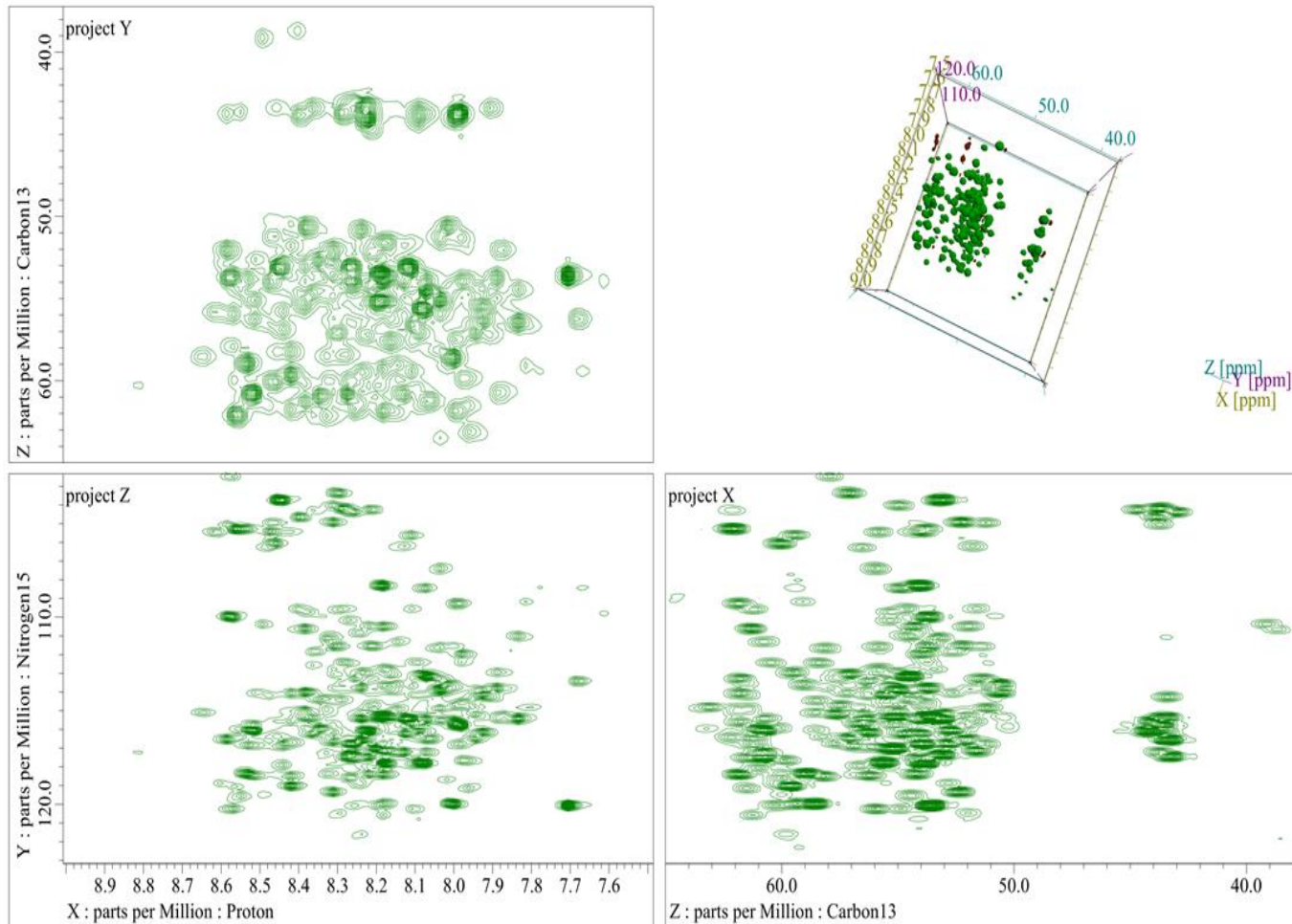
3D structure

# Protein spectra using UltraCOOL HCN 800

3D HNCOCA-TROSY, 2H decoupled, 20 hours

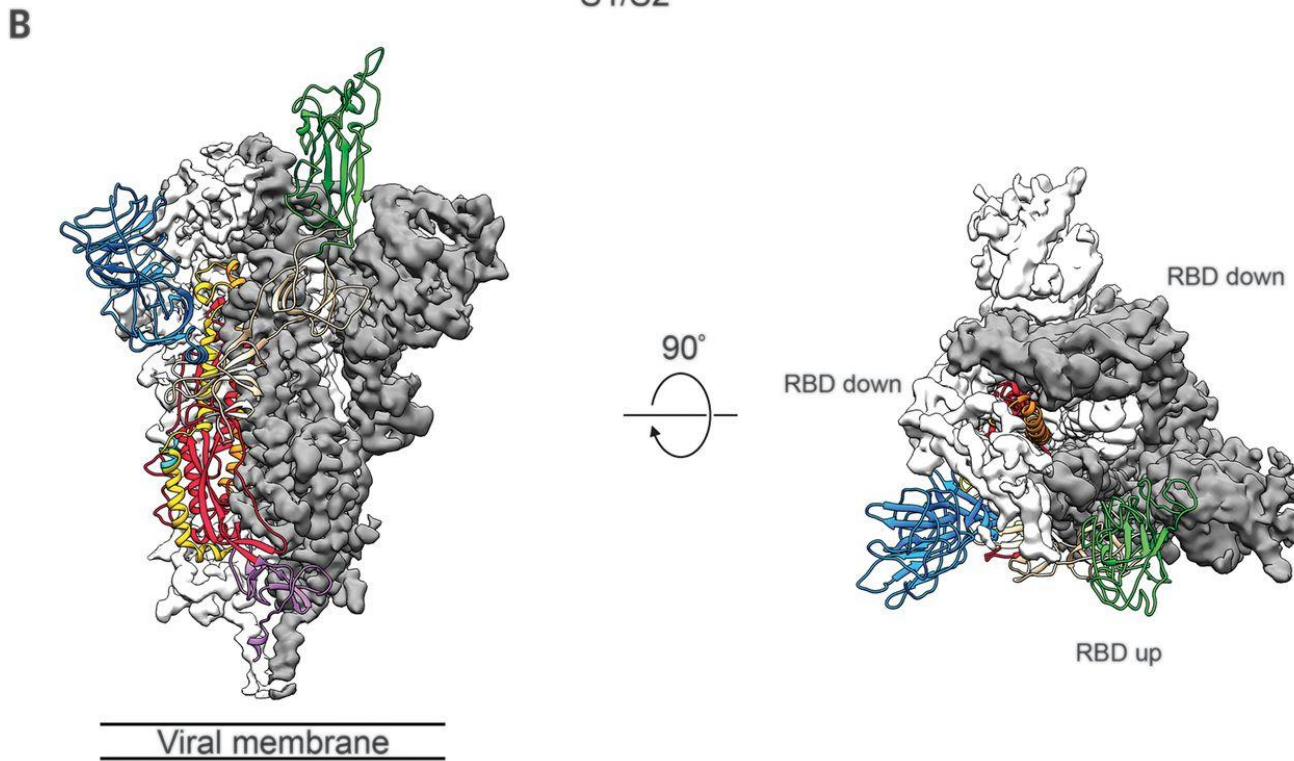
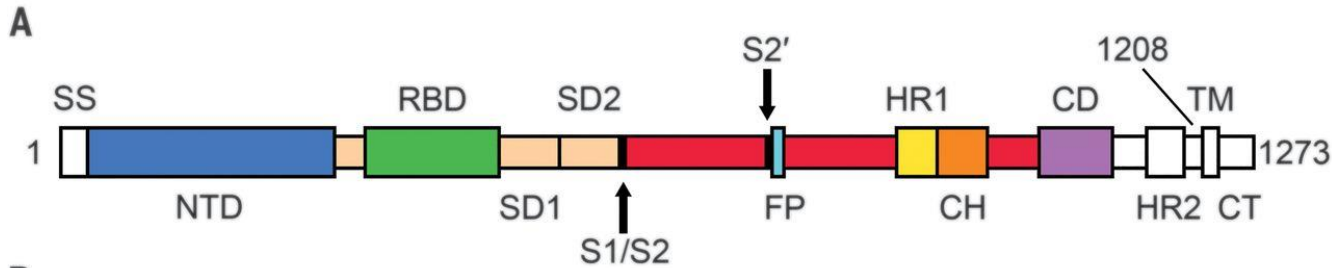
~0.2 mM Stt3p\* (~30kDa) in lipid micelles at 55C

\*Protein sample was kindly provided by Professor Smita Mohanty, Oklahoma State University.



<https://www.protein-nmr.org.uk/solution-nmr/spectrum-descriptions/hncoca/>

# But... March 2020, in Science

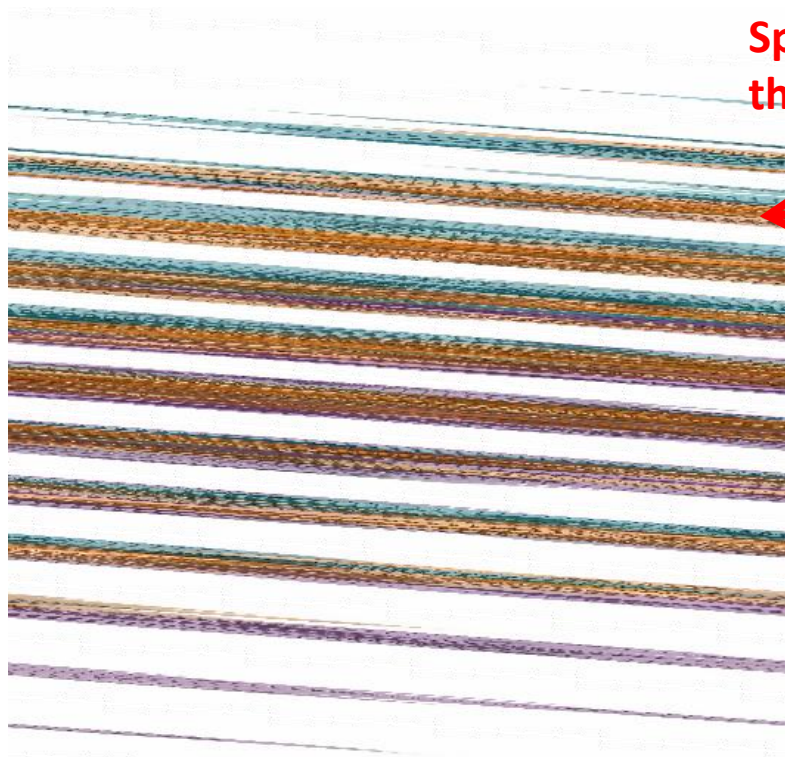


**Cryo-EM enabled quick structure identification of spike protein of COVID-19**  
DOI:  
[10.1126/science.abb2507](https://doi.org/10.1126/science.abb2507)

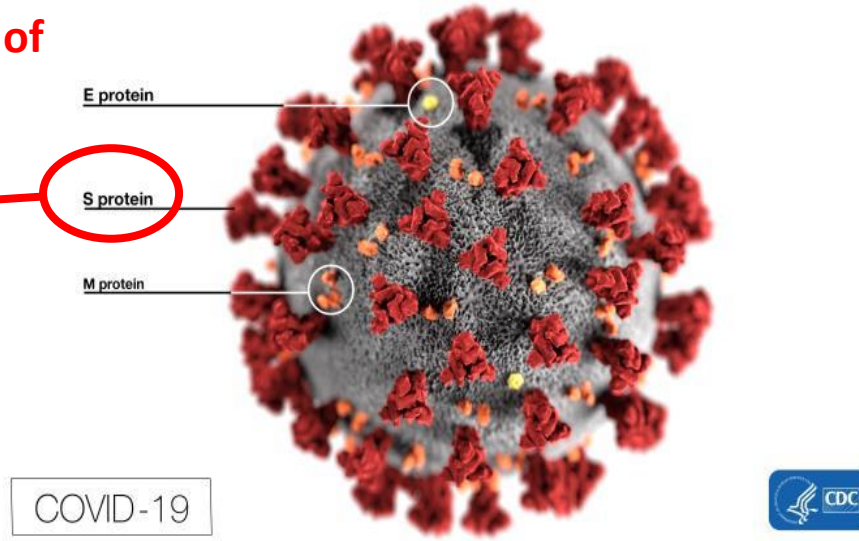
**X-Ray and cryo-EM are preferred for determining protein structures**  
**NMR still main method to determine structure in disordered proteins**



# But... Molecular dynamics (simulation with cryo-EM and SAXS data & Folding@Home)



Spike protein of  
the COVID-19



COVID-19

<https://blogs.plos.org/dnascience/files/2020/02/ESM-proteins.jpg>



**Greg Bowman** @drGregBowman · Apr 3

Replying to @drGregBowman

The three colors are the three proteins that come together to form the spike. Each is made of a linear chain of chemicals called amino acids. The ribbons trace out each chain. The transparent surface is the surface of the spike.

2

9

92



**Greg Bowman** @drGregBowman · Apr 3

The three proteins that make up the #Demogorgon/spike must spread apart to reveal the ACE2 binding site, which initiates infection by attaching to a protein called ACE2 on the surface of human cells. This movie captures part of that opening motion.

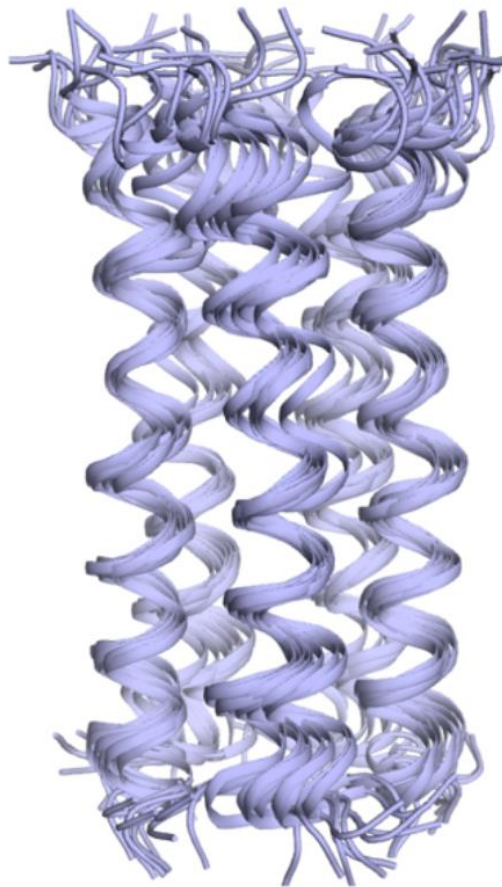


**Greg Bowman** @drGregBowman · Apr 3

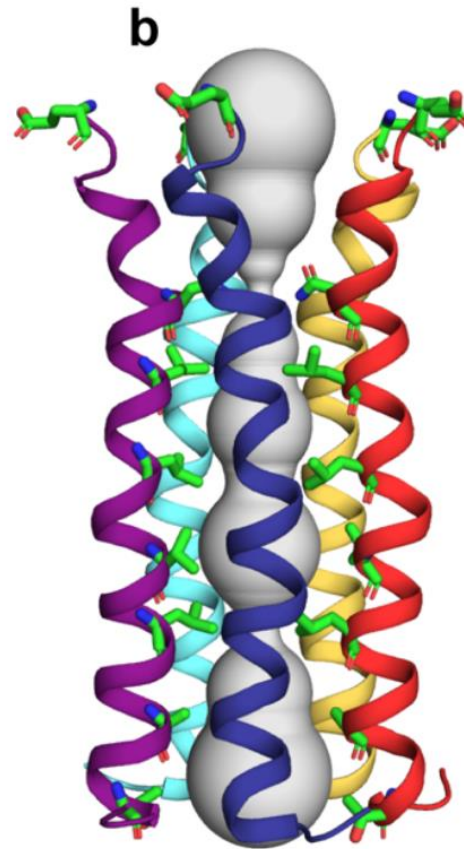
The jitteriness of the movie isn't an artifact, its representative of the stochastic motions on this size scale (i.e. random motions with different probabilities)

<https://twitter.com/drGregBowman/status/1246106316864708608>

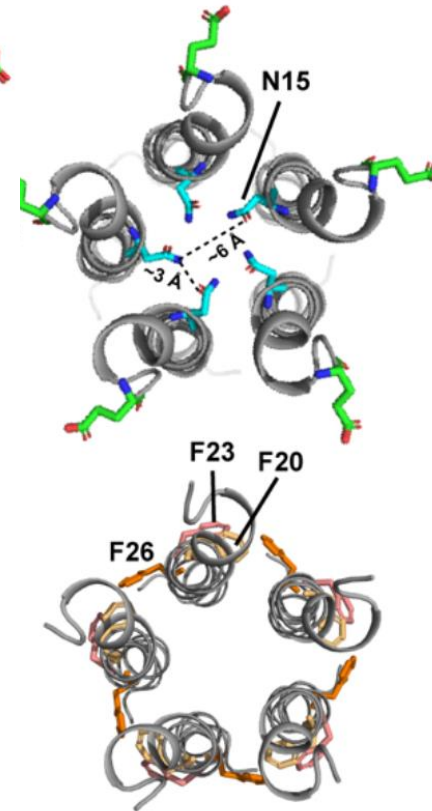
# And... structure of the SARS-CoV-2 envelope protein TM domain via solid-state NMR (3.2mm and 1.9mm MAS)



Ensemble of the ten lowest-energy structures

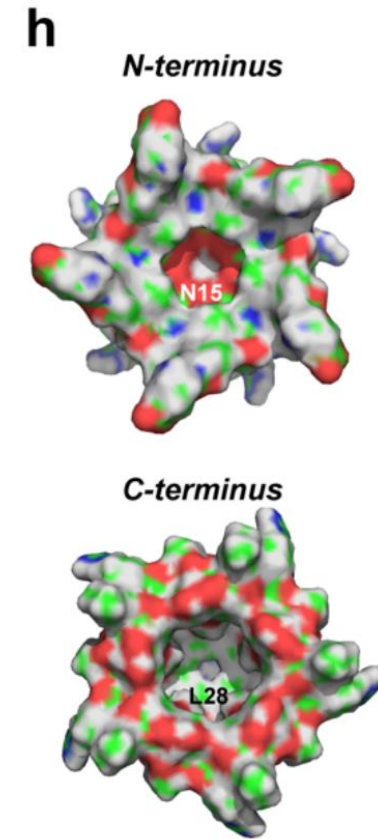


Sideview of the lowest-energy structure along with pore water(gray)



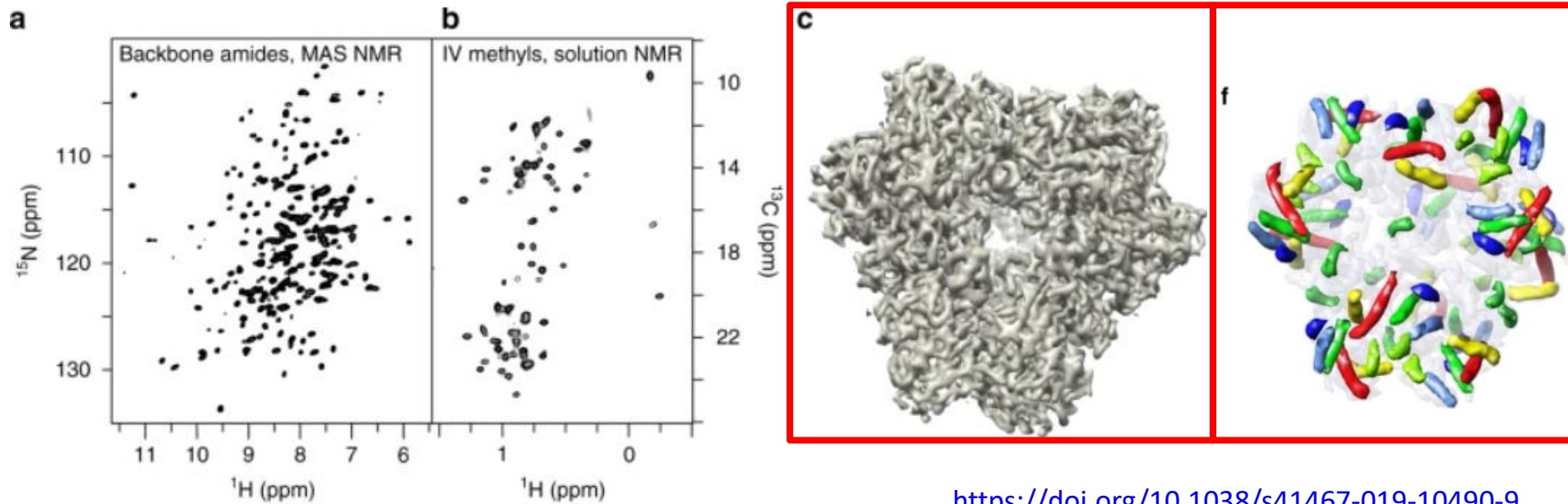
Top views of the **disordered** N-terminal E8 and surface plots

<https://doi.org/10.21203/rs.3.rs-77124/v1>

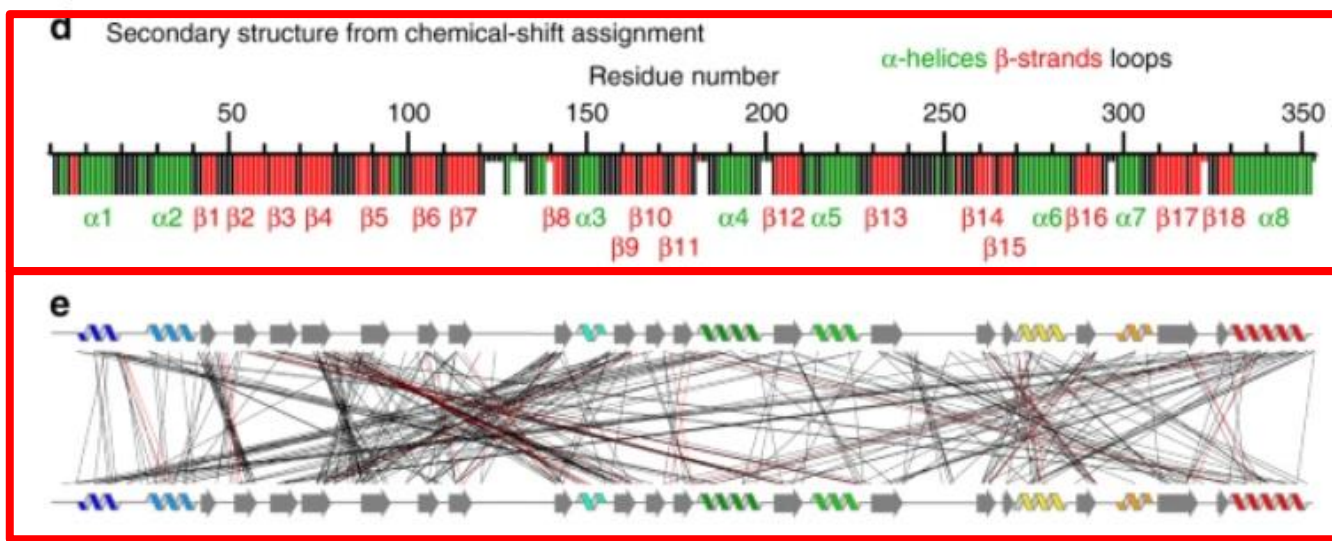




# Structural determination of 468 kDa TET2 with NMR and cryo-EM



<https://doi.org/10.1038/s41467-019-10490-9>

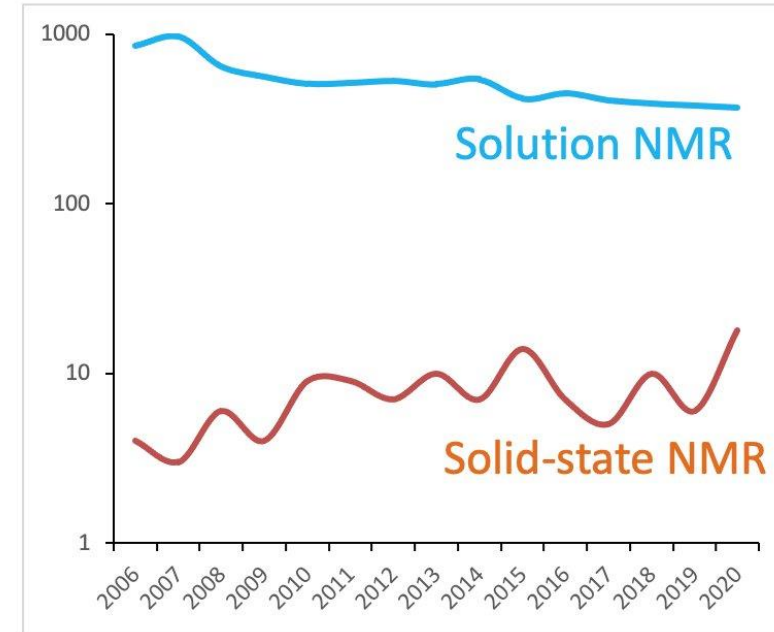
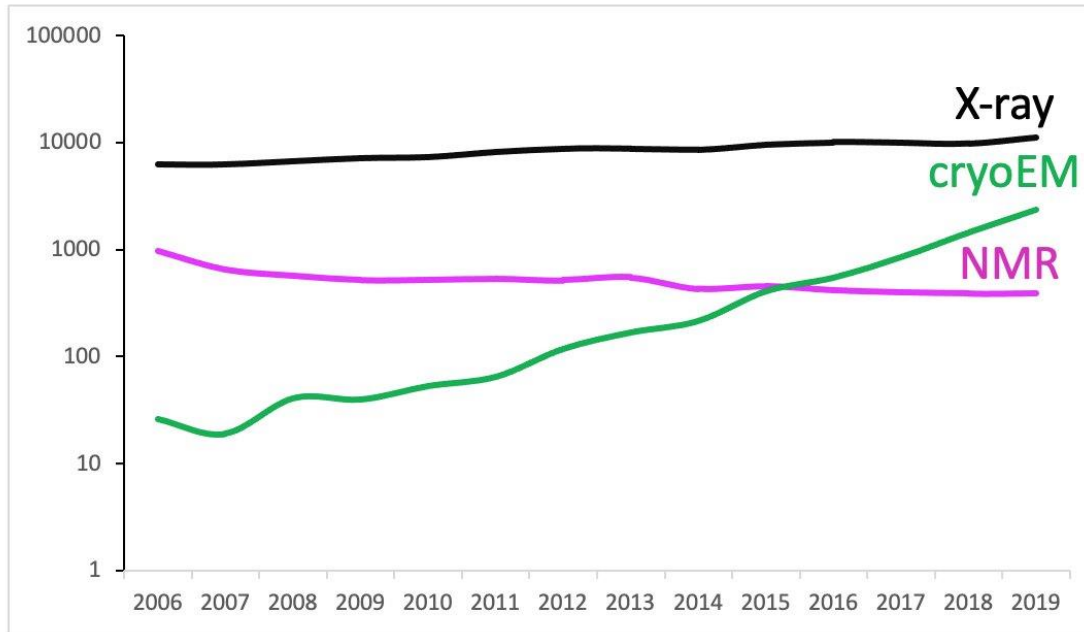


Tertiary structures can now also be predicted from primary structure  
<https://deepmind.com/blog/article/alphafold-a-solution-to-a-50-year-old-grand-challenge-in-biology>

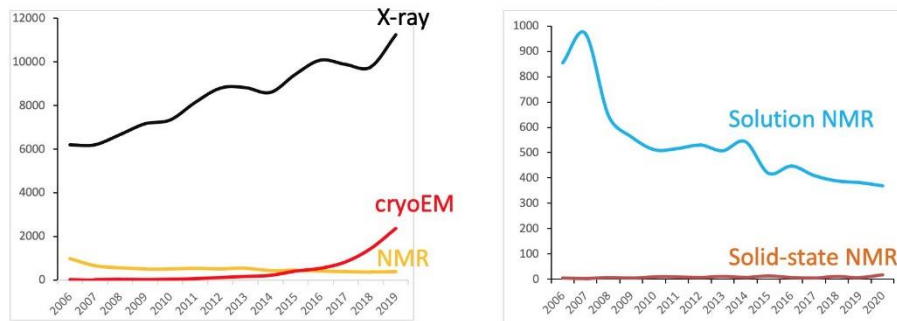


# Protein structures submitted to pdb

New entries to the pdb database/year



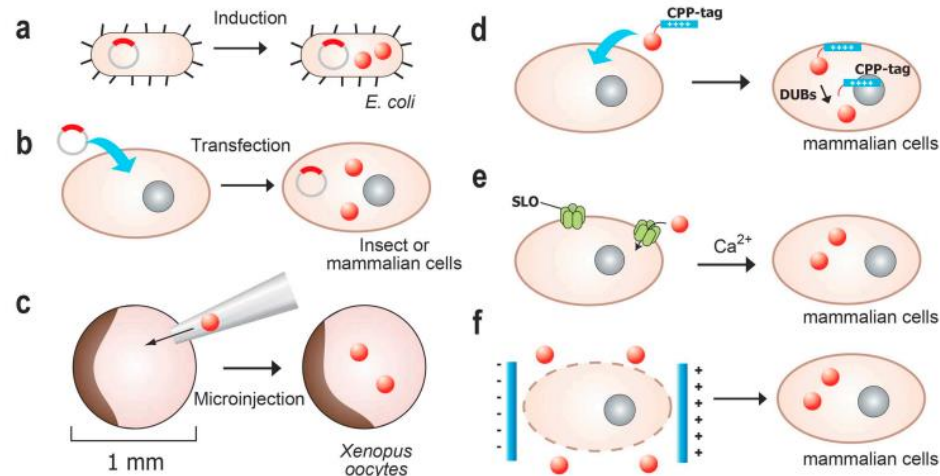
New entries to the pdb database/year



Linear scale

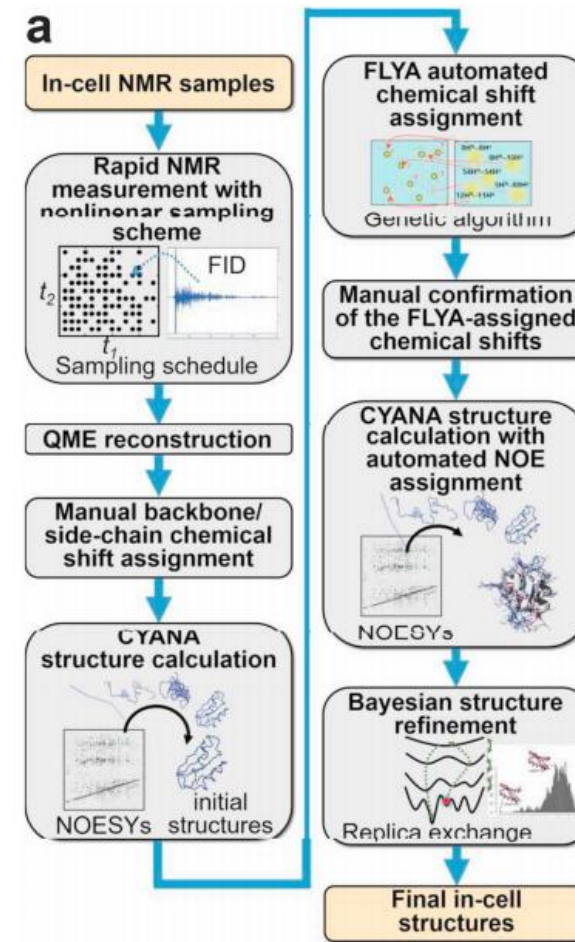
<https://twitter.com/MarkusWeingarth/status/1443299113026654215>

# In-cell NMR

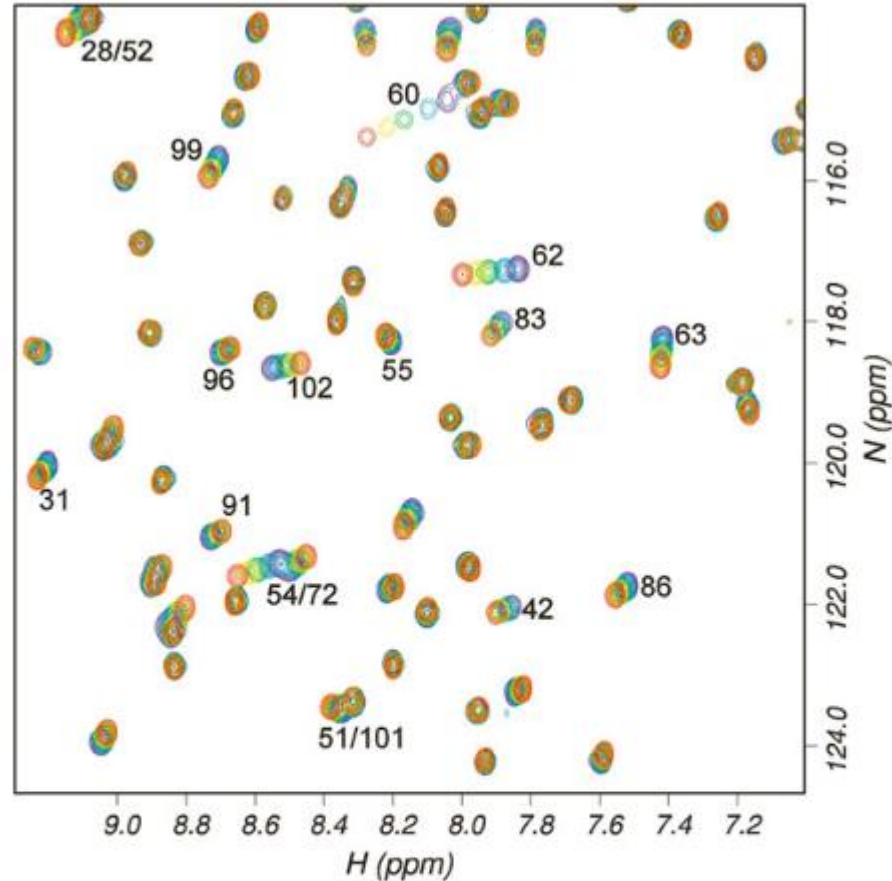


**Fig. 1.** Overview of the methods to prepare the cells enriched with isotope-labeled proteins. (a–b) Overexpression of target proteins within the cells for (a) bacterial cells and (b) eukaryotic cells, such as insect cells or mammalian cells. Cells were cultured in the medium containing the isotope enriched medium after IPTG induction or transfections of expression vectors. (c) The target proteins or nucleic acids were injected into *X. laevis* oocytes by microinjection. (d) The arginine-rich cell-penetrating peptide (CPP) guide the translocation of the target protein (Ub) across the plasma membrane. The CPP tag, which contains a positive charge cluster, tends to stick to a negatively charged surface, such as a plasma membrane. Therefore, the release of the target protein from the CPP tag by endogenous DUBs is necessary for NMR observation. (e) The target proteins are introduced into the mammalian cells through the pores formed by SLO. (f) The cells are permeabilized by electroporation to transduce the target proteins.

<https://doi.org/10.1039/C3MT00224A>



# Chemical shift perturbation



**Fig. 1** Part of a  $^1\text{H}$ ,  $^{15}\text{N}$  HSQC spectrum of the ribonuclease barnase. Each peak is the  $^1\text{H}$ ,  $^{15}\text{N}$  correlation for a different amino acid residue. The figure shows a superposition of HSQC spectra with addition of increasing amounts of the deoxynucleotide ligand d(GC), going from *red* (no addition) to *blue*. Protein concentration is 200  $\mu\text{M}$ , and ligand concentration goes from zero to a 4.5-fold excess. Peaks that shift are numbered by the amino acid (From Ref. [29], with permission from Springer Science + Business)

[https://doi.org/10.1007/978-3-319-28388-3\\_76](https://doi.org/10.1007/978-3-319-28388-3_76)