

# NMR Training Course

9<sup>th</sup> September 2021  
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JEOL UK Demo Lab



# Diffusion-Ordered Spectroscopy

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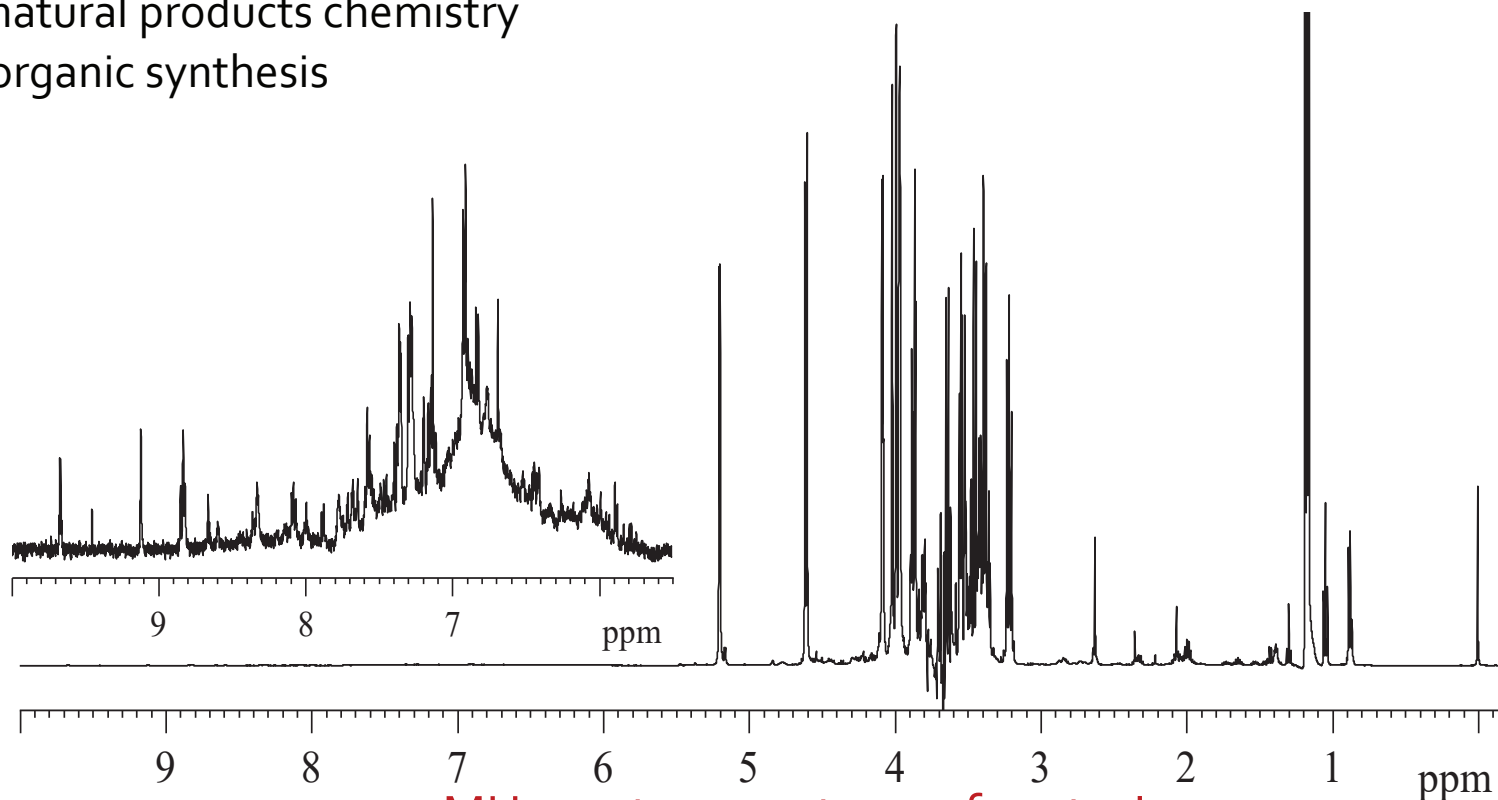
# Mixture analysis by NMR

## applications

metabolomics  
drug development  
process chemistry  
food science  
natural products chemistry  
organic synthesis

## pros/cons

- + structural information
- + nondestructive
- low(ish) sensitivity
- usually needs separation (e.g. LC-NMR)



500 MHz proton spectrum of port wine

# Self-Diffusion

- Molecules experience both rotational and translational Brownian motion
- Stokes-Einstein equation (spherical molecules) can be used to determine the mobility of the molecules

$$D = \frac{k_B T}{6\pi\eta r_H}$$

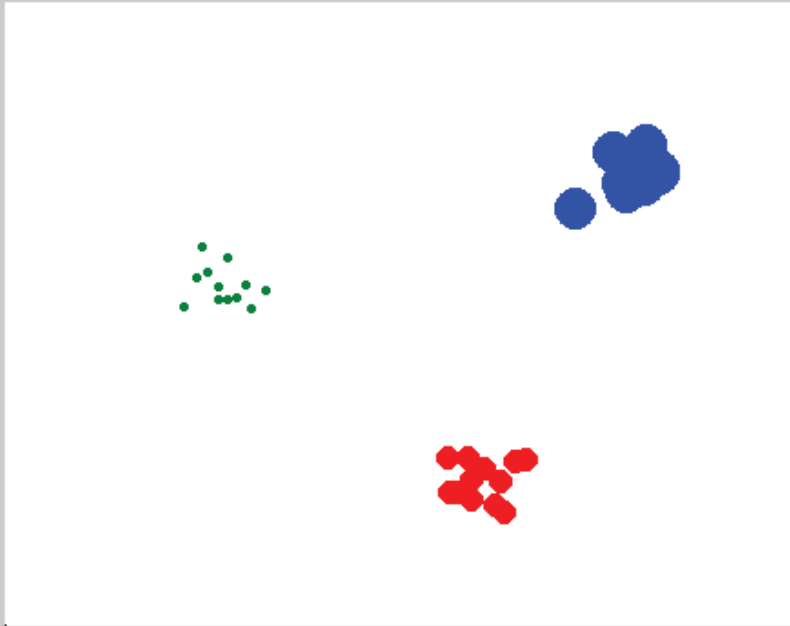
$k_B$  : Boltzmann constant

$T$ : temperature

$\eta$ : viscosity

$r_H$ : hydrodynamic radius

- The equation is valid for solute molecules at infinite dilution diffusing through a continuum solvent (i.e. where the solvent molecules are much smaller than the solute).



# The diffusion coefficient

$$D = \frac{k_B T}{6\pi\eta r_H}$$

The hydrodynamic radius  $a$  is the effective average radius of the solvated solute molecules, and will depend on the molar mass  $MW$ . Assuming similar chemistries (i.e. constant density)

- for a spherical molecule such as a globular protein,  $D \propto (MW)^{-1/3}$
- for a 'random coil' polymer or a flat disk,  $D \propto (MW)^{-1/2}$
- for a rigid linear molecule  $D \propto (MW)^{-1}$

In practice  $D$  will also depend on concentration, molecular shape, interactions etc.

# Self-Diffusion and molecular weight

- Morris approximation for the correlation between molecular weight and self-diffusion coefficient
- More advanced than previous estimation of  $r_H \propto \sqrt[3]{MW}$
- Takes into account the molecular interactions of solvent and solutes
- **Calculator available: Temperature, solvent and MW or D**

$k_B$ : Boltzmann constant

$T$ : temperature

$\eta$ : viscosity

$\rho_{\text{eff}}$ : effective density of the molecule  
(packing effects, geometry,  
solvation and flexibility)

$MW$ : molecular weight of the molecule

$MW_S$ : molecular weight of the solvent

$N_A$ : Avogadro number

$$D = \frac{k_B T \left( \frac{3 \sqrt[3]{\frac{MW_S}{MW}}}{2} + \frac{1}{1 + \sqrt[3]{\frac{MW_S}{MW}}} \right)}{6\pi\eta \sqrt[3]{\frac{3MW}{4\pi\rho_{\text{eff}} N_A}}}$$

R. Evans et al, *Angew. Chem. Int. Ed.* (2013), 52: 3199–3202

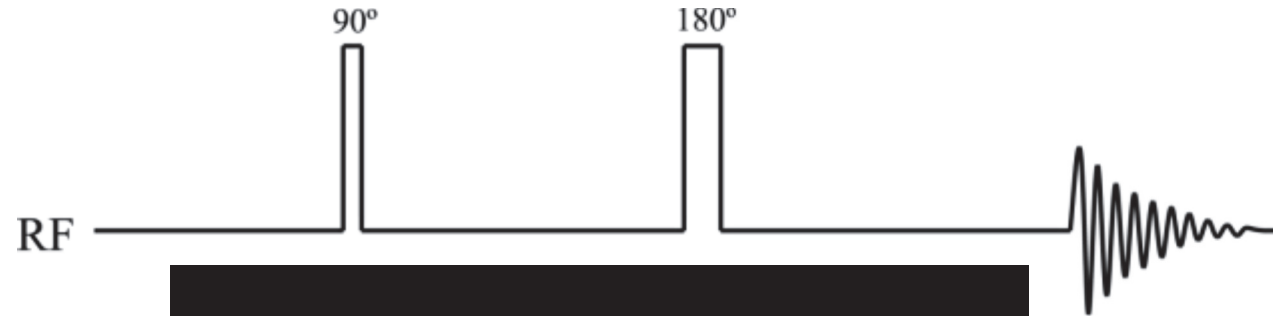
R. Evans et al, *Anal. Chem.* (2018), 90 (6), 3987–3994

R. Evans, *Prog Nucl Magn Reson Spectrosc* (2020), 117, 33-69

<https://www.nmr.chemistry.manchester.ac.uk/?q=node/432>

# Spin echo

Signal is refocused...



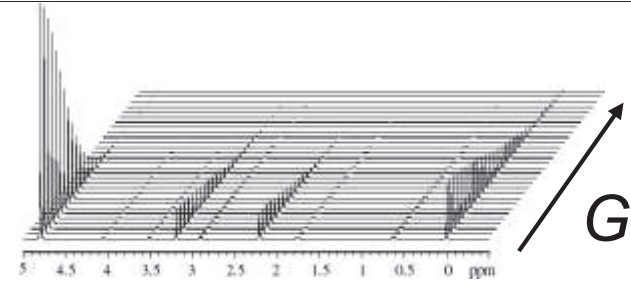
as droplets in corn syrup



<https://twitter.com/wonderofscience/status/1320692387925032961>

# Pulsed field gradient spin echo

- Application of PFG will result in a diffusivity dependent attenuation



$$S = S_0 e^{-D\gamma^2\delta^2 G^2 \Delta'}$$

$S$  : signal amplitude

$S_0$ : signal amplitude without diffusion

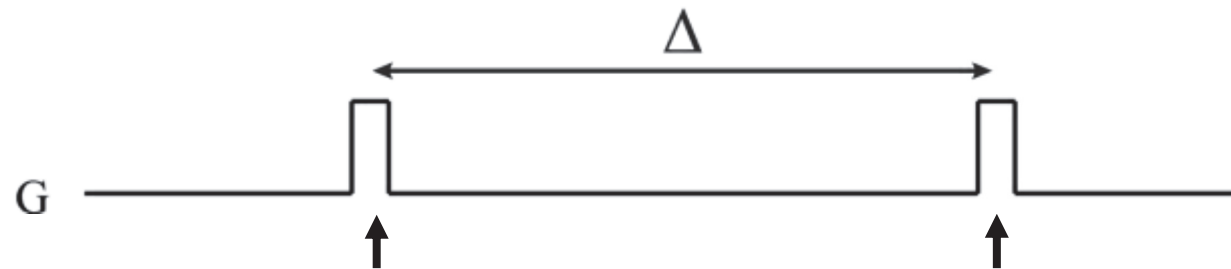
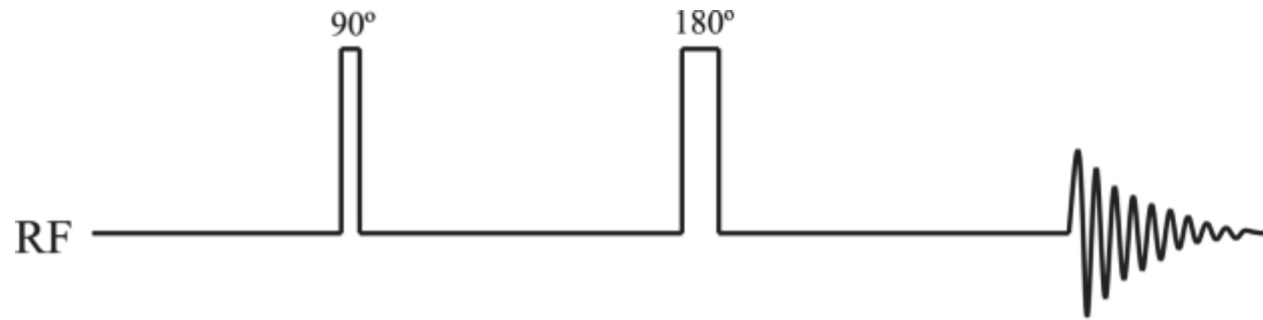
$D$  : diffusion coefficient

$\gamma$  : gyromagnetic ratio

$\delta$  : gradient pulse width

$G$  : gradient amplitude

$\Delta'$  : corrected diffusion time



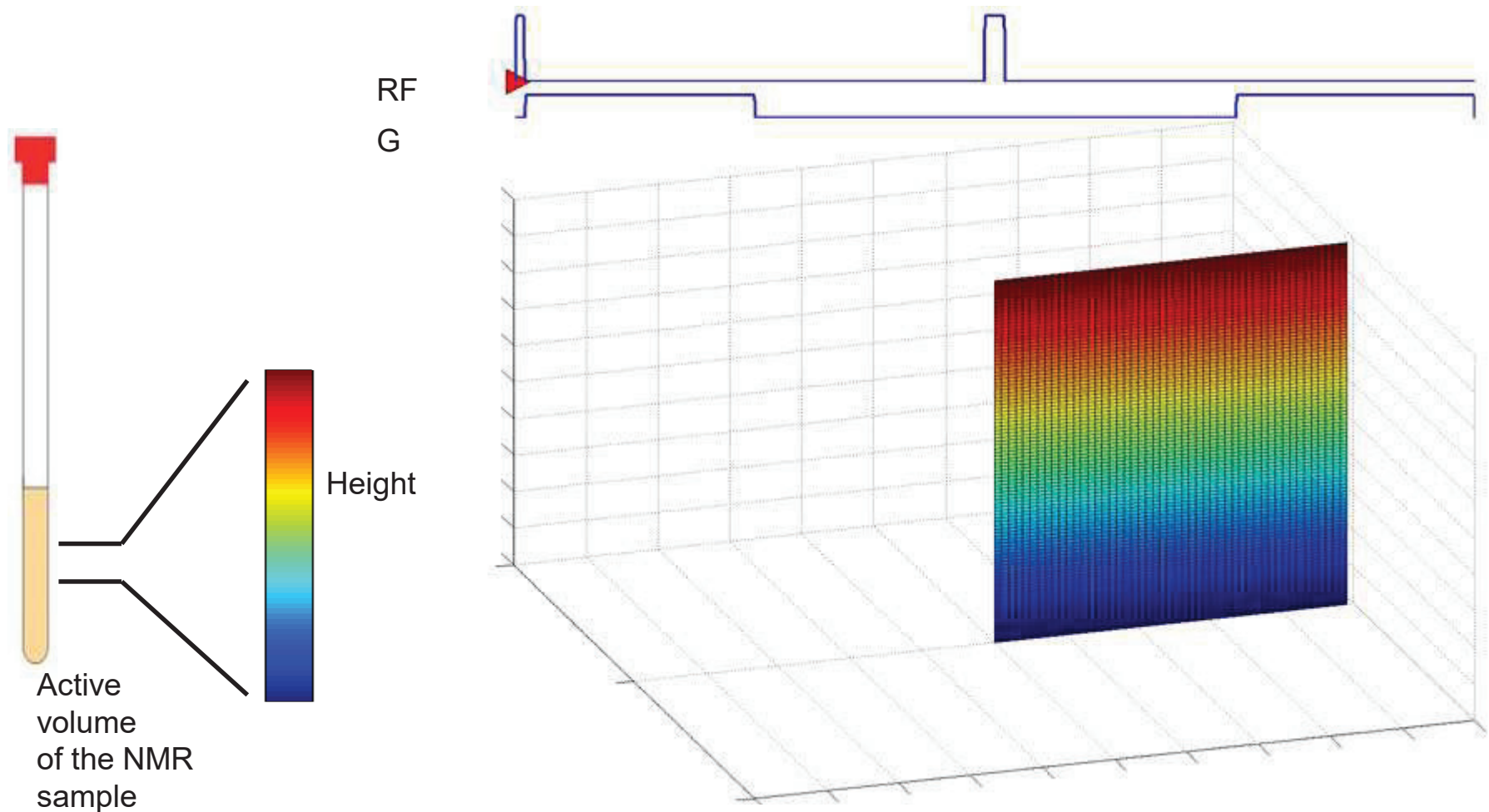
spin phases scrambled  
[  $\phi = \phi(z)$  ]

phases unscrambled  
[  $\phi = \phi(z) - \phi(z) = 0$  ]  
if spins haven't moved



# Pulsed field gradient spin echo

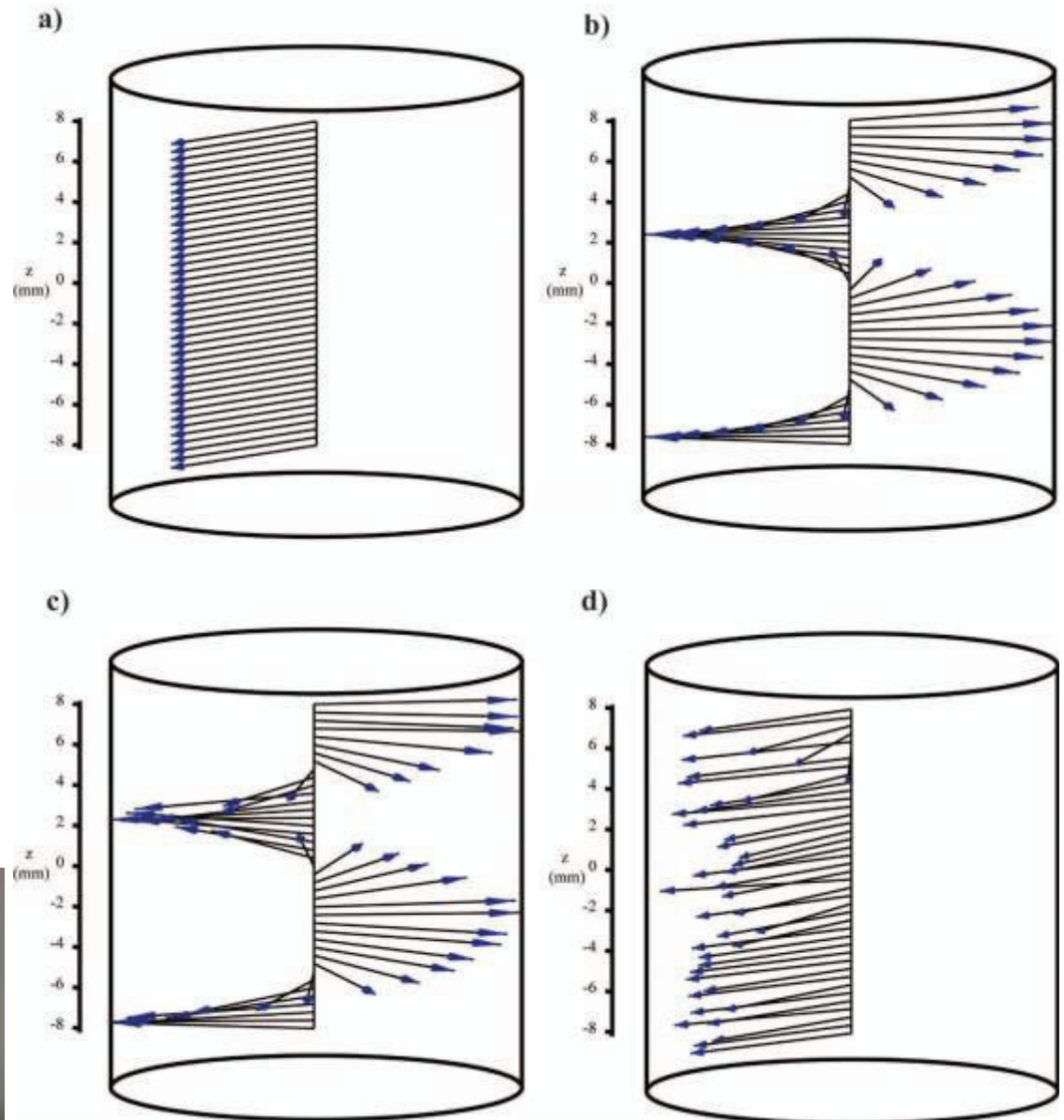
- Magnetization evolution without diffusion



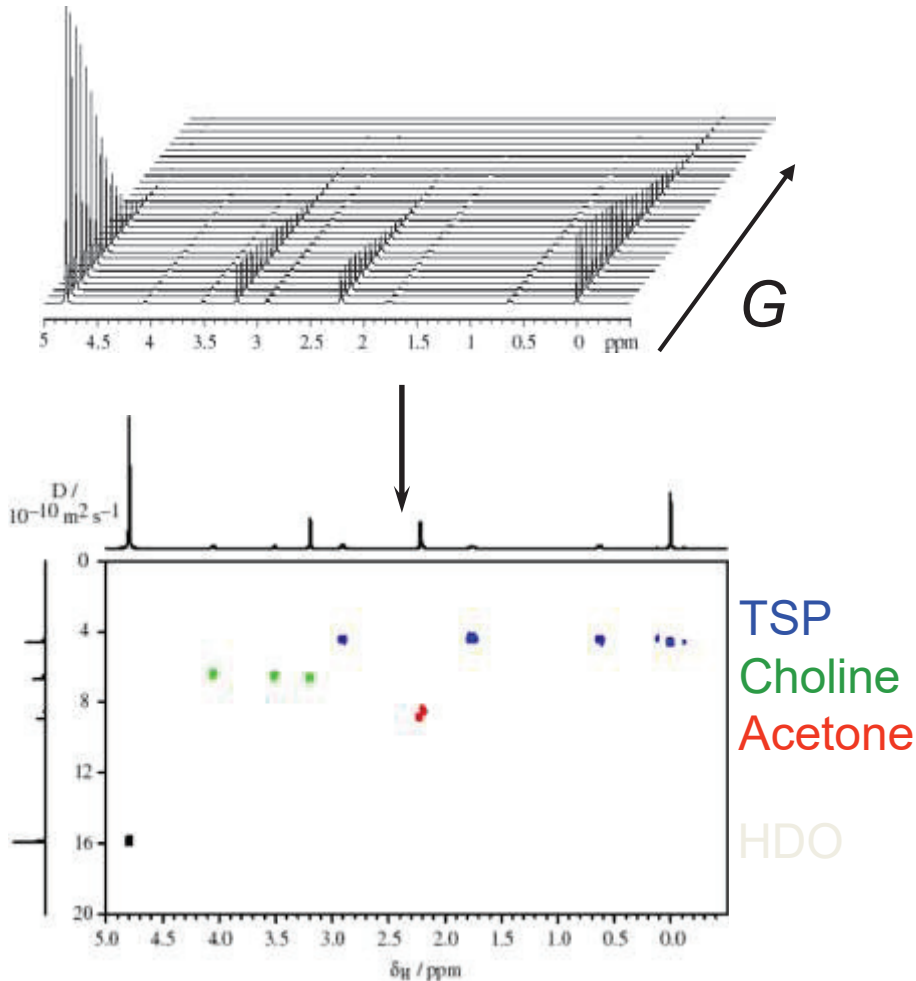
# Pulsed field gradient spin echo

- Magnetization evolution with diffusion
- Diffusion results in imperfect refocusing of magnetization.
- More diffusion leads to worse refocusing, and thus more signal attenuation

No diffusion vs diffusion



# Diffusion-Ordered Spectroscopy (DOSY)



- Measure spectra as a function of  $G$
  - Fit peak heights to get diffusion coefficients  $D$
- $$S = S_0 e^{-D\gamma^2\delta^2 G^2 \Delta'}$$
- Extend 1D peaks into a second dimension, with Gaussian shapes centred on the  $D$ 's
  - Widths determined by the standard errors  $\sigma_D$

Virtual chromatography

Despite the name, DOSY is not like COSY/NOESY: spectra are statistical constructs from, not transforms of, experimental data

# Practical consequences of the Stejskal-Tanner formula

Stejskal-Tanner formula for signal intensity:

$$S = S_0 e^{-D\gamma^2\delta^2 G^2 \Delta'}$$

-  $(\gamma \delta G_{zi})^2$  - **gradient area**

nuclei with high  $\gamma$  values are more sensitive for diffusion ( $^1\text{H}$ ,  $^{19}\text{F}$ ,  $^{31}\text{P}$ )  
(i.e.  $^1\text{H}$  is 16 times more sensitive than  $^{13}\text{C}$ )

-  $\delta$  **should be kept short**

during  $\delta$  the magnetization is transverse, homonuclear J-couplings evolve

-  $G$  **the more, the better**

provided the gradient hardware allows it

-  $\Delta$  **should be kept short:**

to minimize convection effects

# Why do we lose signal intensity?

## 1. Instrumental imperfections

gradient system

recovery - Eddy currents with multiple time constants

phase and lineshape distortions

gradient linearity and symmetry

gradient reproducibility

noise produced by the gradient amplifier

Use suitable pulse sequence

## 2. $T_1$ or $T_2$ relaxation Minimize pulse sequence delays

## 3. Diffusion - the physical basis of DOSY

## 4. Convection - the enemy of every diffusion experiment!

Minimize sample volume

Increase sample viscosity

Minimize temperature gradients

Use convection compensated sequences

# PFG Spin echo

Field gradient pulses result in signal attenuation

$$S = S_0 e^{-D\gamma^2\delta^2 G^2 \Delta'}$$

$S$  : signal amplitude

$S_0$ : signal amplitude  
without diffusion

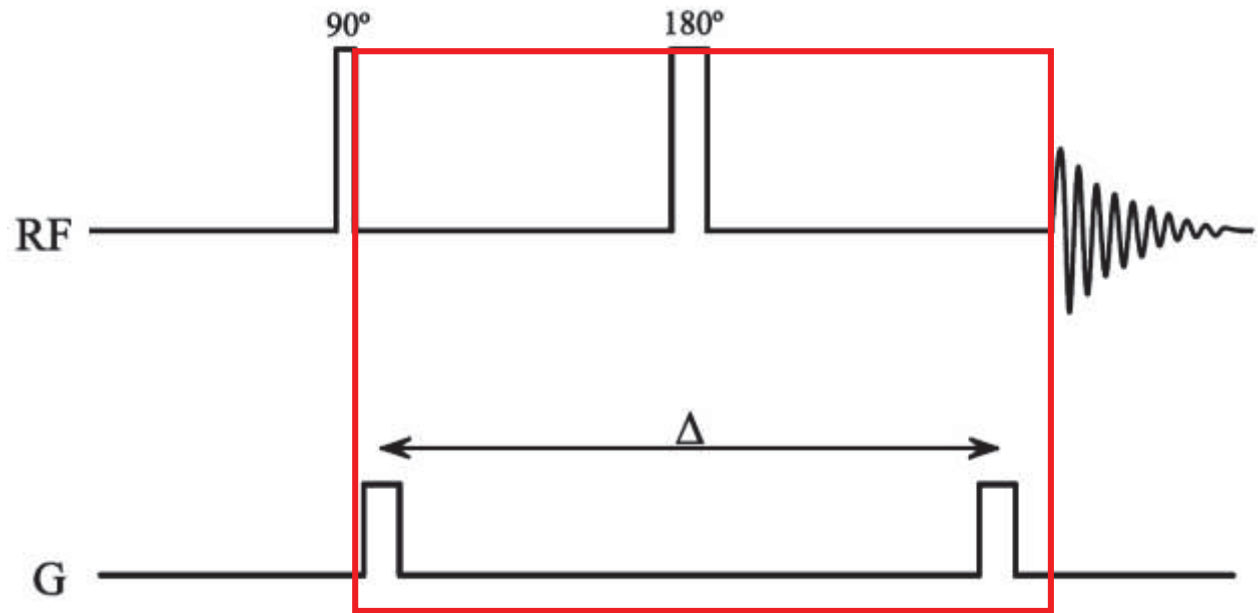
$D$  : diffusion coefficient

$\gamma$  : gyromagnetic ratio

$\delta$  : gradient pulse width

$G$  : gradient amplitude

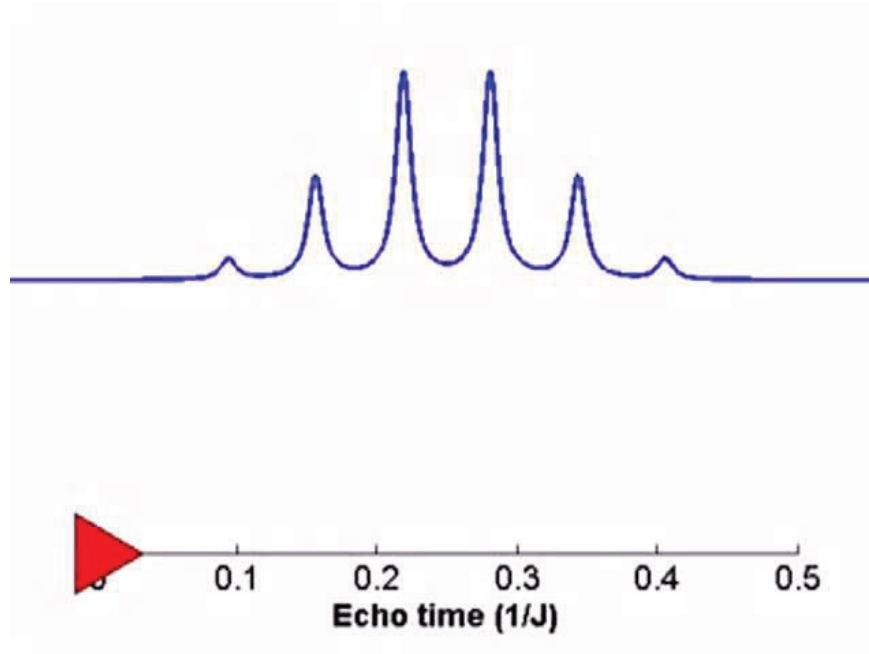
$\Delta'$  : corrected diffusion time



↑  
spin phases scrambled  
[  $\Phi = \Phi(z)$  ]

↑  
phases unscrambled  
[  $\Phi = \Phi(z) - \Phi(z) = 0$  ]  
if spins haven't moved

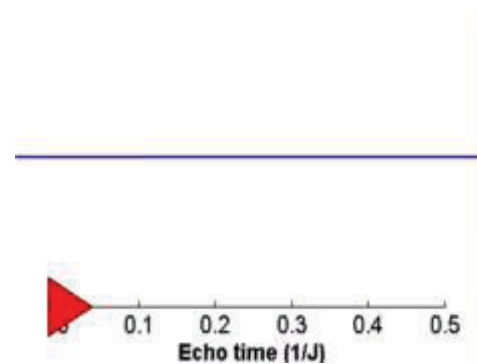
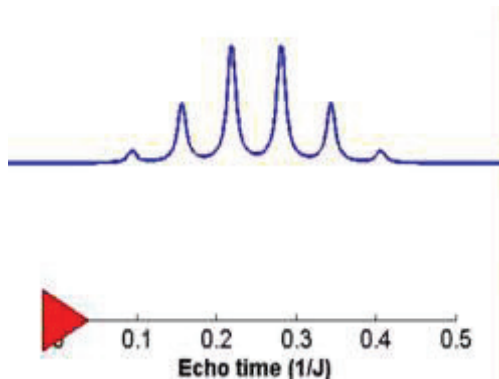
# J-modulated sextet



In-phase magnetization

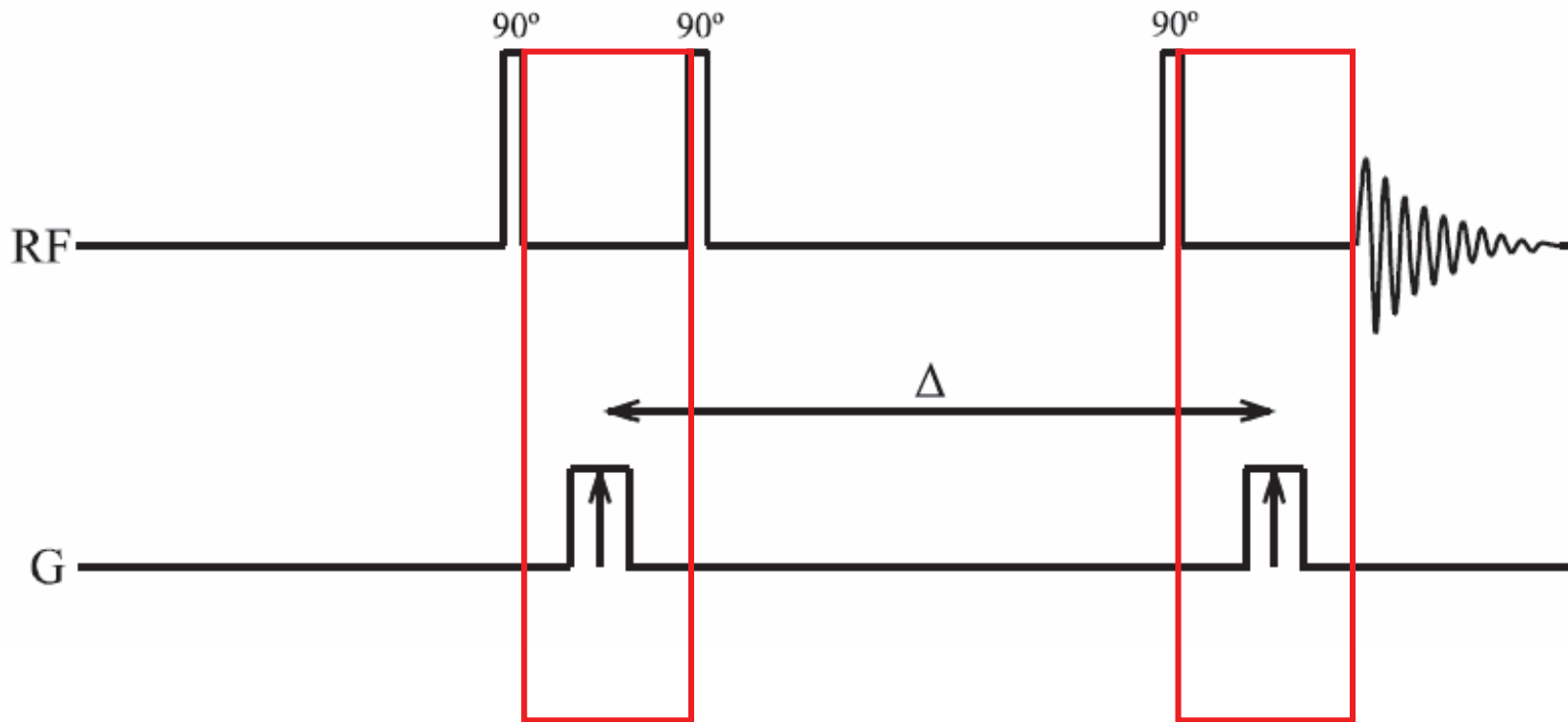
+

Anti-phase magnetization



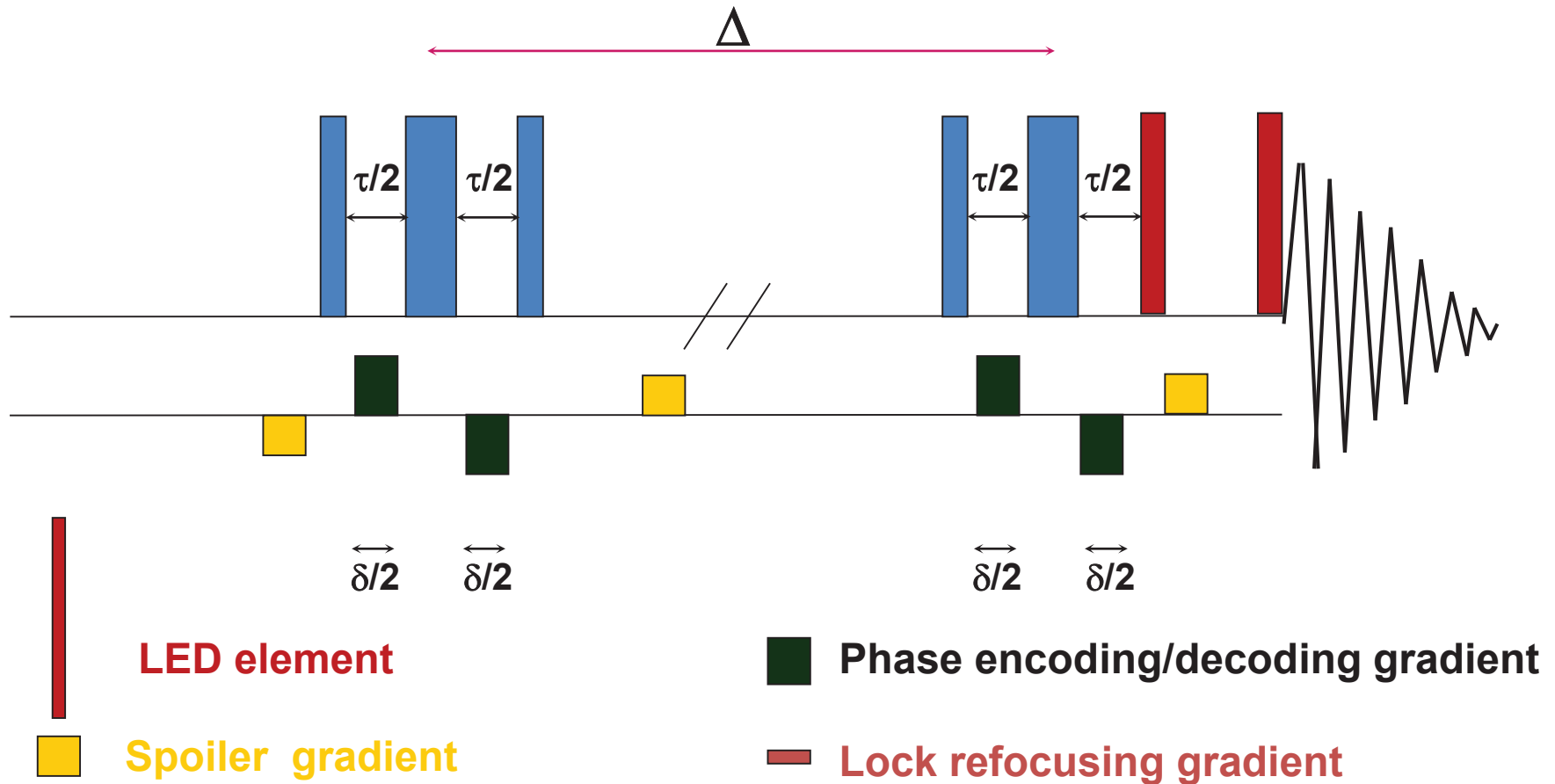
# PFG Stimulated echo

- Magnetization is stored along the z-axis for most of  $\Delta$
- Reduced J-modulation
- Lose 50% of magnetization

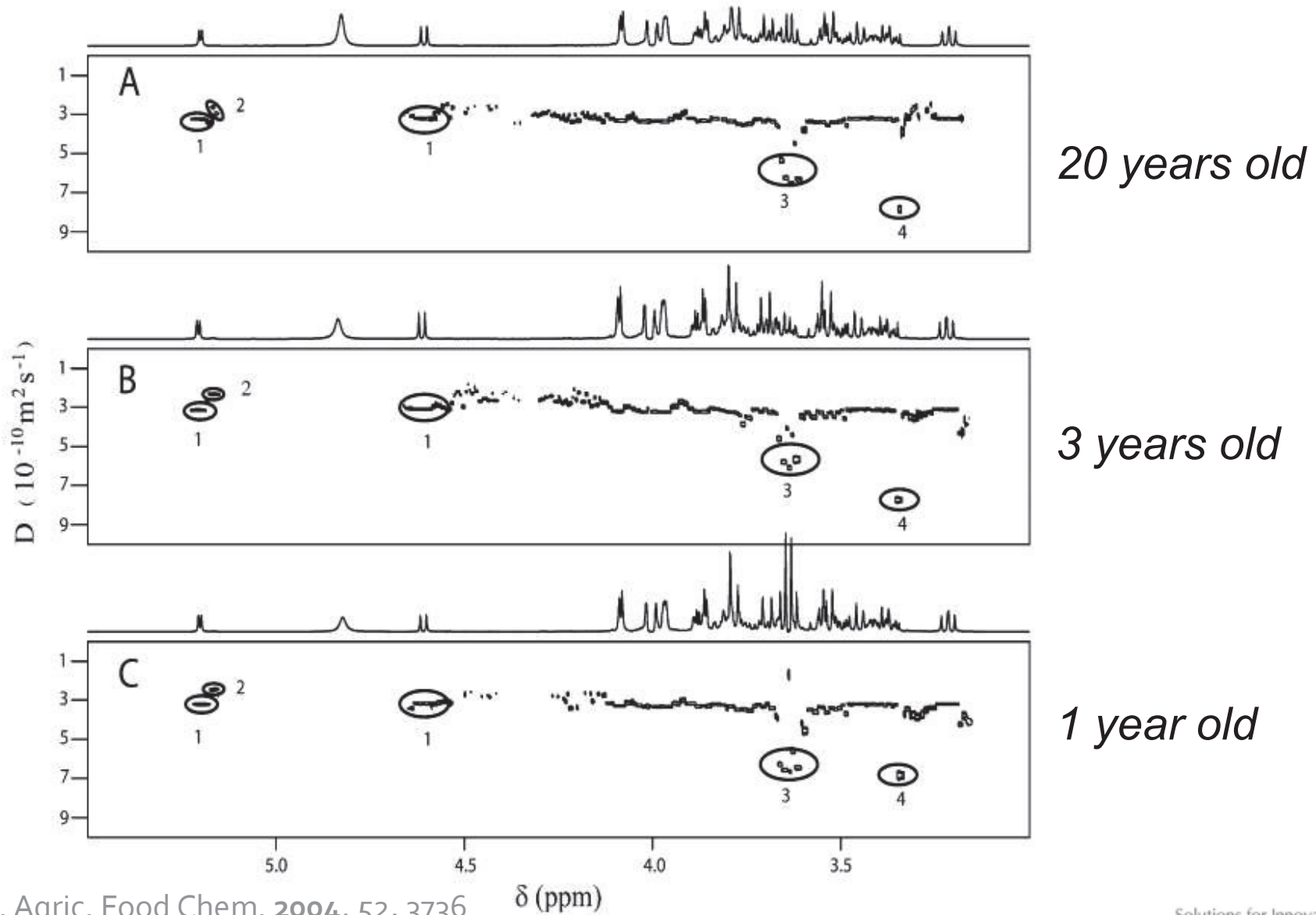




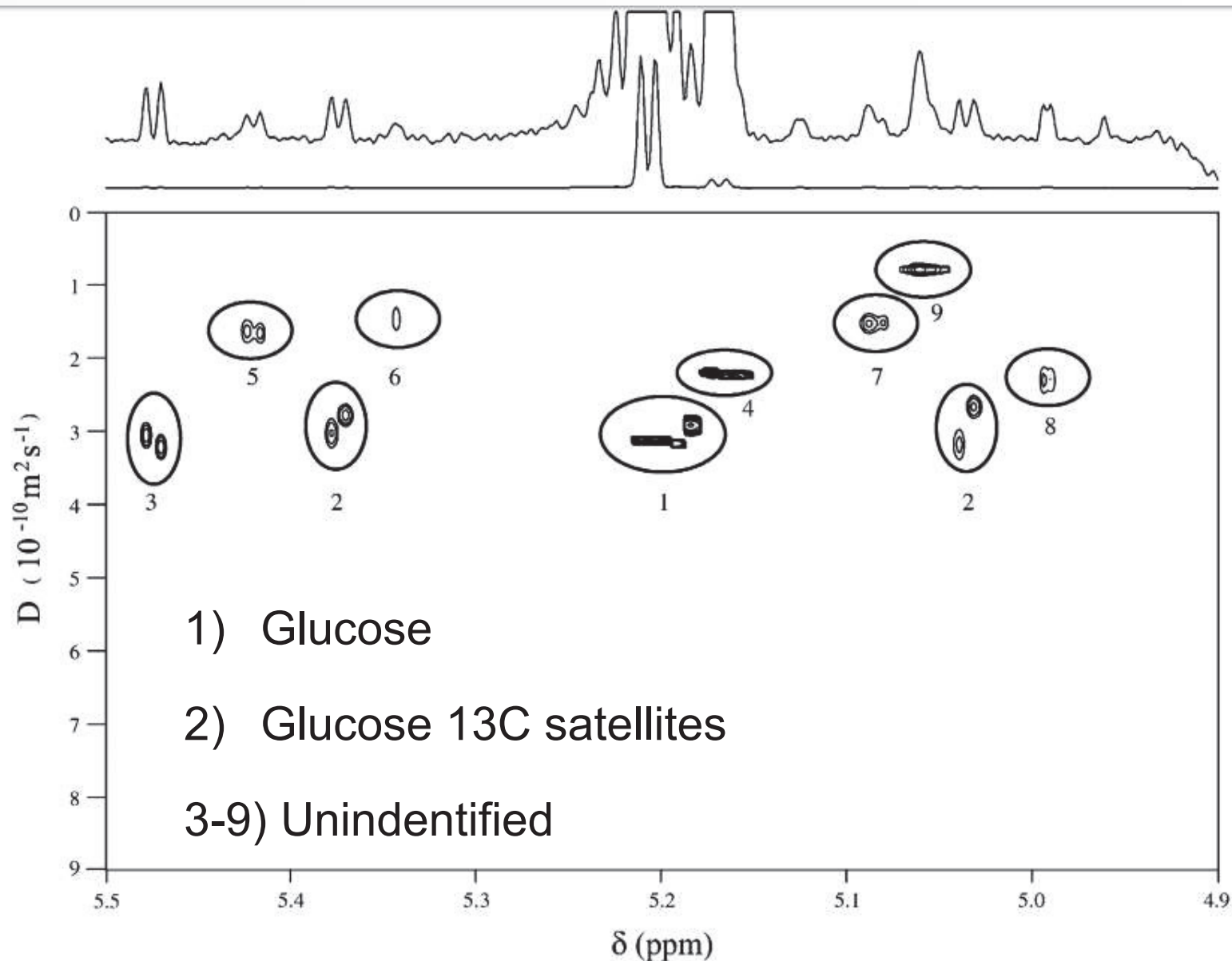
# In practice: Bpp\_led\_dosy\_pfg.jxp



# DOSY of 3 port wines



# DOSY expansion of 3 years old port wine



# Convection

- Once a critical Raleigh number is reached (1700 for a Benard configuration, i.e., two parallel horizontal boundaries separated by a distance  $d$ ), natural convection appears.

$$Ra = \frac{\rho\beta\Delta T l^3 g}{\eta\alpha}$$

$g$ : gravity,

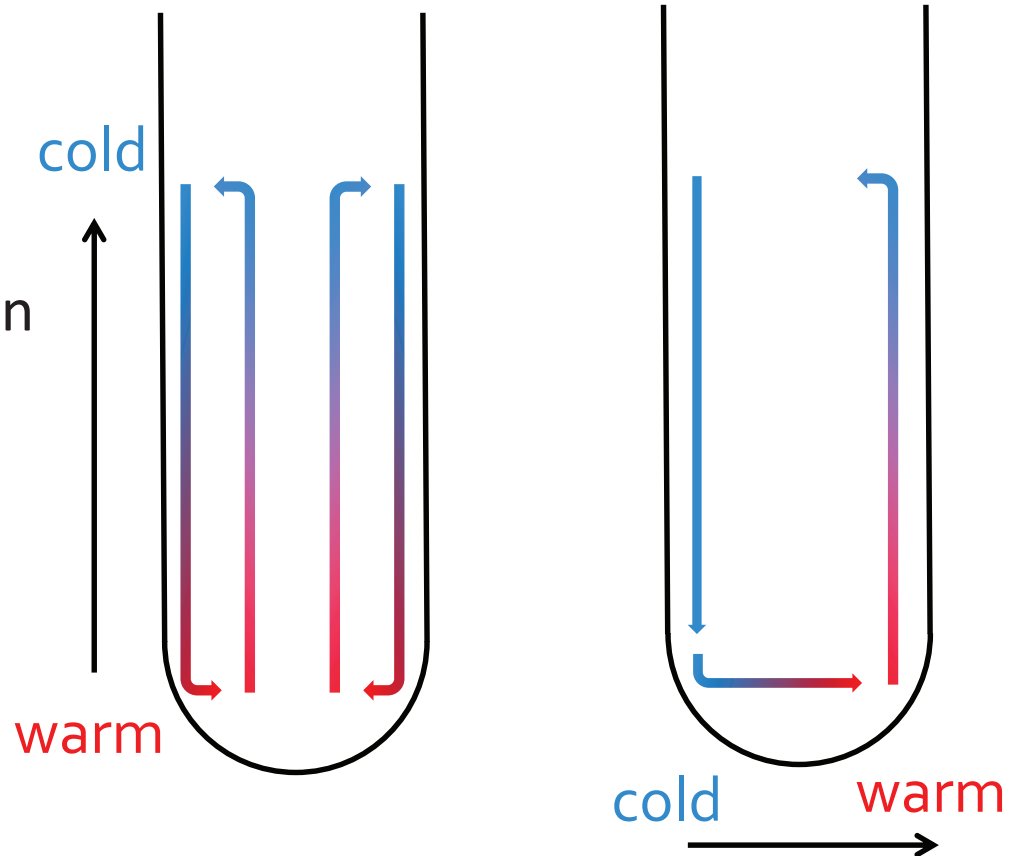
$\beta$ : coefficient of thermal expansion

$\Delta T$ : temperature difference

$l$ : length

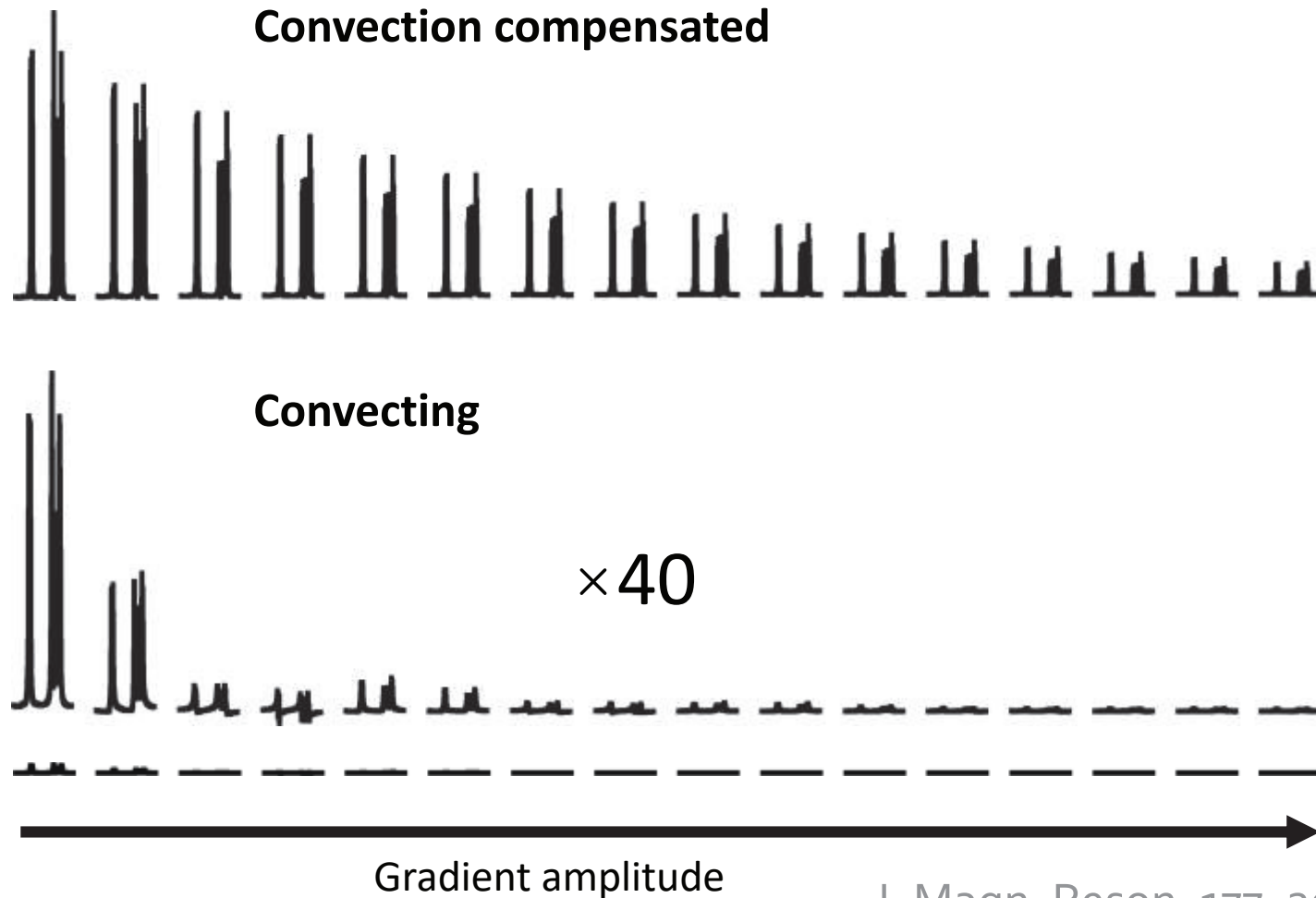
$\eta$ : viscosity

$\alpha$ : thermal diffusivity



# Convection in an NMR tube

Aromatic signals from quinine (7.1 to 7.6 ppm ) as a function of increasing gradient strength at 25 °C.



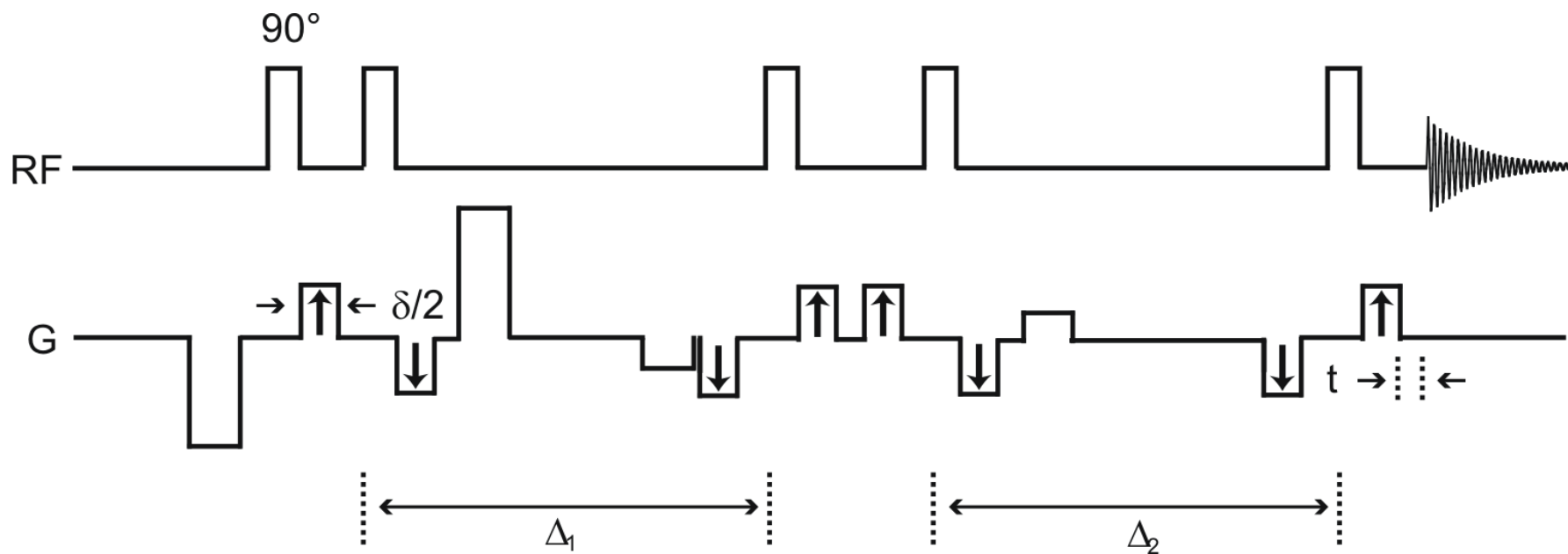
# Convection in NMR

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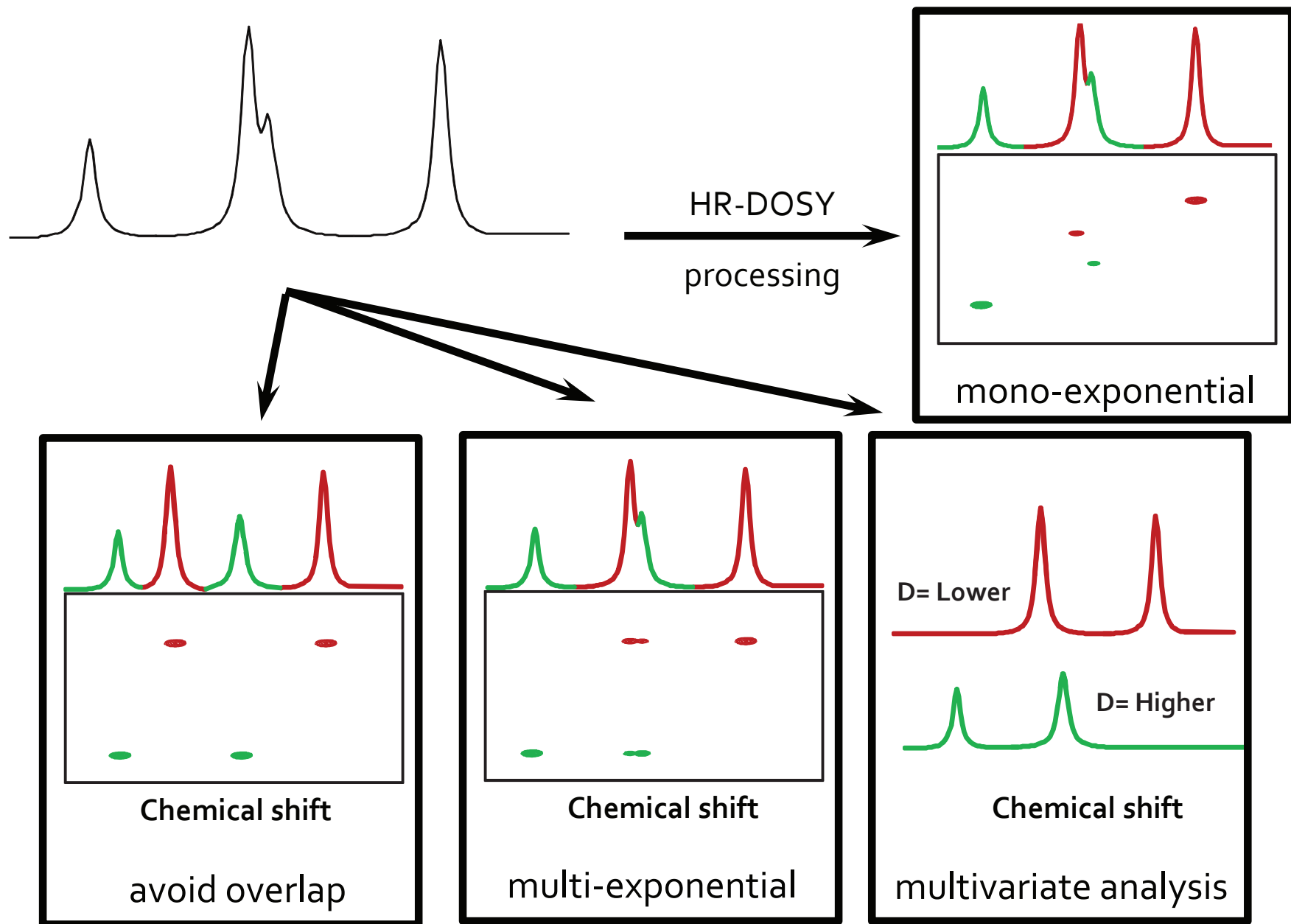
- Any pulse sequence with encoding-decoding gradient pairs suffers from convection
  - Gradient shimming (ask for alternative)
  - Gradient selective 1D
  - g....jxp
  - ...

# Convection compensated diffusion measurement: bpp\_dste\_led\_dosy\_pfg.jxp

Double stimulated echo: lose 75% of signal and longer phase cycling

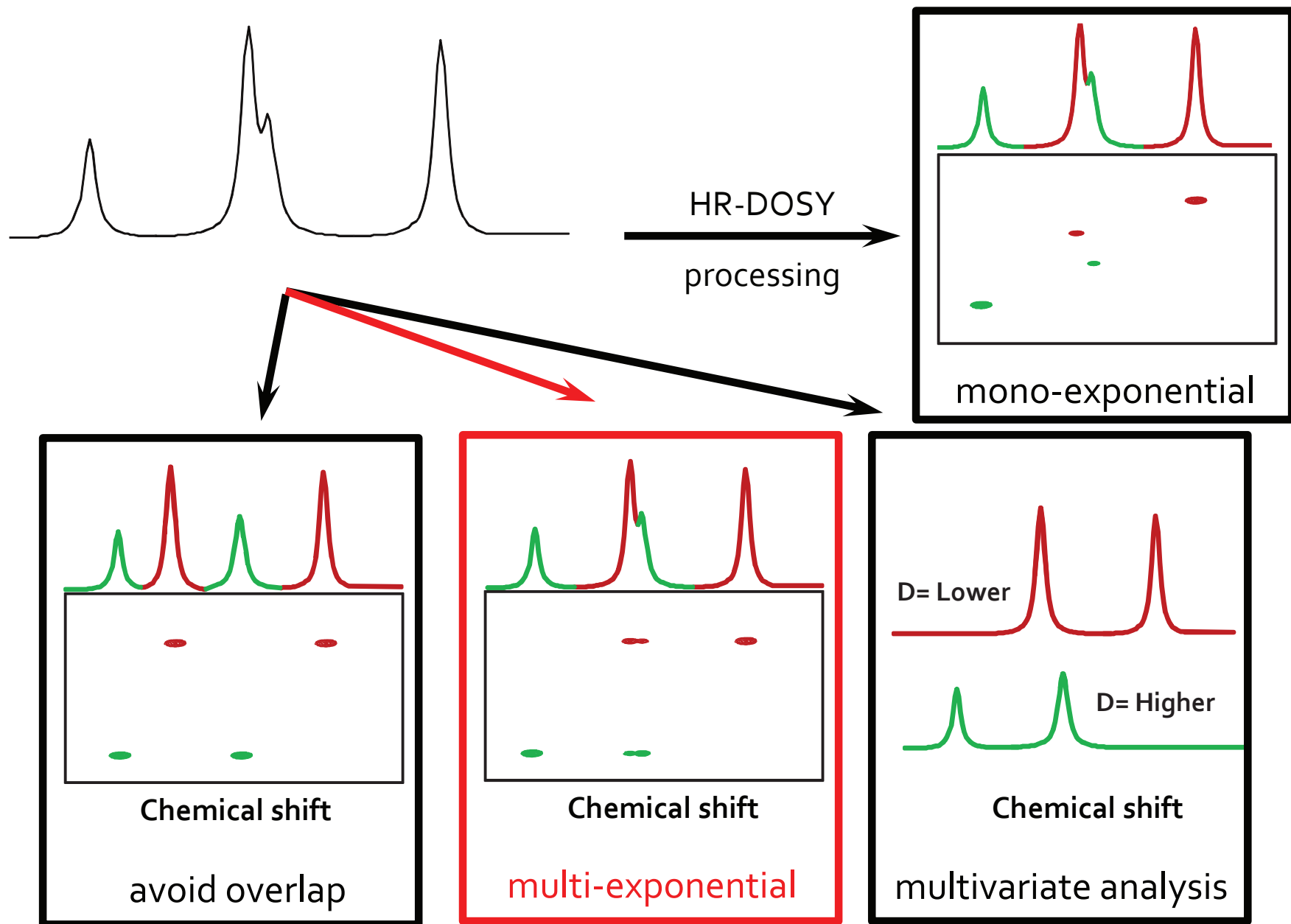


# signal overlap in DOSY processing





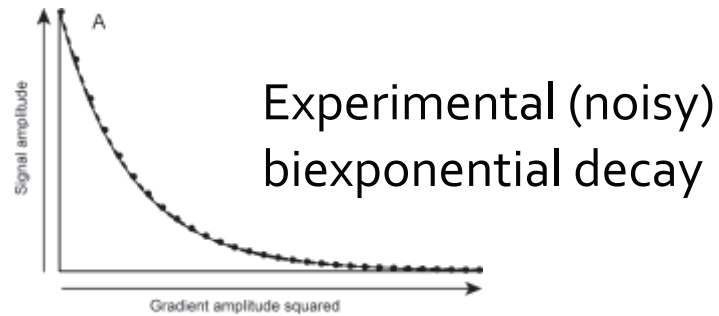
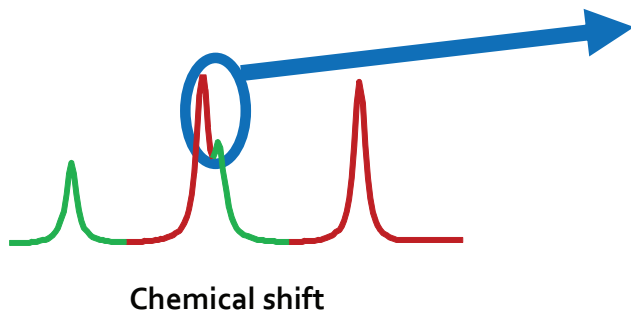
# signal overlap in DOSY processing



# Resolving superimposed exponentials

Superimposed exponentials is a very difficult mathematical problem (ill-posed and numerically unstable).

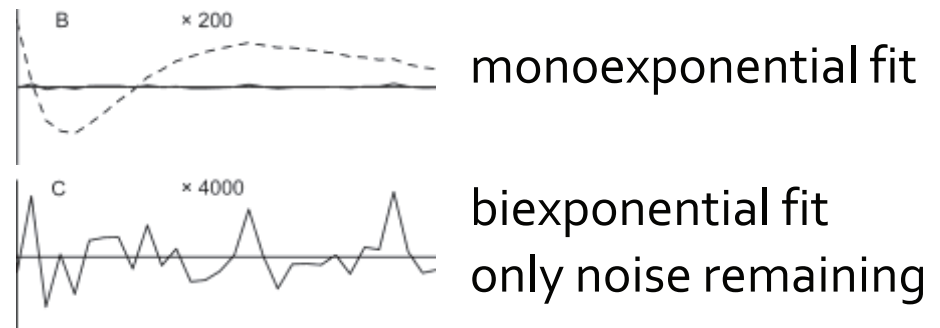
It is only practically feasible with high signal to noise ratio and for a limited (2-3) number of exponentials.



Residuals (**E**) are the fit (**F**) subtracted from the experimental data (**X**)

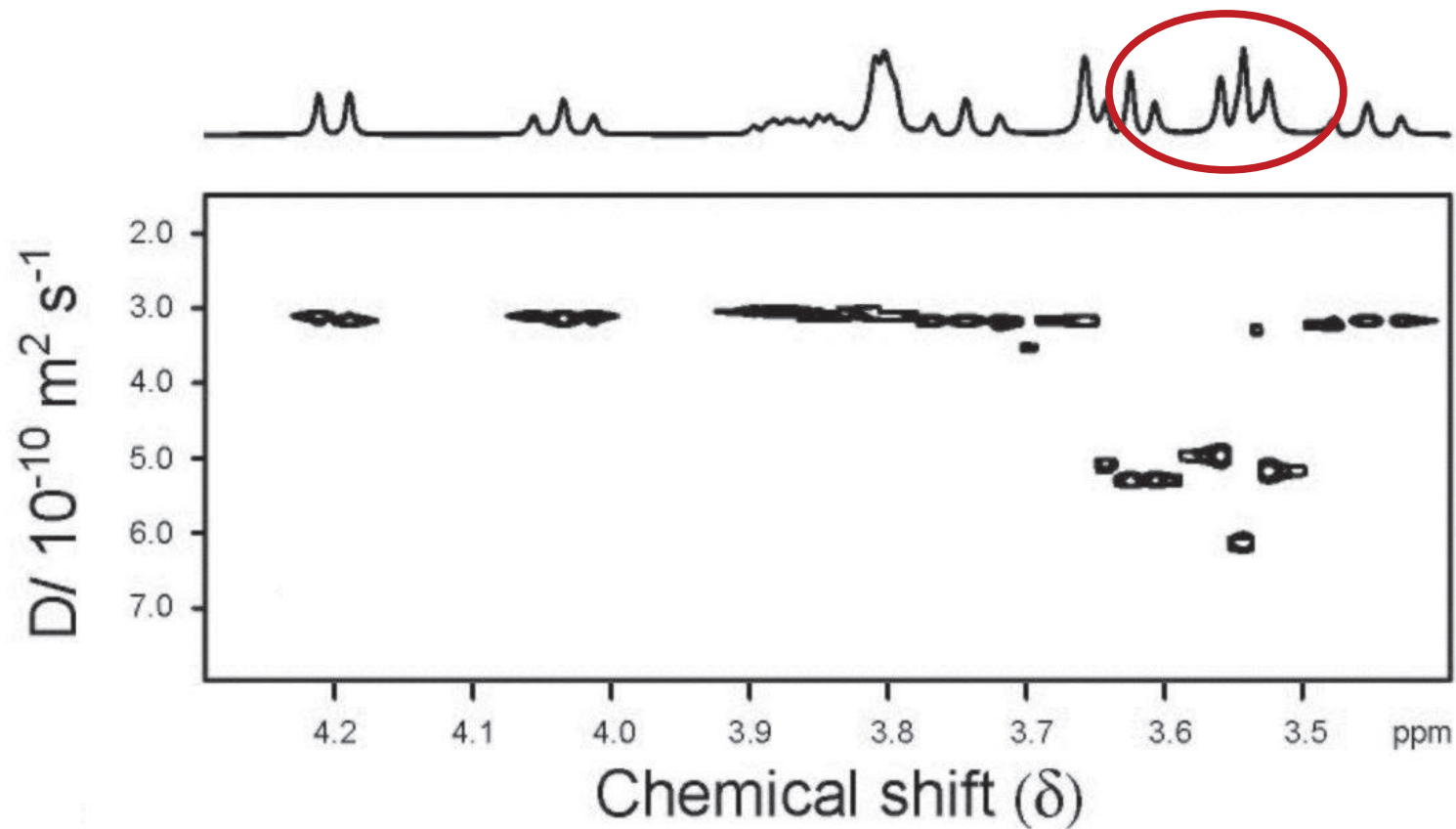
$$\mathbf{R} = \mathbf{X} - \mathbf{F}$$

Residuals:



# overlap in 2D DOSY: monoexponential fitting

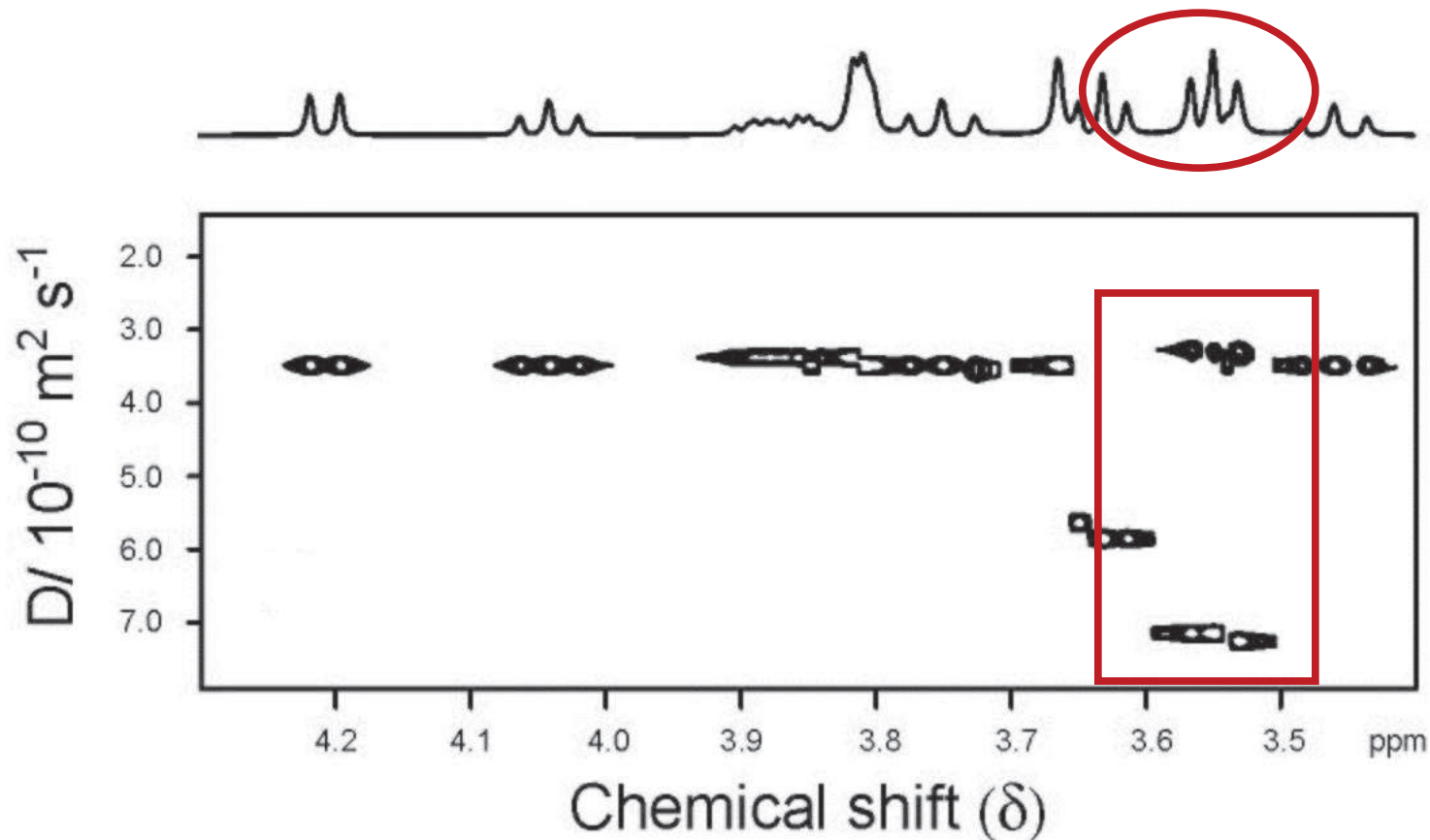
400 MHz oneshot spectrum of sucrose, isopentanol and propan-1-ol in D<sub>2</sub>O



Overlapping peaks give  
compromise diffusion coefficient

# overlap in 2D DOSY: biexponential fitting

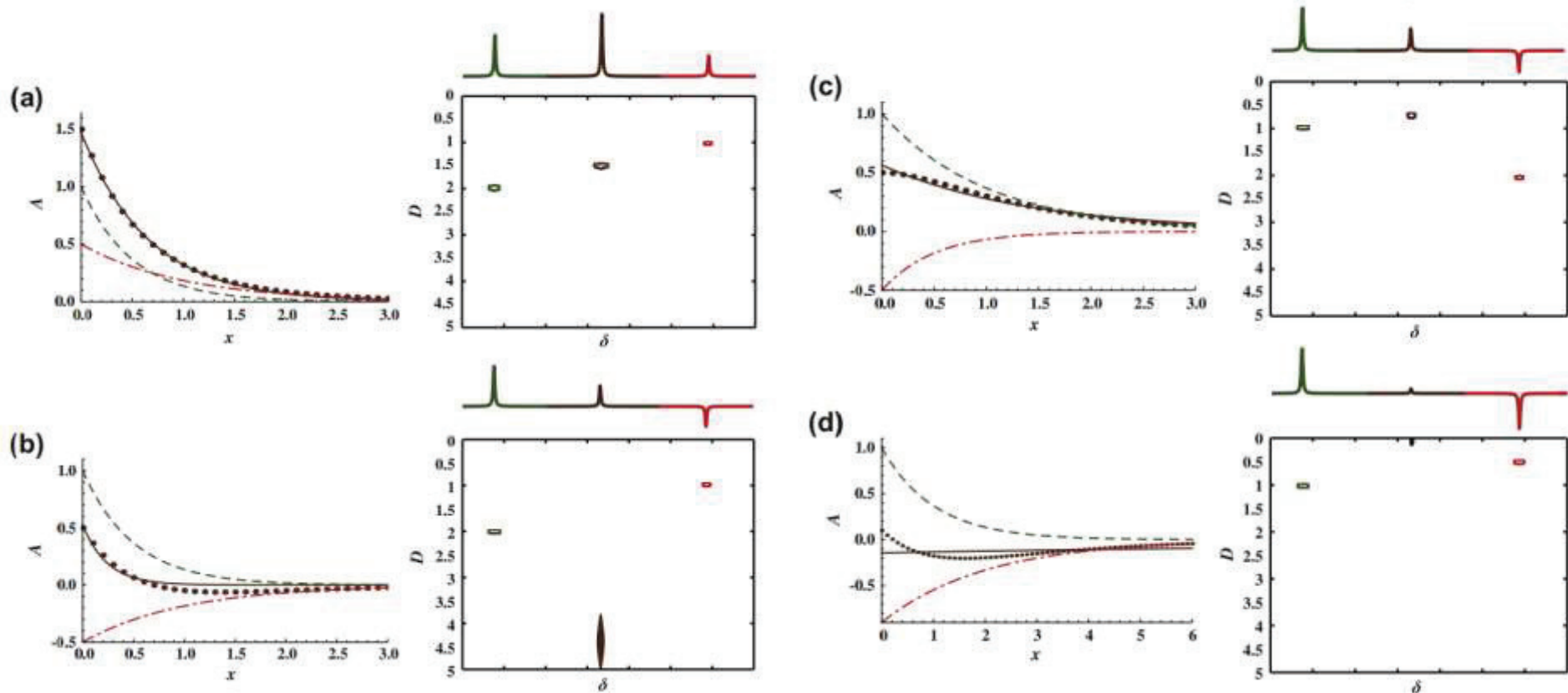
400 MHz oneshot spectrum of sucrose, isopentanol and propan-1-ol in  $D_2O$



$D_A$  and  $D_B$  must differ by at least 30%

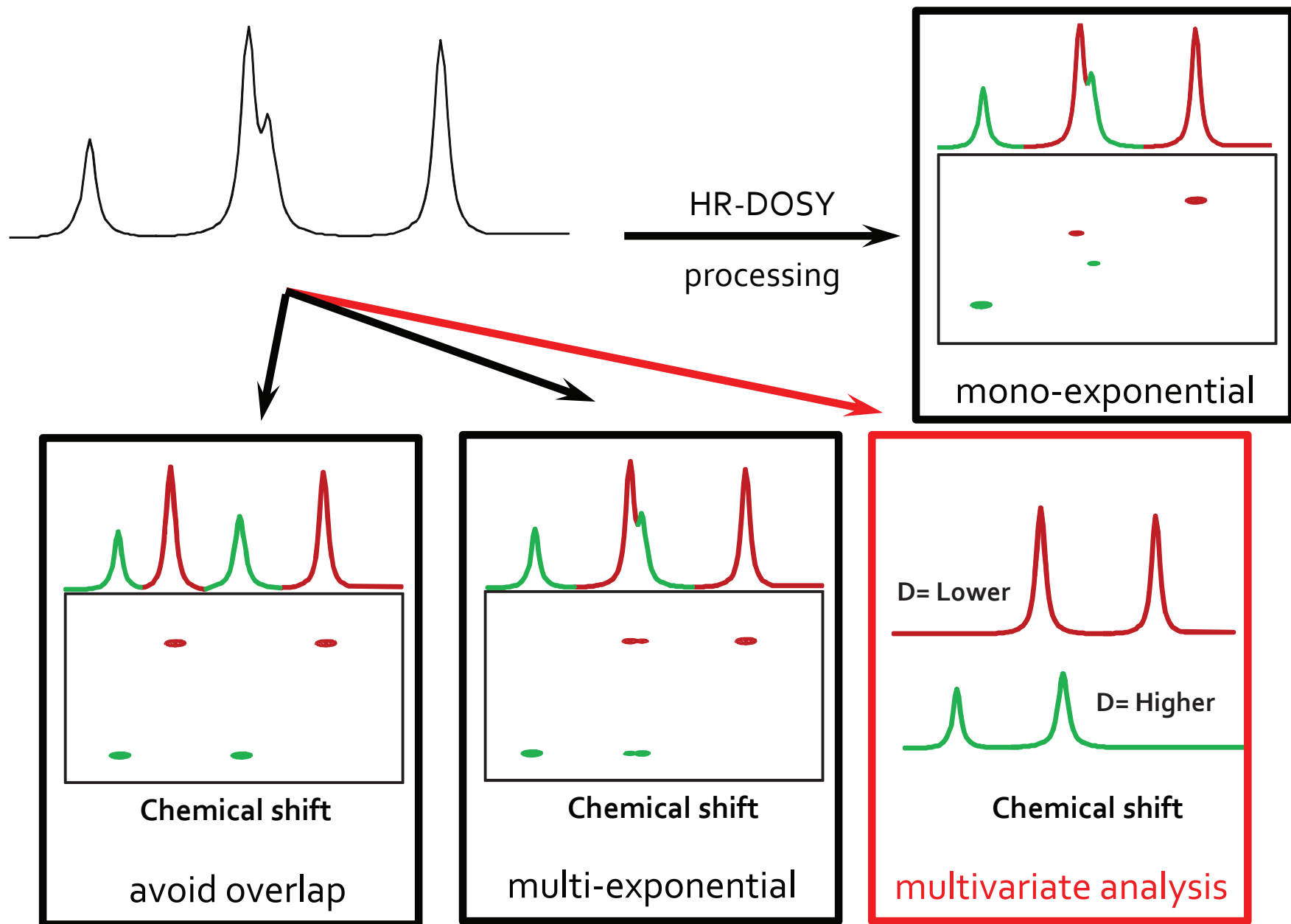
Very dependent on the quality and S/N of data

# Effects of overlap with J-modulation



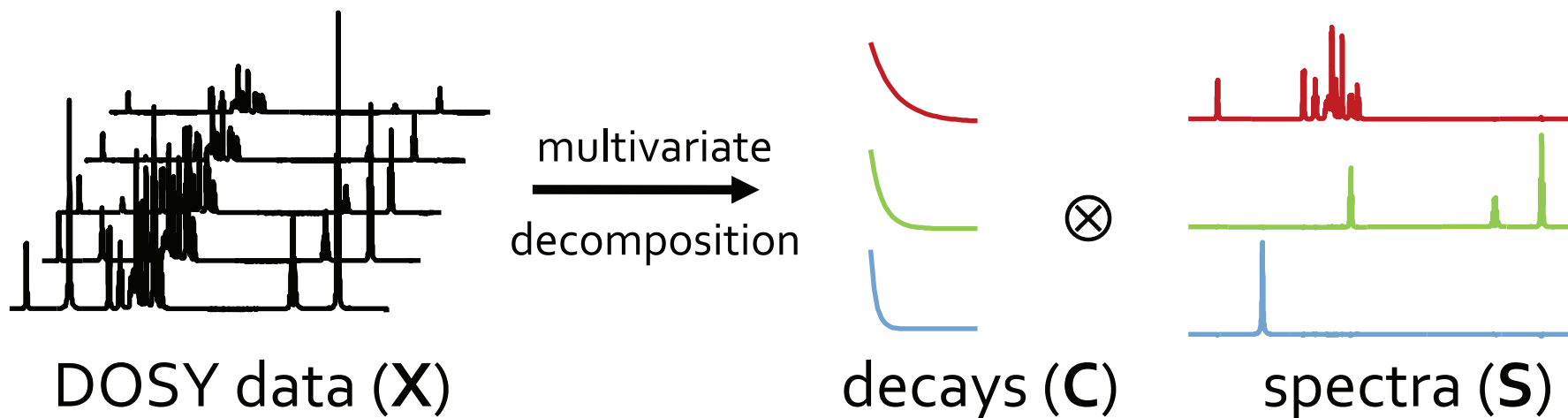
<https://doi.org/10.1016/j.jmr.2010.11.012>

# signal overlap in DOSY processing



# Processing “DOSY” data: SCORE/OUTSCORE

Here we fit the whole spectrum at once rather than each peak at the time as in HR-DOSY. This makes sense as all peaks in a component spectrum decays in the same way



$$\mathbf{X} = \mathbf{C} \mathbf{S}^T + \mathbf{E}$$

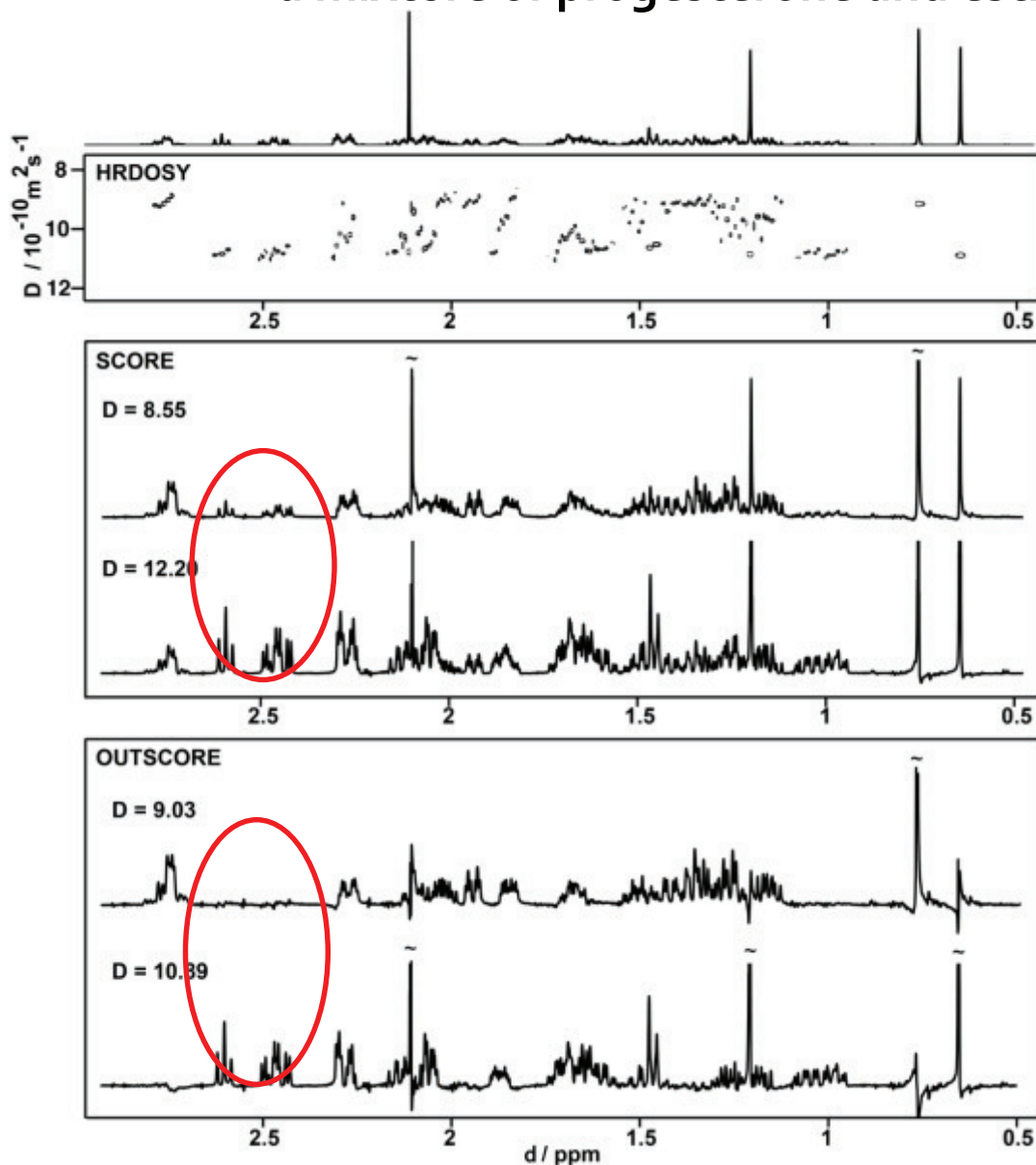
Minimize  $\mathbf{E}$  assuming a known decay form

**SCORE:**  $\mathbf{E} = \mathbf{X} - \mathbf{C} \mathbf{S}^T$  (residuals)

**OUTSCORE:**  $\mathbf{E} = |\mathbf{S}_i| \cdot |\mathbf{S}_j|$  (spectral similarity)

# Optimized Unmixing of True Spectra for COmponent Resolution (OUTSCORE)

a mixture of progesterone and estradiol in DMSO- $d_6$



## HRDOSY

monoexponential fitting  
<1% difference in  $D$   
suffers from overlap

## SCORE

minimizes residuals  
>30% difference in  $D$

## OUTSCORE

minimizes cross-talk  
<5% difference in  $D$   
fewer components



# Multivariate analysis

- Reference deconvolution to improve data:

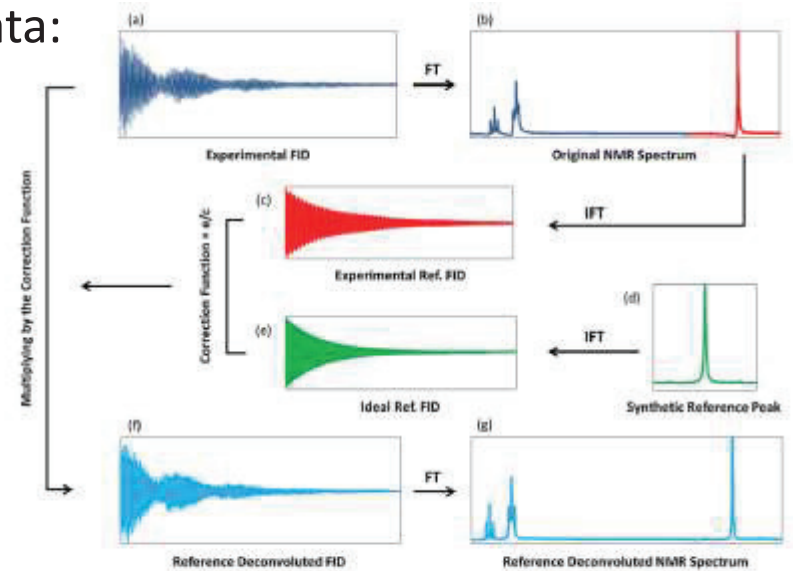
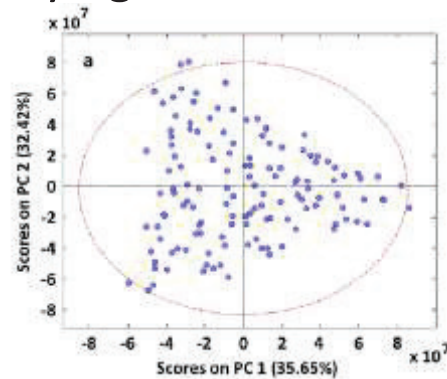
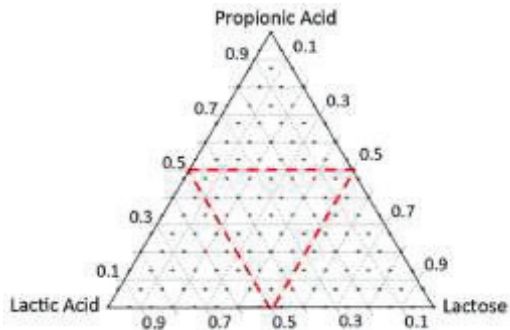
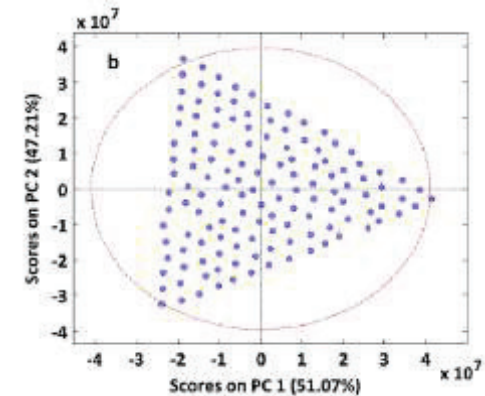


Figure 1. Schematic illustration of the FIDDLE algorithm for reference deconvolution. The reference peak is extracted from the experimental spectrum (b), and its inverse Fourier transform (c) is compared to that of "perfect" FID (e) to yield a correction function (e/c). The correction is then applied in the time domain to the entire experimental FID (a) to produce the corrected FID (f).

- Example (3 compounds with varying concentrations):



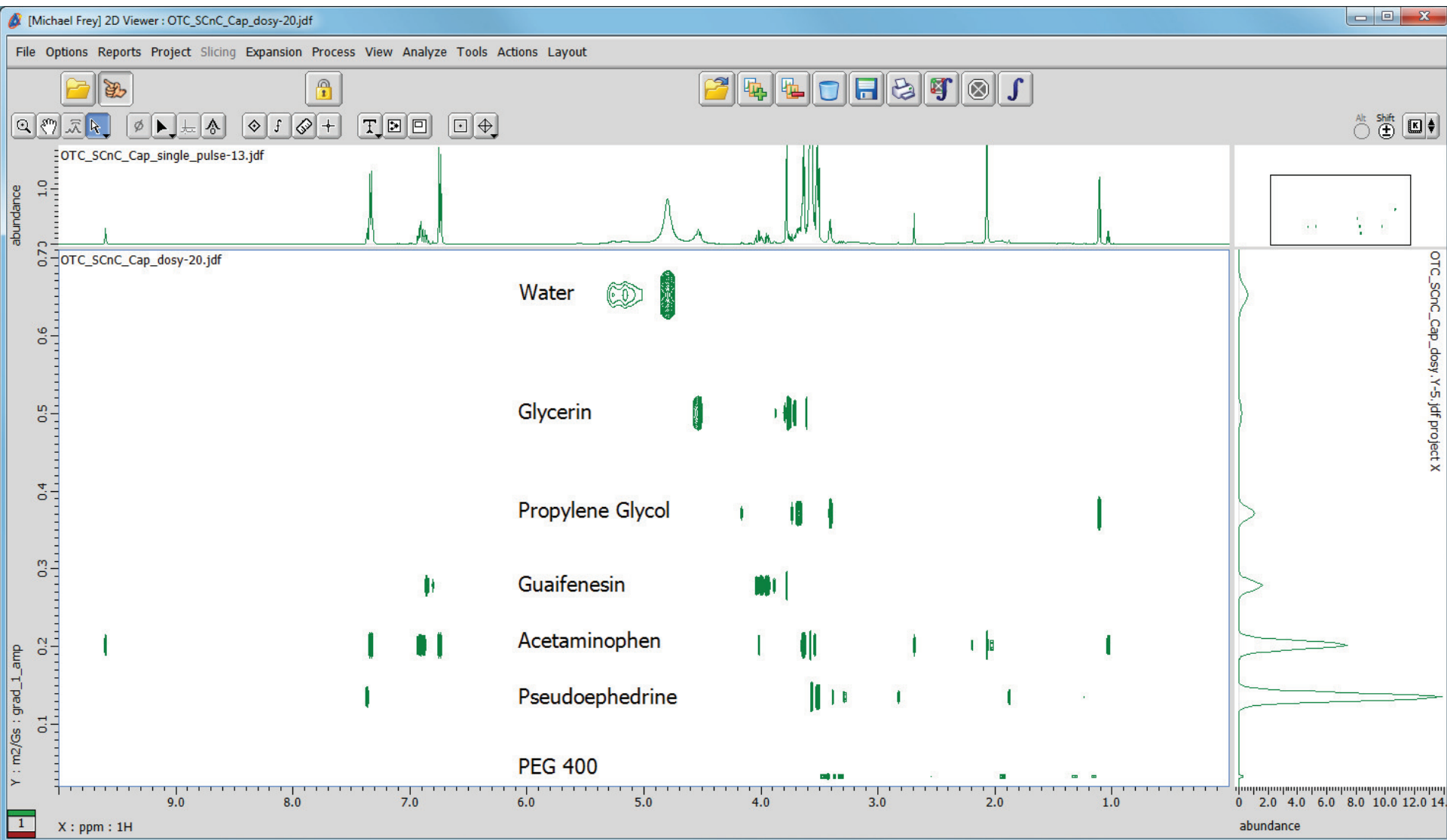
Raw data



Reference deconvoluted data

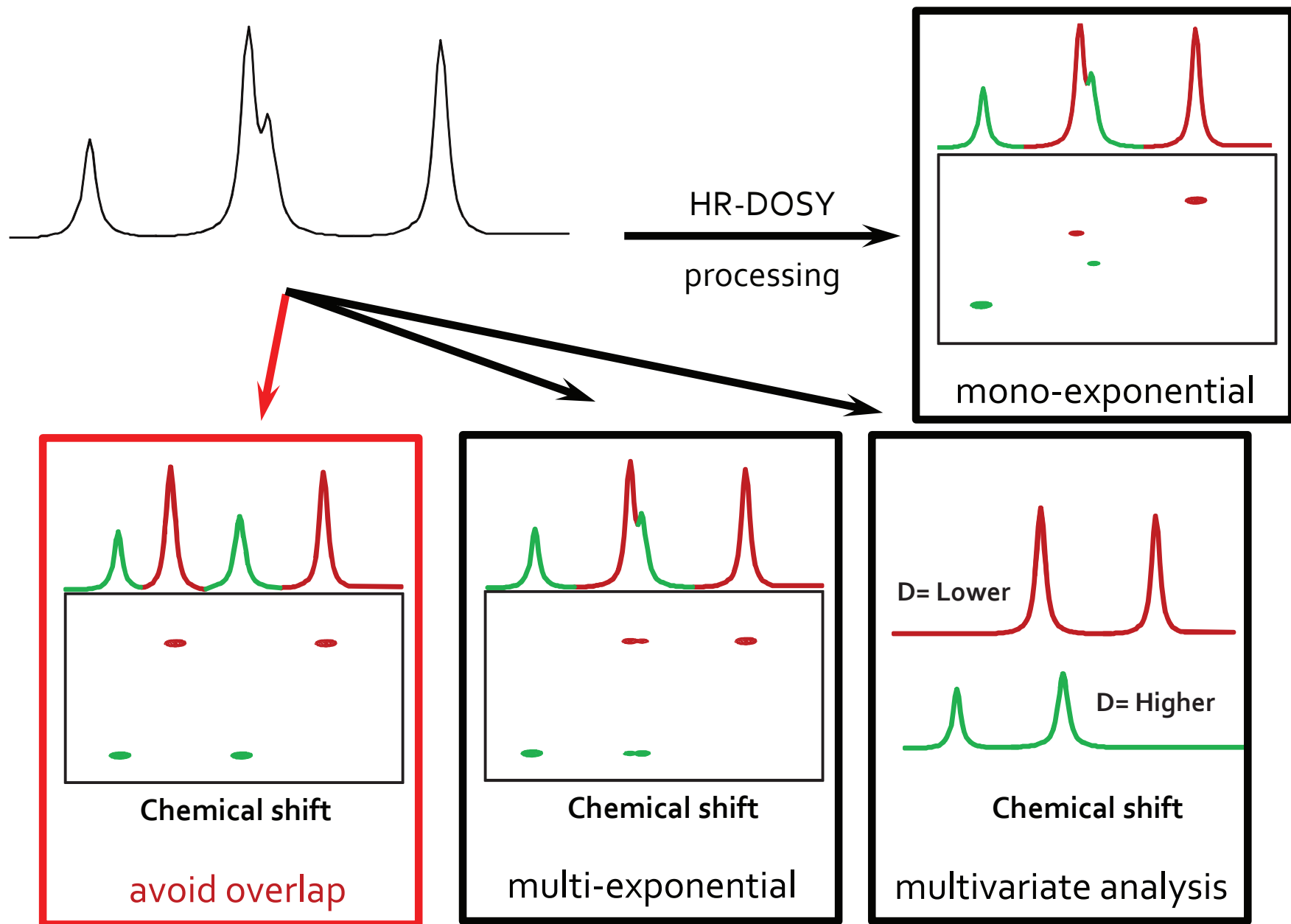
<https://doi.org/10.1002/cem.2607>

# Multivariate DOSY NMR of paracetamol tablet



Solvent Methanol-d<sub>4</sub>

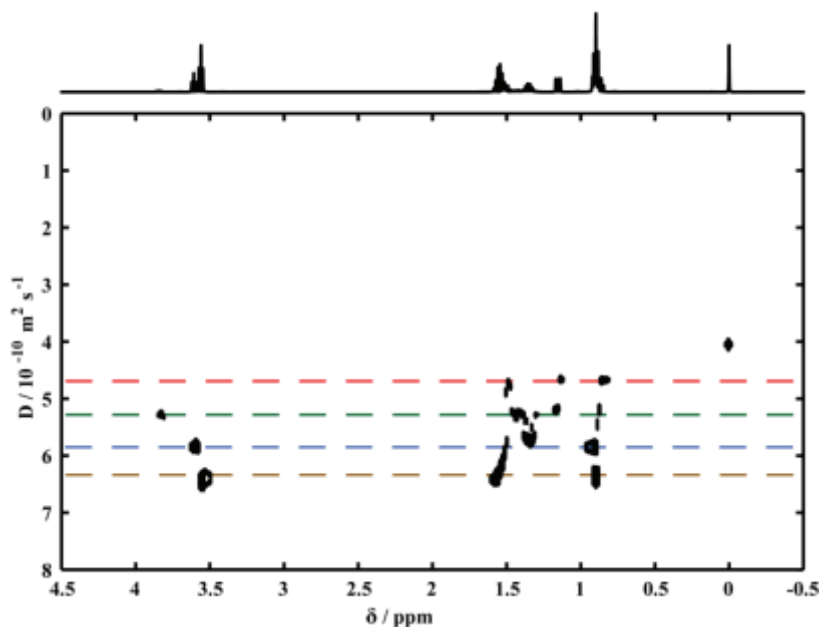
# signal overlap in DOSY processing



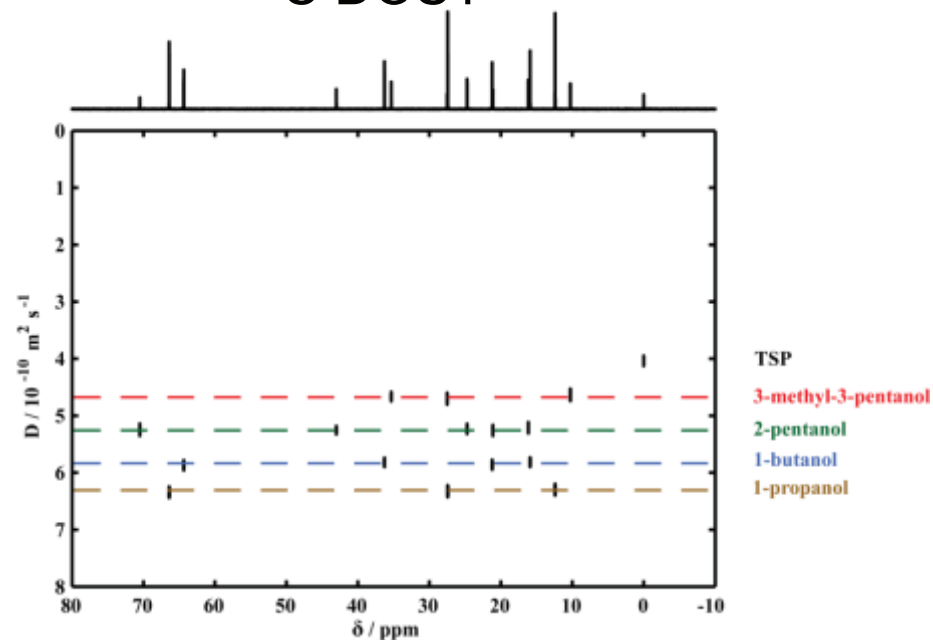
# overlap in 2D DOSY: $^{13}\text{C}$ DOSY

500 MHz  $^1\text{H}$  and  $^{13}\text{C}$  DOSY spectra of mixture of alcohols in  $\text{D}_2\text{O}$

$^1\text{H}$  DOSY



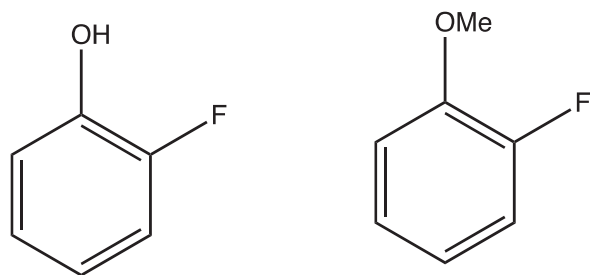
$^{13}\text{C}$  DOSY



No overlap in the  $^{13}\text{C}$  spectrum greatly facilitates interpretation

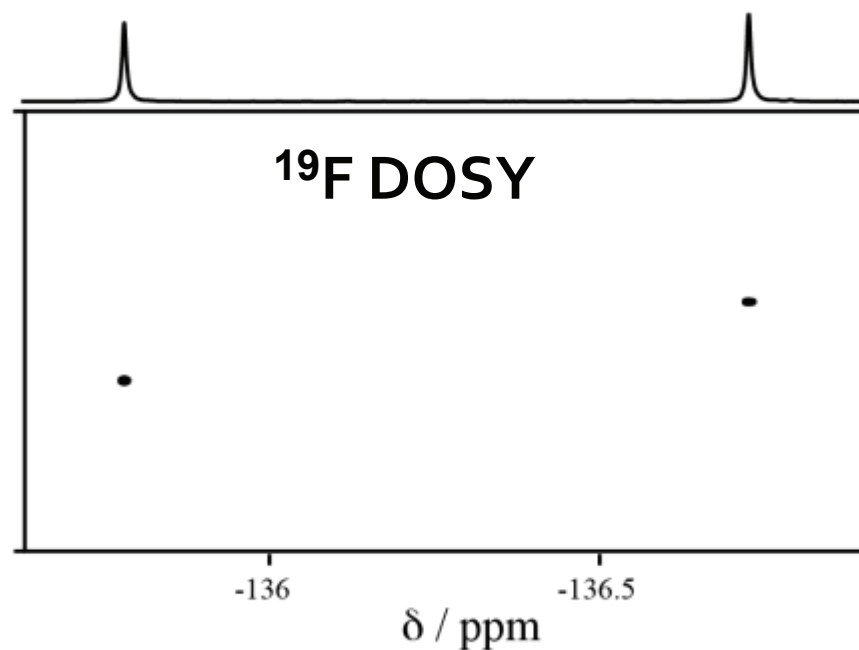
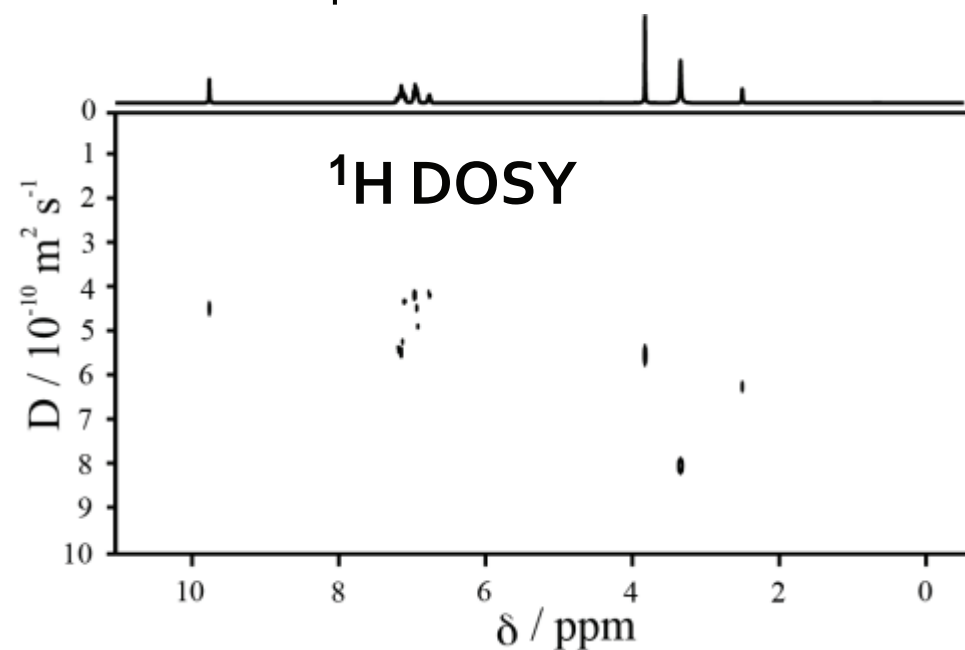
# overlap in 2D DOSY: $^{19}\text{F}$ DOSY

600 MHz  $^1\text{H}$  and  $^{19}\text{F}$  DOSY spectra of fluorinated compounds in  $\text{DMSO-d}_6$



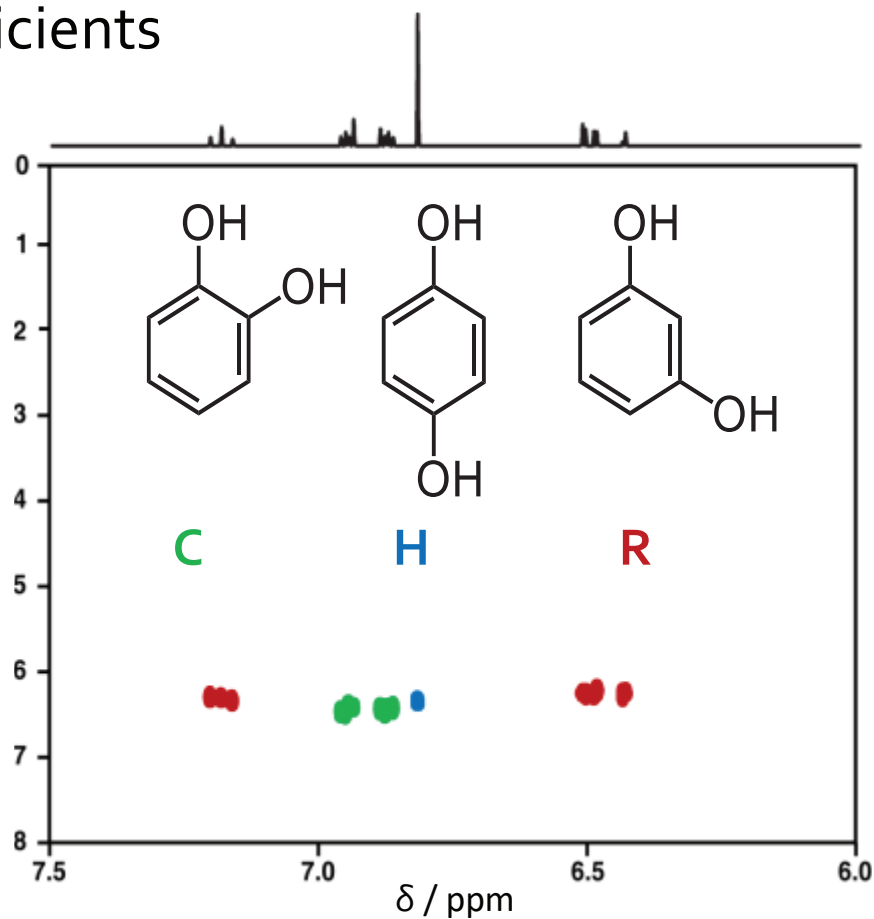
2-fluorophenol    2-fluoroanisole

the simple and well dispersed  $^{19}\text{F}$  spectrum allows unambiguous assignment



# Matrix-Assisted DOSY (MAD)

DOSY can only separate signals from species that have different diffusion coefficients



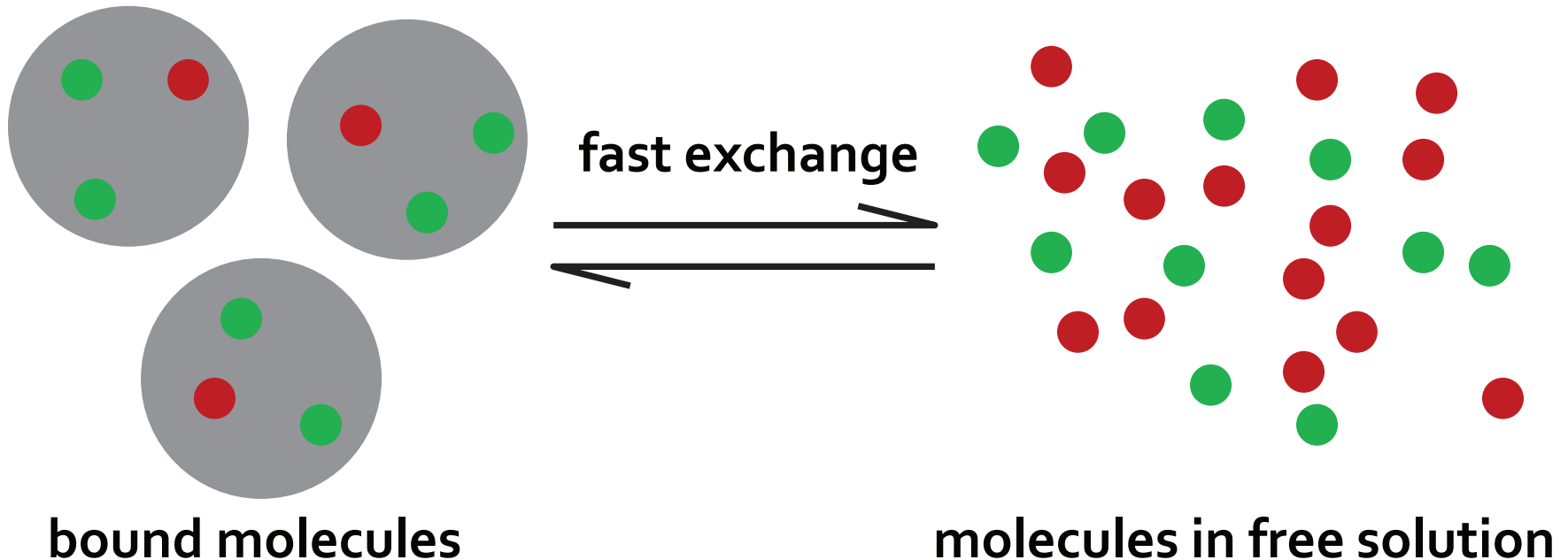
Can we manipulate the way different species diffuse?

# Interaction with a matrix

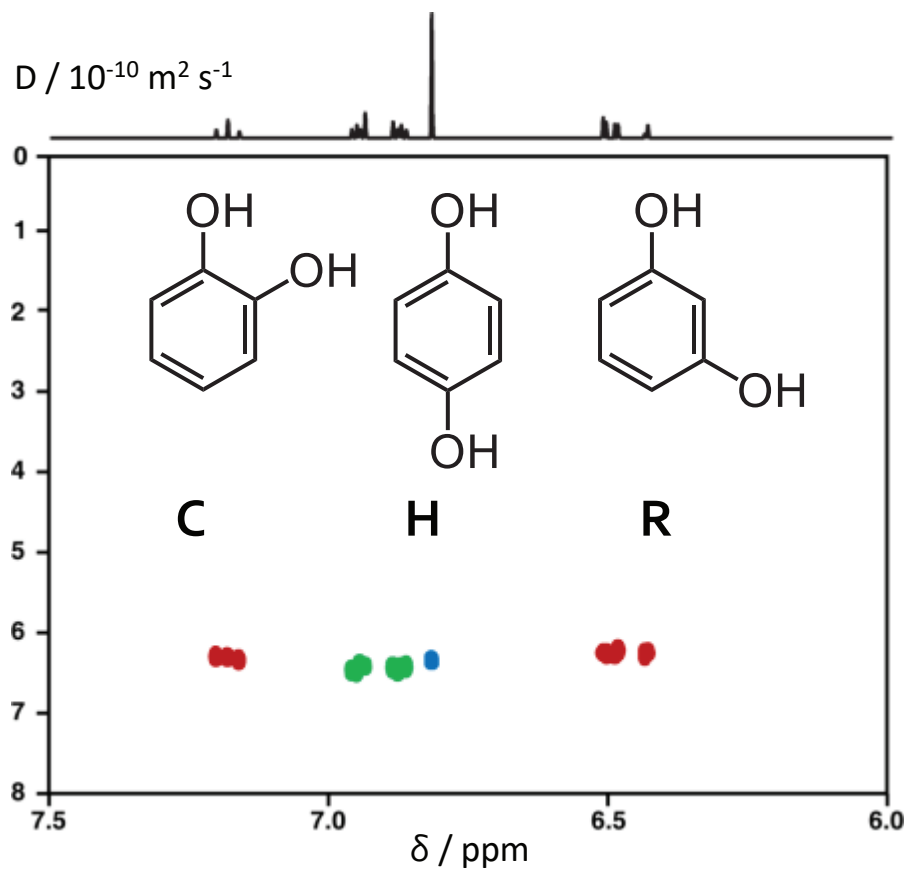
interaction of a solute with a [more slowly diffusing] matrix reduces its apparent diffusion in proportion to the strength of interaction.

**a simple two-site model**

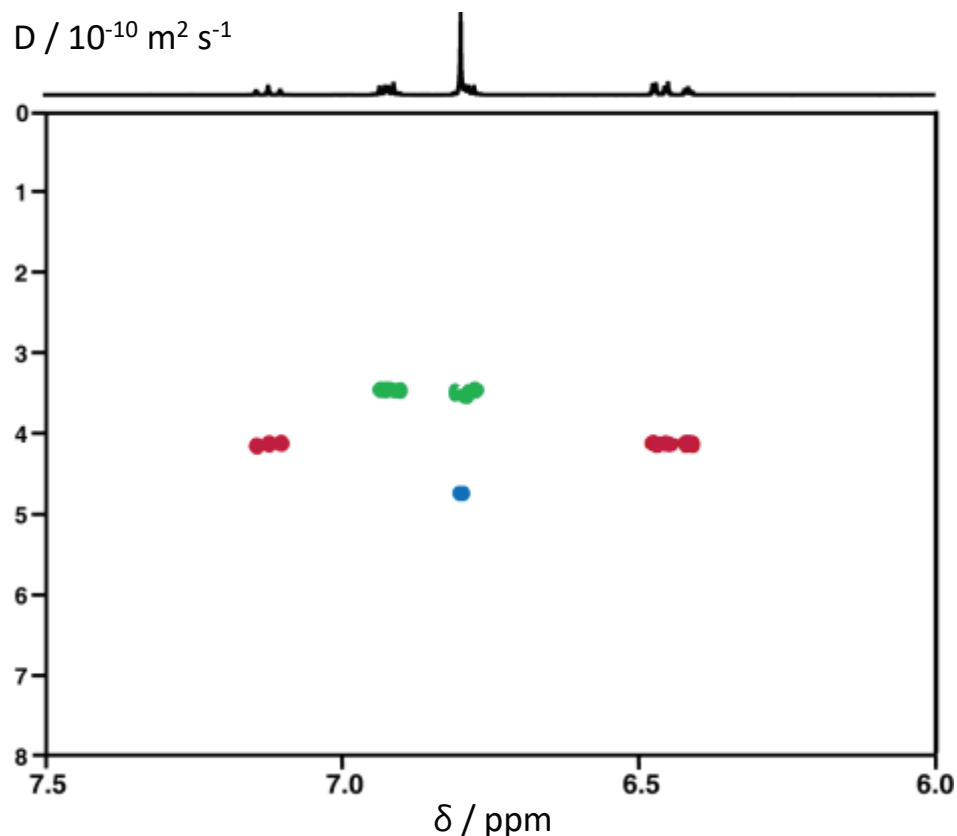
$$D_{\text{apparent}} = f_{\text{bound}} D_{\text{matrix}} + (1 - f_{\text{bound}}) D_{\text{free}}$$



# isomers resolved using micelles



$\text{D}_2\text{O}$

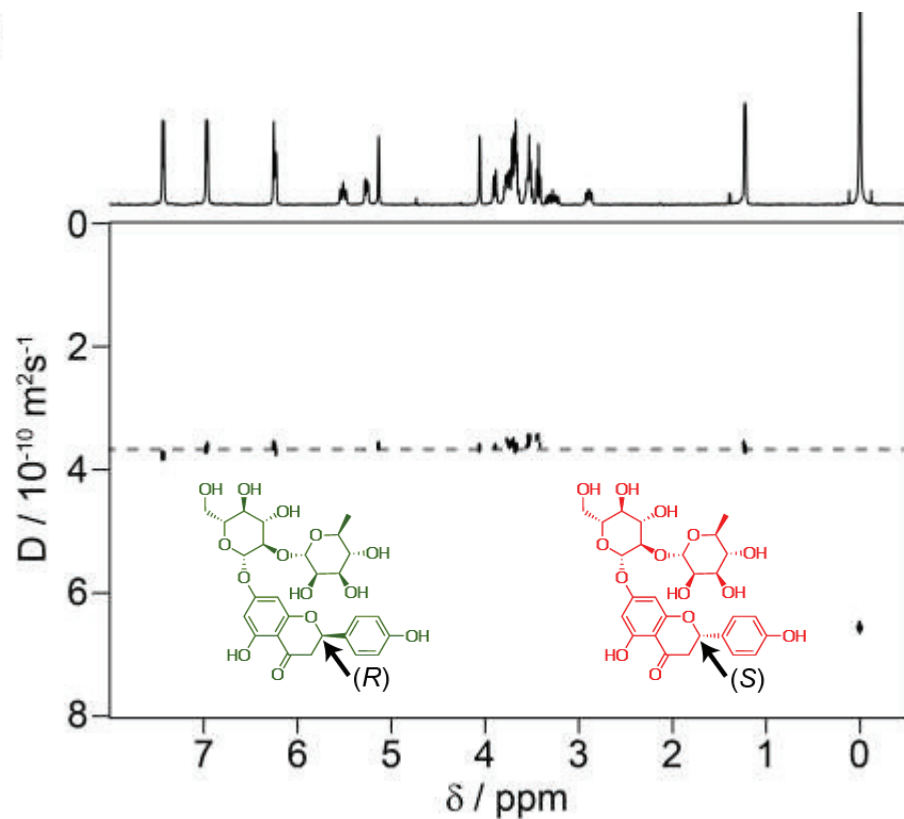


$\text{D}_2\text{O}$  with 150 mM SDS

using a micellar matrix exploits differences in binding to separate the signals of species with similar or identical diffusion coefficients

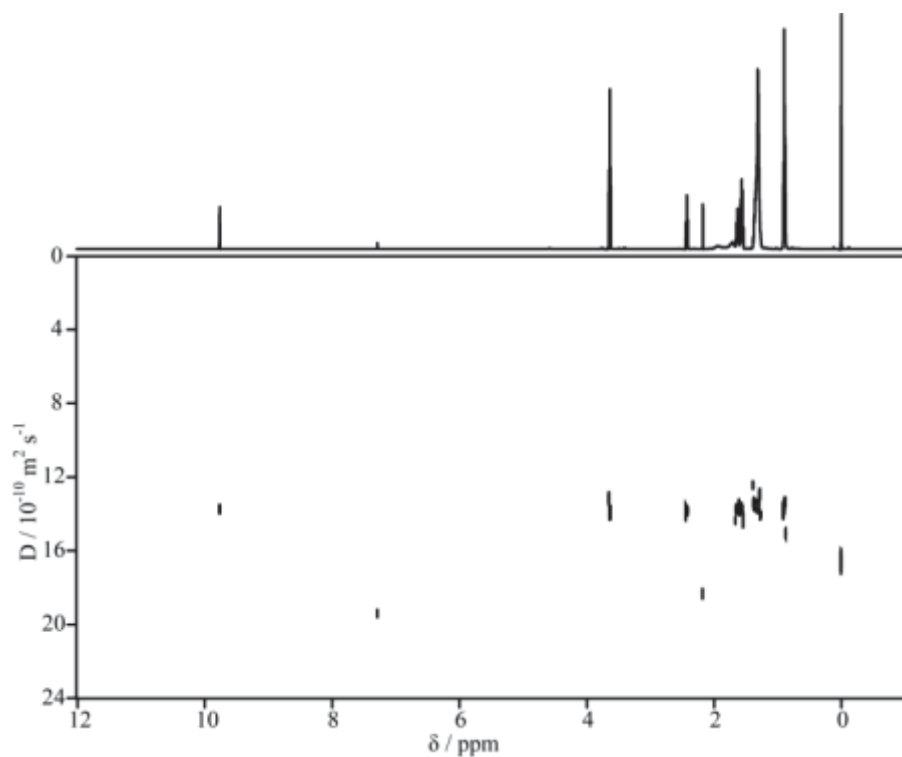


# Chiral MAD: epimers resolved using cyclodextrins

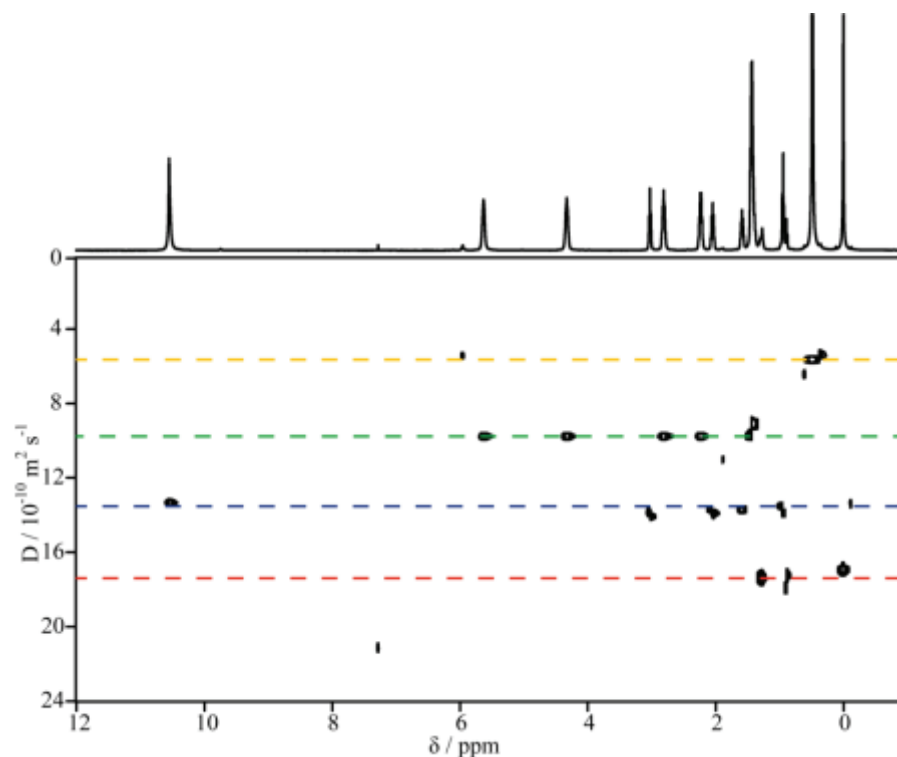


differential inclusion by  $\beta$ -cyclodextrin of the epimers of the natural product naringin is exploited, allowing separation of the naringin epimer signals by high resolution DOSY.

# Lanthanide shift reagents



an “impossible” mixture of hexane, hexanol and hexanal.



adding **Eu(fod)<sub>3</sub>** resolves the signals in both dimensions. The signals from **hexane**, **hexanal** and **hexanol** can now be identified

# Thank you

You can discover more at

- <http://www.jeol.co.jp/en/>  
(Products -> NMR)
  - Description of our products
  - Free processing software
  - Free natural products database
  - Application notes
  - Events
  - And more

- <http://nmrsupport.jeol.com/> (license)

The screenshot displays the JEOL website interface. At the top, there are navigation links for PRODUCTS, APPLICATIONS NOTES, SUPPORT, and ABOUT US. The main content area is divided into several sections:

- NEWS:** A list of recent news items with dates and titles, such as "2018/08/24 Update: 100% Challenge - moving into the future with an expansion of models".
- CASE STUDY:** A section featuring three case study thumbnails with titles like "Synthetic Organic Chemistry Laboratory (Hokuyama Lab), Department of Chemistry, School of Science, University of Tokyo".
- PRODUCT LINEUP:** A row of product images with labels: "600 NMR spectrometer FT 600", "500 NMR spectrometer FT 500", "Data NMR Software", "Four and Half Magnet", and "Magnet".
- Resource Grid:** A 2x4 grid of resource cards, including "NMR data processing software", "NMR peripherals/consumables", "User stories", "NMR basic knowledge/history", "CH-NMR-ND C", "Liquid/solid state NMR probes", "NMR application note", and "quantitative NMR".